

A Milk-Oriented Microbiota (MOM) in Infants—How Babies Find their MOMs

Insights into next generation prebiotics & probiotics

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1906

The probiotic concept

THE PROLONGATION
OF LIFE
OPTIMISTIC STUDIES

*“the Bulgarian bacillus became a rage, companies were formed, and their directors grew rich off selling these **silly bacilli**.”*

Paul de Kruif “The Microbe Hunters” 1926

Proposed fermented milks would would contain factors (microbes) that prevent putrefaction
→ they should also help prevent putrefaction in the gut



Probiotics & prebiotics



Definitions and misperceptions

Probiotics – “live microorganisms that when administered in adequate amounts **confer a health benefit on the host**” (UNFAO/WHO 2001).

Prebiotics – “a prebiotic is a selectively fermented ingredient that allows specific changes, both in the composition and/or activity in the gastrointestinal microflora, that **confer benefits upon host well-being and health**” (J. Nutr. 2007 137:830S-837S)

Synbiotics – combinations of prebiotics and probiotics

Two consumers...infants and the infant gut microbiota



Tissier H. 1905. Repartition des microbes dans l'intestin du nourisson. Ann. de l'Institut de Pasteur 19:109

“...to be constituted, by microscopic examination, of only one species, *Bacterium bifidus*, a strictly anaerobic bacterium...”

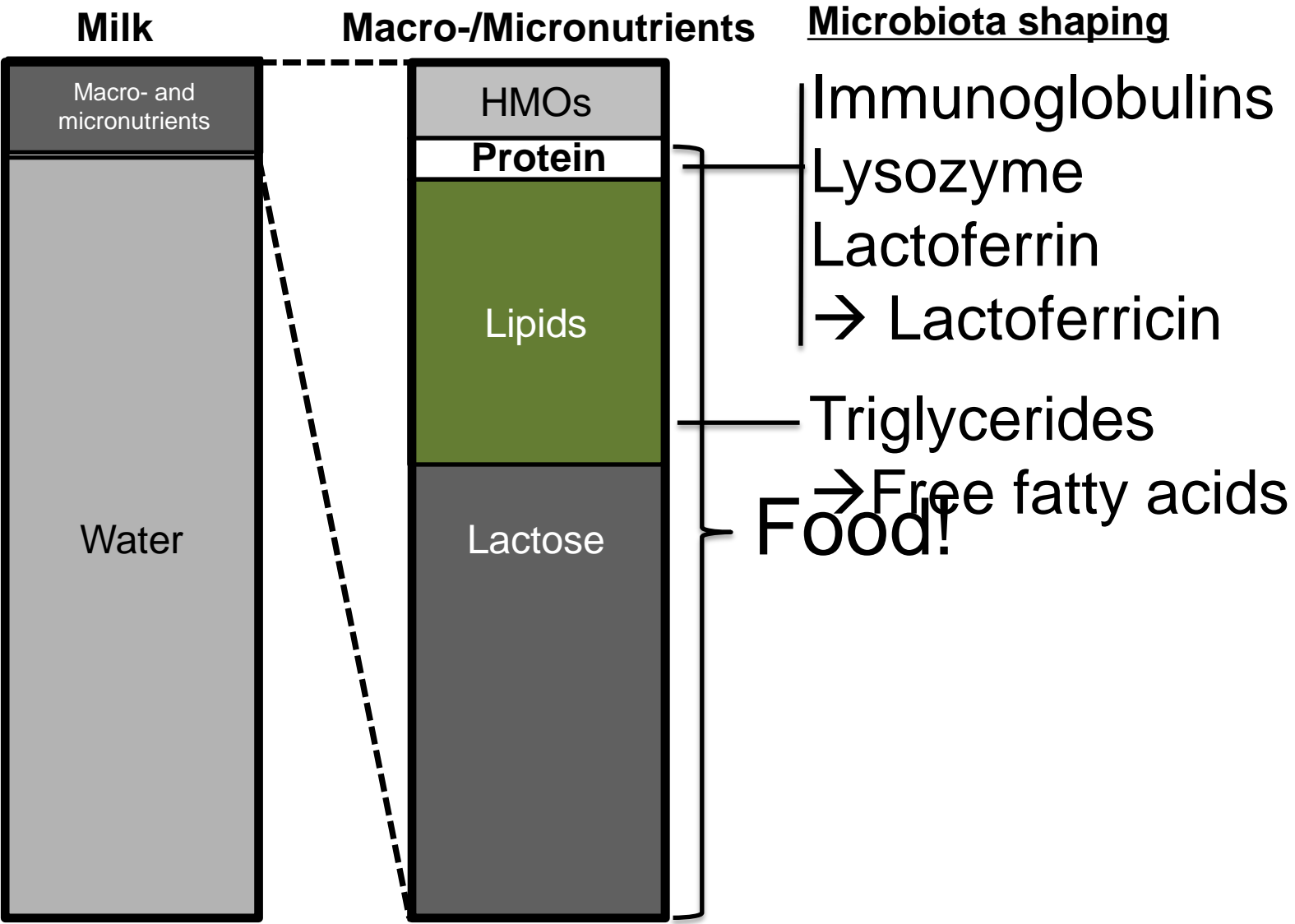
Gyorgy, P (1971) The uniqueness of human milk, biochemical aspects. AJCN 24 970.

“The bifidus factor contains in various proportion, lactose, galactose, fucose, N-acetylglucosamine and N-acetylneuraminic acid (sialic acid)”

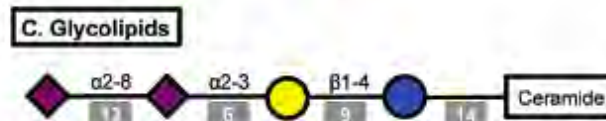
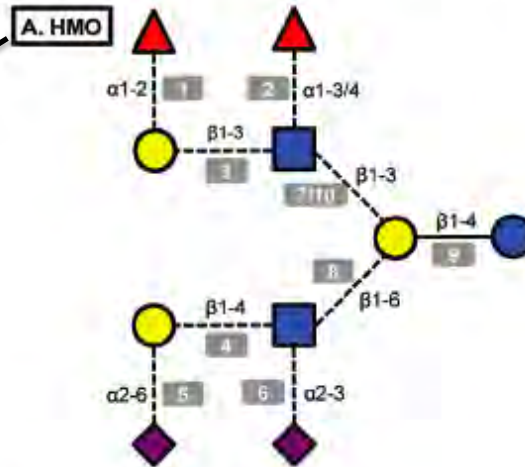
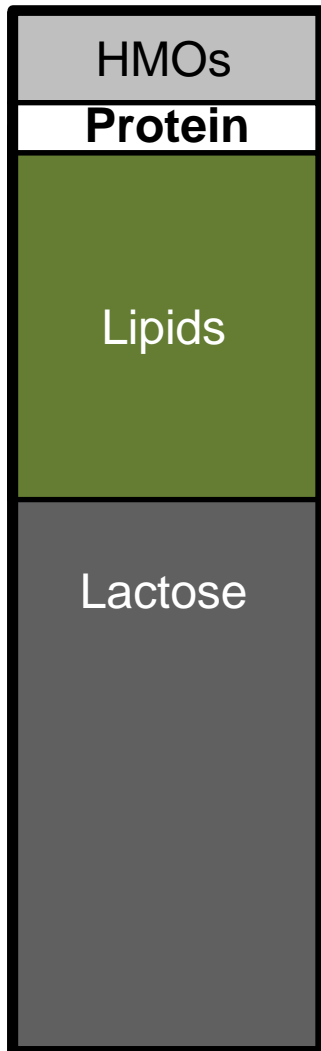
What factors in milk shape the microbiota?



