Learning Chromatin States from ChIP-seq data Luca Pinello

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Outline

• Chromatin structure, histone modifications and combinatorial patterns

• How to segment the genome in chromatin states

• How to use ChromHMM step by step

• Further references

Epigenetics and chromatin structure

• All (almost) the cells of our body share the same genome but have very different gene expression programs....



The code over the code

- The chromatin structure and the accessibility are mainly controlled by:
- 1. Nucleosome positioning,
- 2. DNA methylation,
- Histone modifications.



Histone Modifications

- Specific histone modifications or combinations of modifications confer unique biological functions to the region of the genome associated with them:
 - H3K4me3: promoters, gene activation
- H3K27me3: promoters, poised enhancers, gene silencing
- H2AZ: promoters
- H3K4me1: enhancers
- H3K36me3: transcribed regions
- H3K9me3: gene silencing
- H3k27ac: active enhancers



ChIP-seq to measure histone data

Measuring the regulome (e.g., protein-binding of the genome)



Adapted from Dewey lecture and Peter Park Nature Genetics Review

We can "call peaks" but...

chr6:30,614,231-31,337,674



Idea: We need a way to summarize the combinatorial patterns of multiple histone marks

ChromHMM

"ChromHMM is a Java program for the learning and analysis chromatin states using a multivariate Hidden Markov Model that explicitly models the observed combination of marks"

http://compbio.mit.edu/ChromHMM/

ARTICLE

doi:10.1038/nature09906

Mapping and analysis of chromatin state dynamics in nine human cell types

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ChromHMM and Chromatin States

 Chromatin states are defined based on different combinations of histone modifications and correspond to different functional regions



Chromatin Mark Observation Frequency (%)

• The goal is to segment the genome into biologically meaningful units.

ChromHMM and segmentation



ChromHMM in practice: gather the ingredients

- Required:
 - 1. Java virtual machine (<u>http://java.com/</u>)
 - 2. ChromHMM software (<u>http://compbio.mit.edu/ChromHMM/ChromHMM.zip</u>)
 - Aligned ChIP-seq files for different histone modifications for example from the ENCODE portal (<u>https://www.encodeproject.org/</u>)
- Optionally, if we want to use it on your data:
 - 1. Raw or aligned reads for different histone modifications
 - A fast aligner aligner like Bowtie (<u>http://bowtie-bio.sourceforge.net/bowtie2</u>) or BWA (<u>http://bio-bwa.sourceforge.net/</u>)
 - 3. Bedtools (<u>https://github.com/arq5x/bedtools2</u>)

The Workflow

- 1. Get ChIP-seq raw reads for different histone modifications
- 2. Align the reads to a reference genome
- 3. Convert aligned reads in bed format
- 4. Create Binned and Binarized Tracks
- 5. Train the model
- 6. Infer the states
- 7. Interpretation

Align the reads

• Starting from a file containing raw reads (usually a fastq file) you need to align them to a reference genome to get a .bam file (binary aligned file). You can use Bowtie or BWA (links in slide #11).

 Or you can download many aligned samples from the encode portal <u>https://www.encodeproject.org/</u>

Convert aligned reads to bed format

ChromHMM needs the aligned reads in .bed format

bedtools bamtobed -i cell1_mark1.bam > ~/
data/cell1_mark1.bed

Create Binned and Binarized Tracks

• ChromHMM quantify the presence or absence of each mark in bins of fixed size



Genomic sequence

Create Binned and Binarized Tracks

 java —mx4000M —jar ChromHMM.jar BinarizeBed —b 200 CHROMSIZES/hg18 ~/ data/ cellmarkfiletable.txt SAMPLEDATA_HG18

• Inside the cellmarkfiletable.txt:

cell1 mark1 cell1_mark1.bed cell1_control.bed cell1 mark2 cell1_mark2.bed cell1_control.bed cell2 mark1 cell2_mark1.bed cell2_control.bed cell2 mark2 cell2_mark2.bed cell2_control.bed

Train the model and segment the genome



java -mx1600M -jar ChromHMM.jar LearnModel SAMPLEDATA_HG18 OUTPUTSAMPLE 10 hg18

Output of ChromHMM

- ChromHMM generates a nice HTML report called webpage_N.html (N is the number of states used) with many useful information :
 - 1. Model learned: transition and emission parameters
 - 2. Enriched functional categories
 - 3. Bed files to visualize the segmentation

Transition and emission Parameters





State To (Emission order)

Enriched functional category

Fold Enrichment GM12878_10



Fold Enrichment GM12878_10 RefSeqTSS



Position

Visualize the segmentation

- Genome Browser: https://genome.ucsc.edu/
- IGV: https://www.broadinstitute.org/igv/

	chr1 p36.22 p35.3 p34.1 p32.1 p31.1 p22.1 p13.3 q11 q21.1 q23.3 q25.2 q31.2 q32.2 q42.11 q43
	Image: wide wide wide wide wide wide wide wide
HEPG2	13_Heterochrom/lo 6_Weak_Enhancer 13_Heterochrom/lo 8_Insulator 12_Repressed 11_Weak_Ixn
HSMM	13_Heterochrom/lo 13_Heterochrom/lo 13_Heterochrom/lo 12_Repressed 11_Weak_Txn
HUVEC	eterochrom/lo 8_Insulator 7_Weak_Enhancer 11_Weak_Txn 4_Strong_Enhancer 11_Weak_Txn 8_Insulato
H1 hESC	13_Heterochrom/lo 11_Weak_Txn 13_Heterochrom/lo 12_Repressed 12_Repressed 10_Txn_Elongatio
NHLF	13_Heterochrom/lo 8_Insulator 8_Insulator 12_Repressed 8_Insulator 8_Insulato
HMEC	13_Heterochrom/lo 13_Heterochrom/lo 7_Weak_Enhancer 12_Repressed 11_Weak_Txn 8_Insulato
NHEK	13_Heterochrom/lo 11_Weak_Ixn 12_Repressed 12_Repressed 13_Heterochrom/lo 8_Insulato
GM12878	13_Heterochrom/lo 13_Heterochrom/lo 12_Repressed 3_Poised_Promoter 10_Txn_Elongation
K562	eterochrom/lo 8_Insulator 4_Strong_Enhancer 9_Txn_Transition 1_Active_Promoter 11_Weak_Txn 10_Txn_Elo
RefSeq genes	

Further References

• Other models are available to segment the genome in chromatin states:

- 1. Segway: <u>http://pmgenomics.ca/hoffmanlab/proj/segway/</u>
- 2. Spectacle: <u>https://github.com/jiminsong/Spectacle</u>
- 3. DI-HMM (soon available) GC Yuan/M Kellis

Questions?