Human milk composition



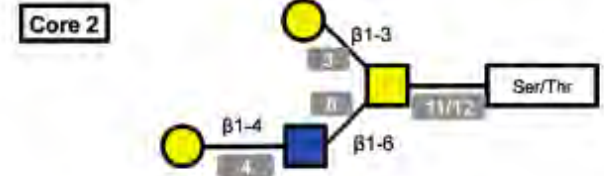
Human Milk Glycans



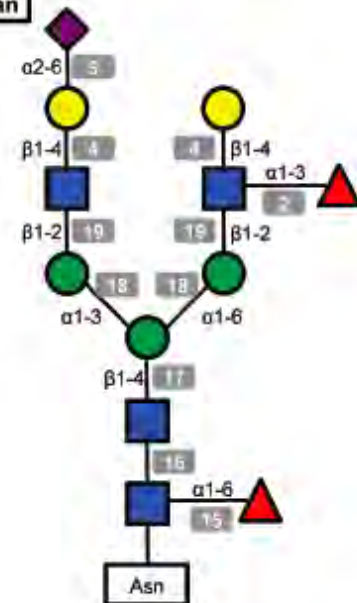
- 1 α 1-2 Fucosidase
- 2 α 1-3/4 Fucosidase
- 3 β 1-3 Galactosidase
- 4 β 1-4 Galactosidase
- 5 α 2-6 Sialidase
- 6 α 2-3 Sialidase
- 7 β 1-3 *N*-Acetylglucosaminidase
- 8 β 1-6 *N*-Acetylglucosaminidase
- 9 β 1-4 Galactosidase (lactase)
- 10 Lacto-*N*-biosidase

- 11 Endo- α -*N*-acetylgalactosaminidase (EngBF)
- 12 Endo- α -*N*-acetylgalactosaminidase (NagBb)
- 13 α 2-8 Sialidase
- 14 Endoglucosylceraminidase
- 15 α 1-6 Fucosidase
- 16 Endo- β -*N*-acetylglucosaminidase
- 17 β 1-4 Mannosidase
- 18 α -Mannosidase
- 19 β 1-2 *N*-Acetylglucosaminidase

B. O-linked glycans



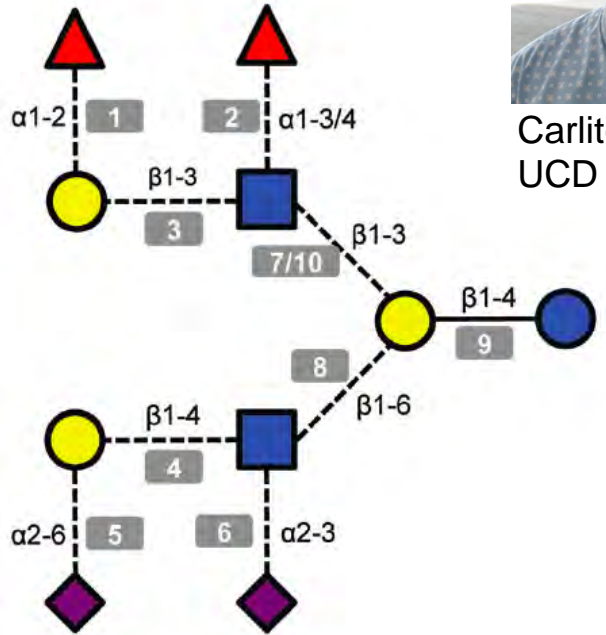
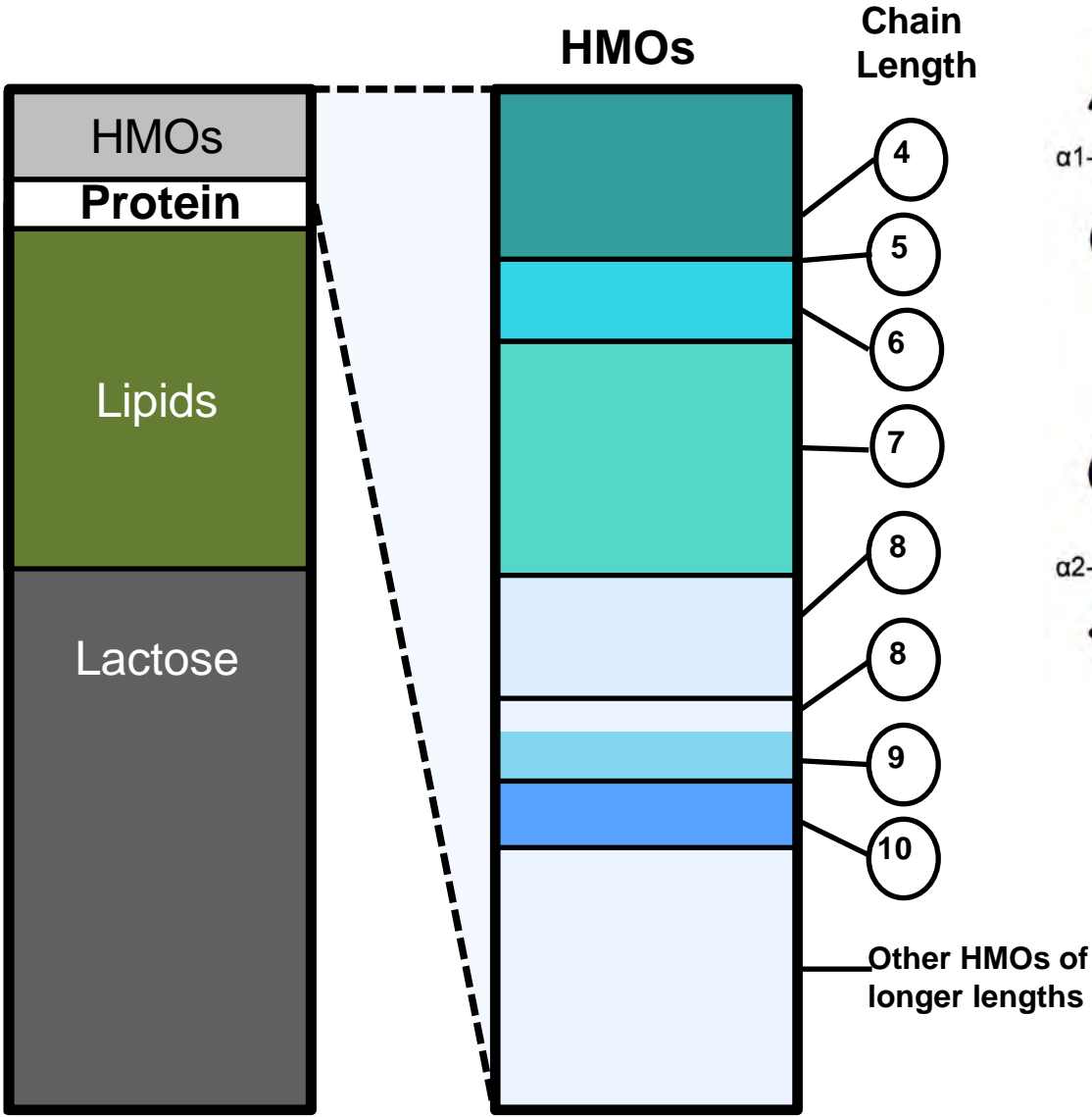
D. Complex N-glycan



Human milk oligosaccharides



Carlito Lebrilla
UCD Chemistry



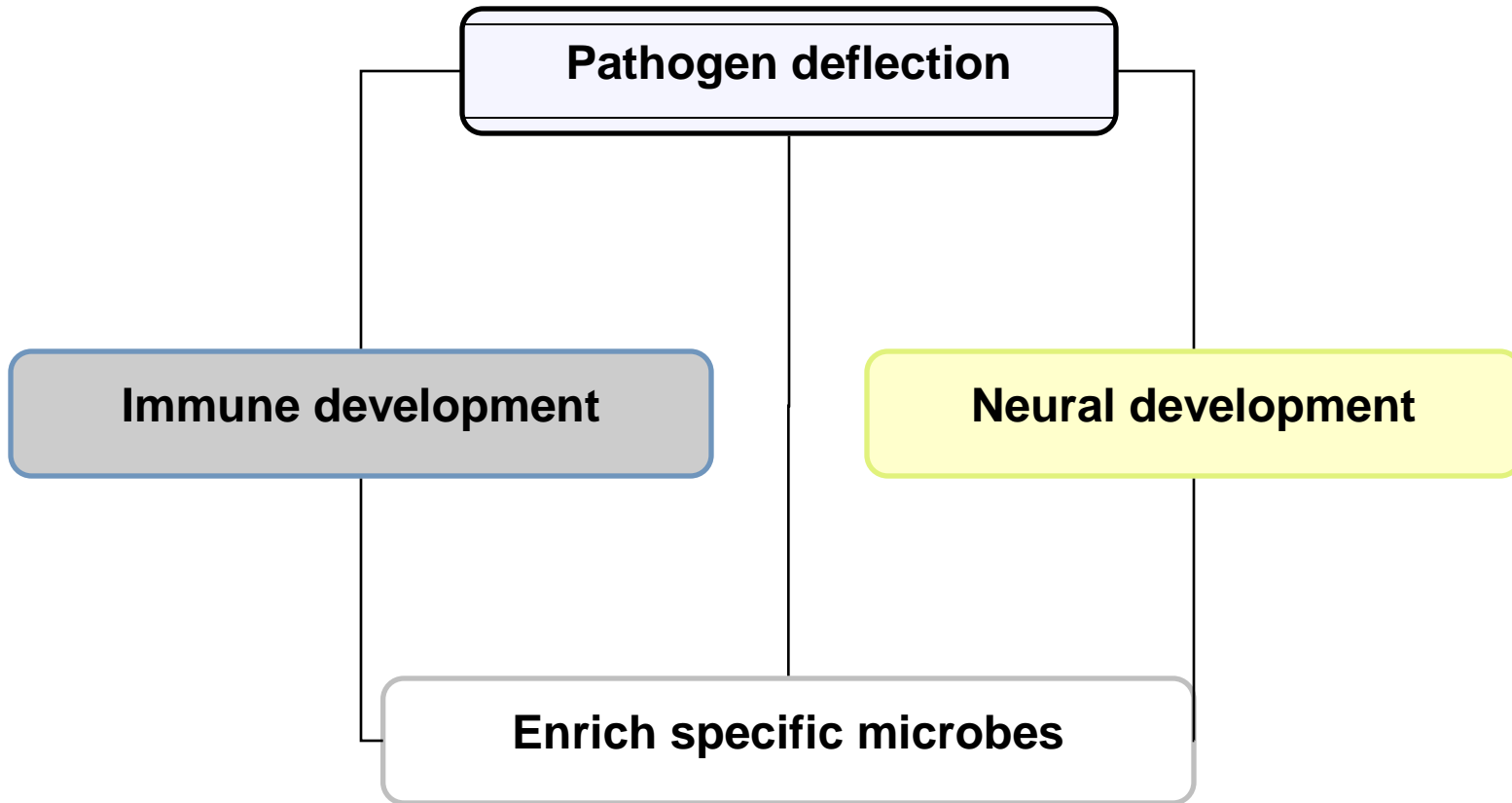
- Human indigestible and highly variable
- Higher proportion of fucosylated (40-70%) than sialyated (4-38%)
- Nearly 200 species in pooled human milk

Nature 468 S5-S7 (23 December 2010)

Garrido et al Microbiology (2013)

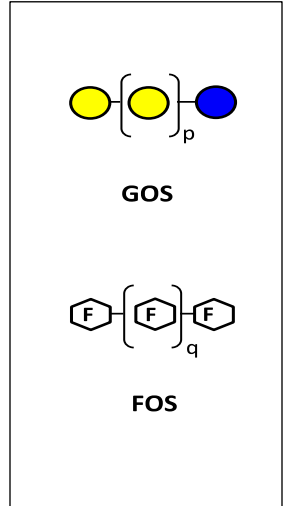
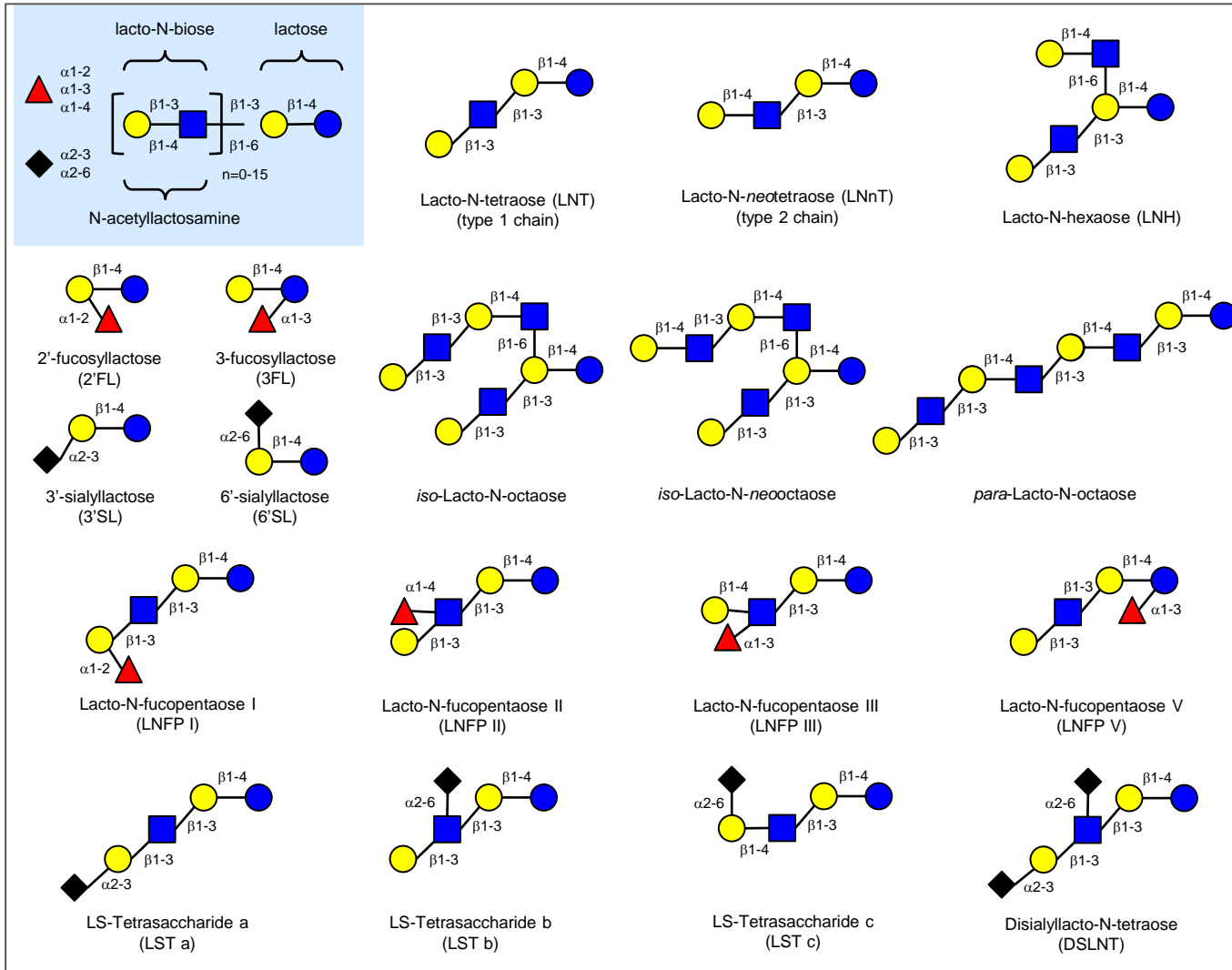
Why Make Glycoconjugates/Free Glycans?

(if they are not consumed by the infant)



HMO Structural Diversity

FOS/GOS



150 – 200 different HMO

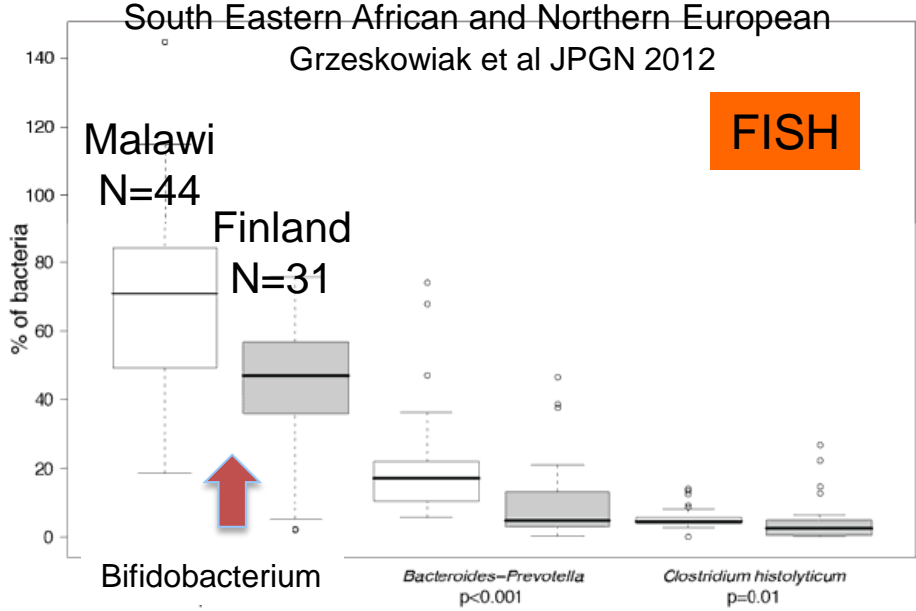
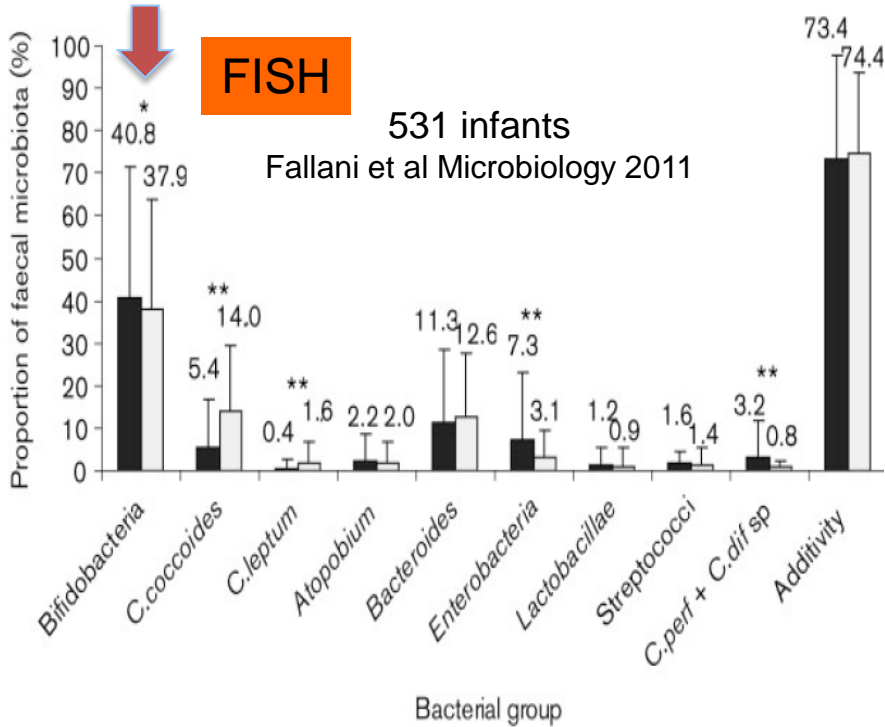
Courtesy of Lars Bode

Breast milk enriches bifidobacterial populations

Penders et al. Pediatrics 2006

TABLE 2 Median Counts and Prevalence of Selected Gut Bacteria in Feces of Infants 1 Month of Age (n = 1032)

Quantitative PCR	Bifidobacteria	<i>E coli</i>	<i>C difficile</i>	<i>B fragilis</i> Group	Lactobacilli	Total
Median counts (range), log ₁₀ CFU/g feces	10.68 (6.84–11.56)	9.35 (5.91–10.79)	5.32 (2.70–9.57)	9.28 (5.74–10.44)	8.66 (7.92–10.73)	11.12 (9.43–12.14)
Prevalence, %	98.6	87.7	25.0	81.6	32.4	100

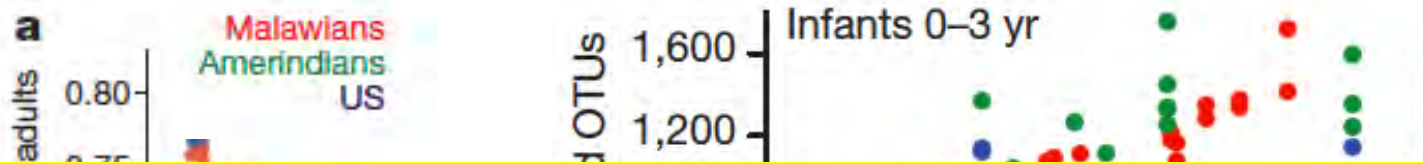


Breast milk
enriches
bifidobacterial
populations

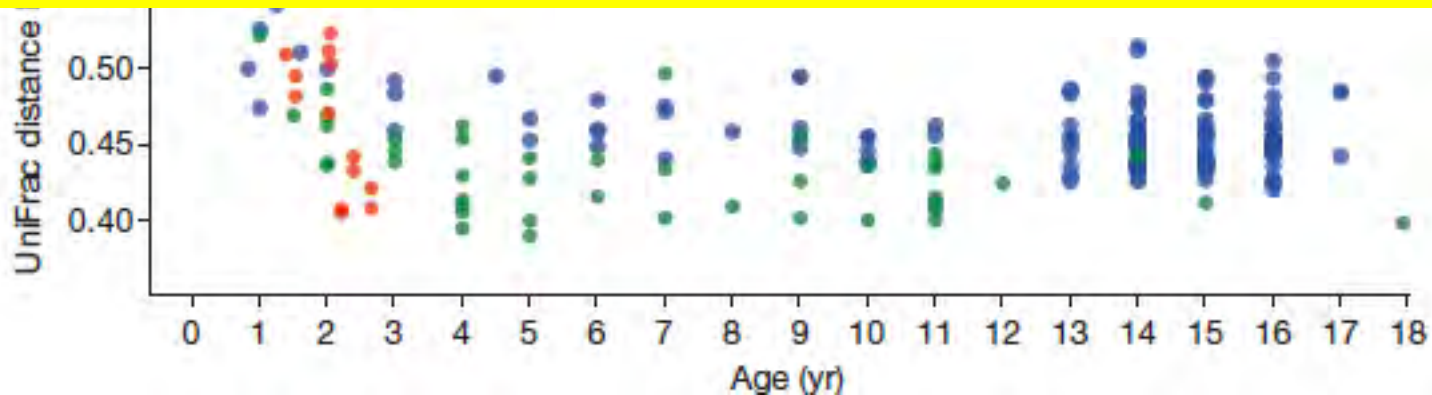
Human gut microbiome viewed across age and geography

Tanya Yatsunenکو¹, Federico E. Rey¹, Mark J. Manary^{2,3}, Indi Trehan^{2,4}, Maria Gloria Dominguez-Bello⁵, Monica Contreras⁶, Magda Magris⁷, Glida Hidalgo⁷, Robert N. Baldassano⁸, Andrey P. Anokhin⁹, Andrew C. Heath⁹, Barbara Warner², Jens Reeder¹⁰, Justin Kuczynski¹⁰, J. Gregory Caporaso¹¹, Catherine A. Lozupone¹⁰, Christian Lauber¹⁰, Jose Carlos Clemente¹⁰, Dan Knights¹⁰, Rob Knight^{10,12} & Jeffrey I. Gordon¹

N= 531 (16S) N=110 (metagenome)



“Most shotgun and 16S rRNA V4 sequences (75 ± 20%) in all babies mapped to members of the *Bifidobacterium* genus.”

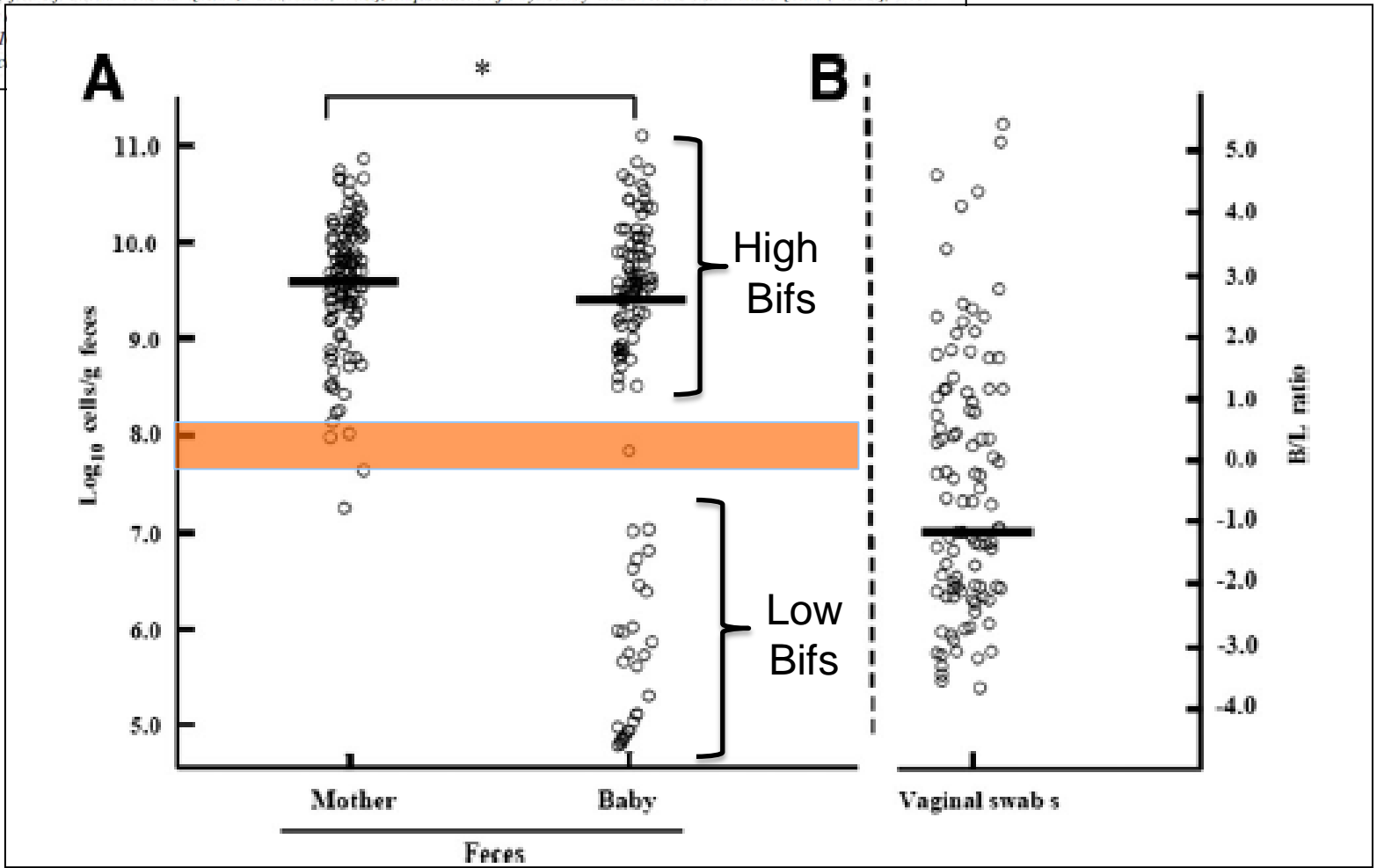


Influence of Maternal Bifidobacteria on the Establishment of Bifidobacteria Colonizing the Gut in Infants

KATSUNAKA MIKAMI, HIDENORI TAKAHASHI, MOTO KIMURA, MITSUHIRO ISOZAKI, KUNIO IZUCHI, RUMIKO SHIBATA, NOBUYUKI SUDO, HIDEO MATSUMOTO, AND YASUHIRO KOGA

Laboratory for Infectious Diseases [K.M., H.T., M.K., Y.K.], Department of Psychiatry and Behavioral Science [K.M., H.M.], and Department of Pediatrics [K.M., H.M.], Fukuoka 81-3-15, Japan

- n=110 babies
- 1 month of age
- Measured by qPCR





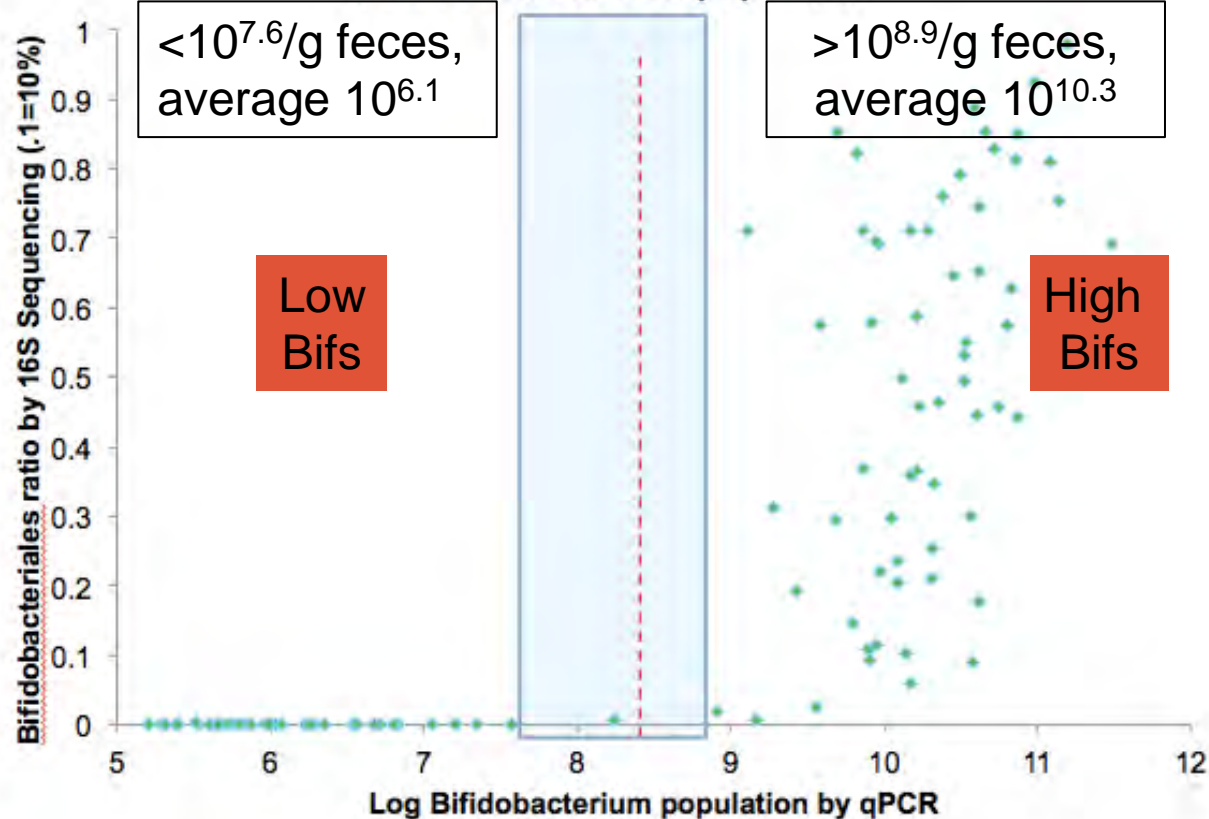
Zach Lewis

Lactation Study

Who is conducting this study?

This clinical study led by J. Bruce German, Ph.D., and colleagues at the UC Davis Foods for Health Institute, and Food Science and Technology, Viticulture and Enology, and Chemistry departments is part of the **Milk Bioactives** and the **Functional Glycobiology Program**. This

% Bif vs Absolute Bif population



48 breast fed infants
4 time points
(Day 6, 21, 71, 120)

A scanning electron micrograph (SEM) showing a dense population of bifidobacteria. The bacteria are rod-shaped and exhibit characteristic bifurcations, appearing in various orientations and colors ranging from bright yellow to dark red. The background is black, making the individual bacterial structures stand out. A white text overlay is centered in the image, and technical SEM data is visible at the bottom left.

Why bifidobacteria?

Acc.V Magn WD | 10 μ m
20.0 kV 3348x 8.9 UCD 272, 25 HMO

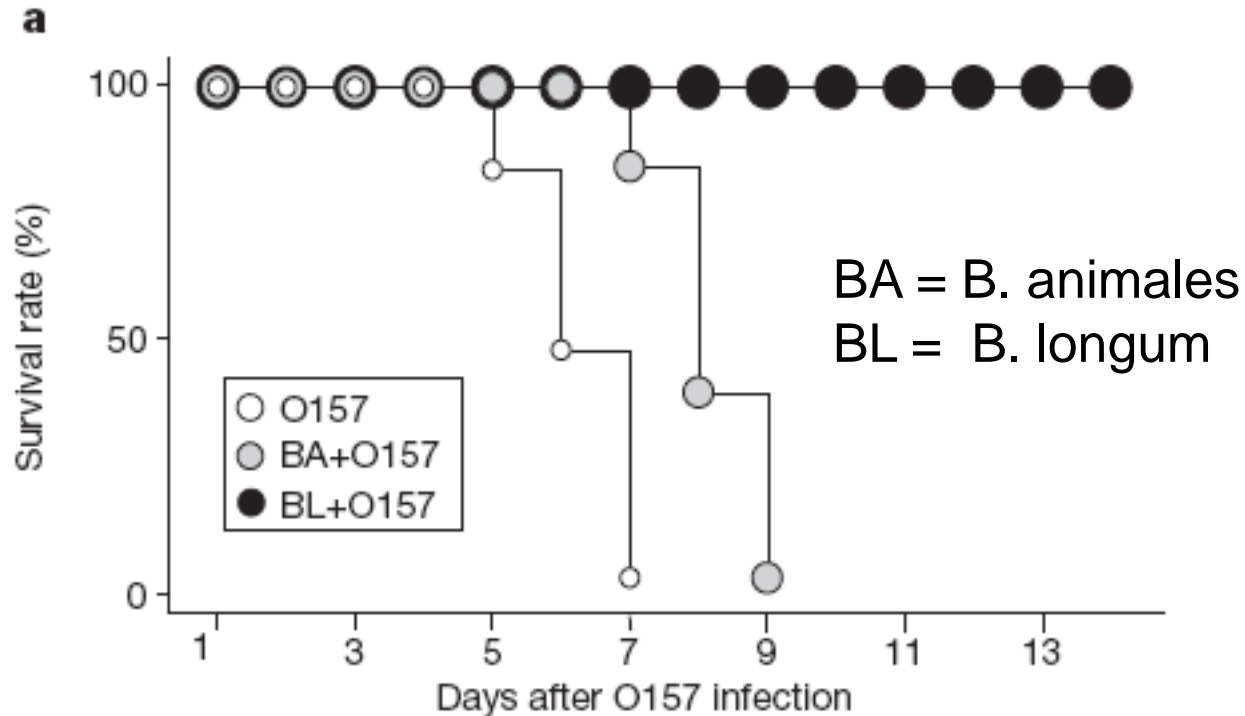
Bifidobacteria can protect from enteropathogenic infection through production of acetate

Shinji Fukuda^{1,2}, Hidehiro Toh³, Koji Hase¹, Kenshiro Oshima⁴, Yumiko Nakanishi^{1,2,5}, Kazutoshi Yoshimura⁶, Toru Tobe⁷, Julie M. Clarke⁸, David L. Topping⁸, Tohru Suzuki⁹, Todd D. Taylor³, Kikuji Itoh⁶, Jun Kikuchi^{2,5,10}, Hidetoshi Morita¹¹, Masahira Hattori⁴ & Hiroshi Ohno^{1,2,12}



Nature 2010

Mouse survival after O157 infection



Bifidobacteria can protect from enteropathogenic infection through production of acetate

Shinji Fukuda^{1,2}, Hidehiro Toh³, Koji Hase¹, Kenshiro Oshima⁴, Yumiko Nakanishi^{1,2,5}, Kazutoshi Yoshimura⁶, Toru Tobe⁷, Julie M. Clarke⁸, David L. Toppino⁸, Tohru Suzuki⁹, Todd D. Taylor³, Kikuii Itoh⁶, Jun Kikuchi^{2,5,10}, Hidetoshi Morita¹¹

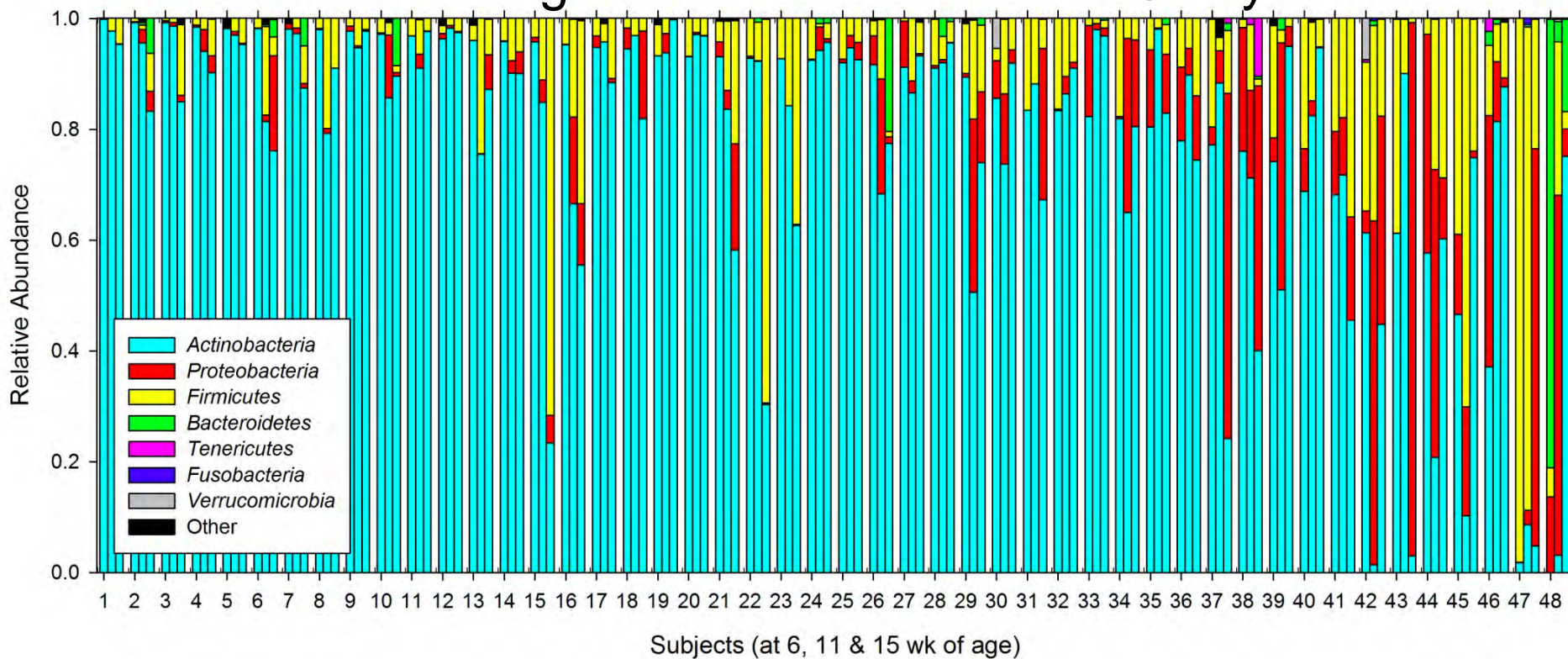
If bifidobacteria can grow well on a targeted sugar *in situ*, growth and accompanying production of acetate is protective

- ABC-type sugar transporter periplasmic component (COG1879)
- ABC-type sugar transporter permease component (COG1172)
- ABC-type sugar transporter ATPase component (COG1129)
- Transcriptional regulator



Why bifidobacteria?

Bangladesh Infant Vitamin A Study



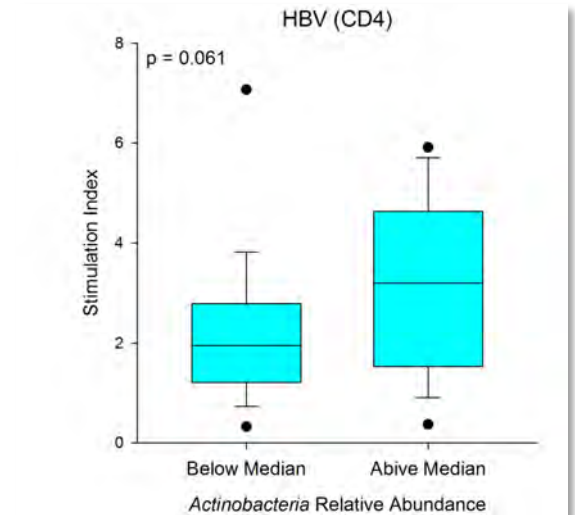
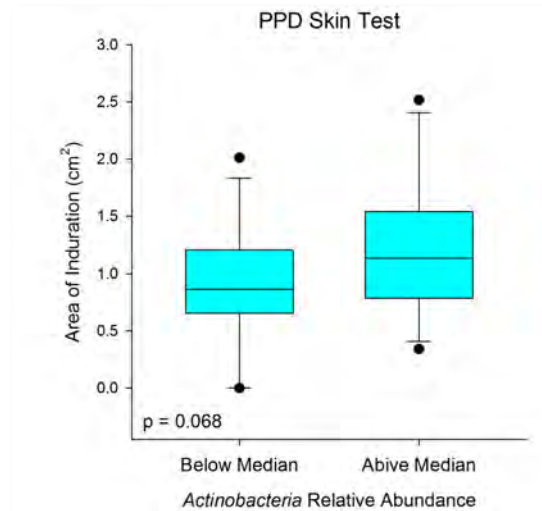
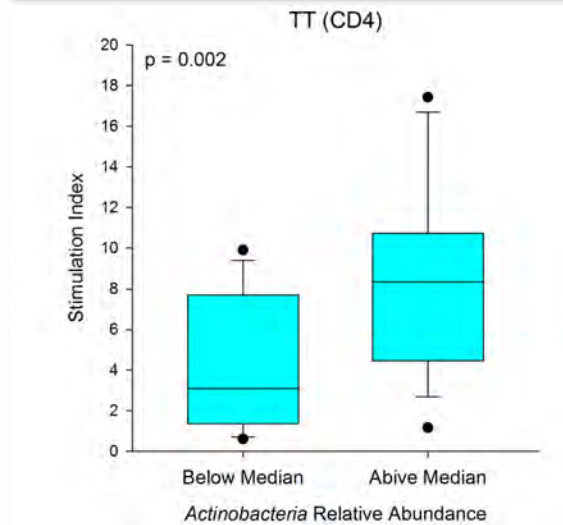
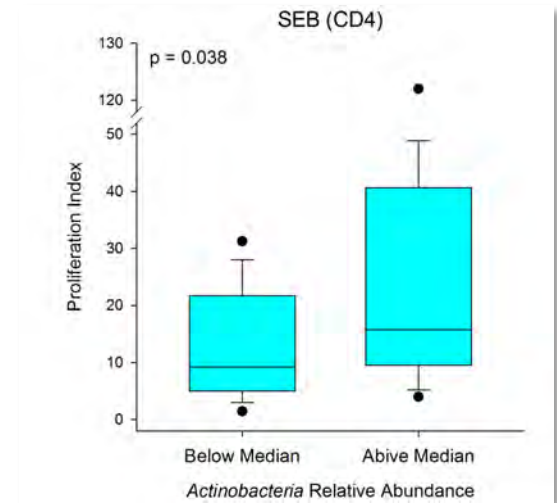
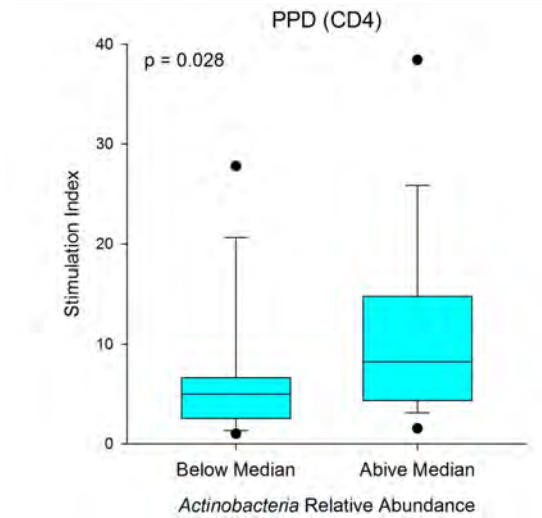
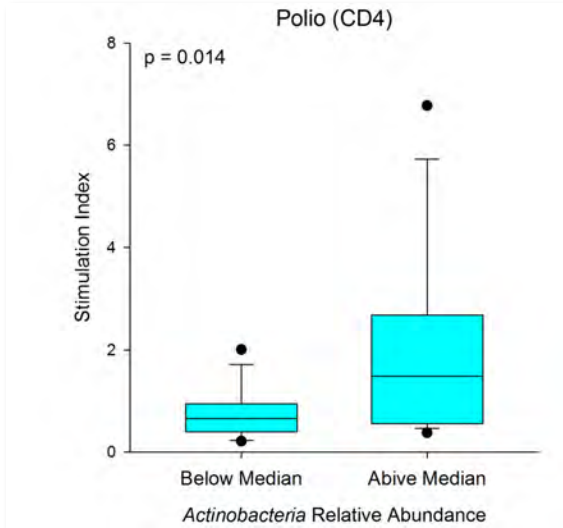
International Center for Diarrhoeal Disease
Research (Bangladesh)



Western Human Nutrition Research Center
(Davis, CA)

Huda, Stephensen et al In Prep

Positive correlation between Actinobacteria and vaccine response

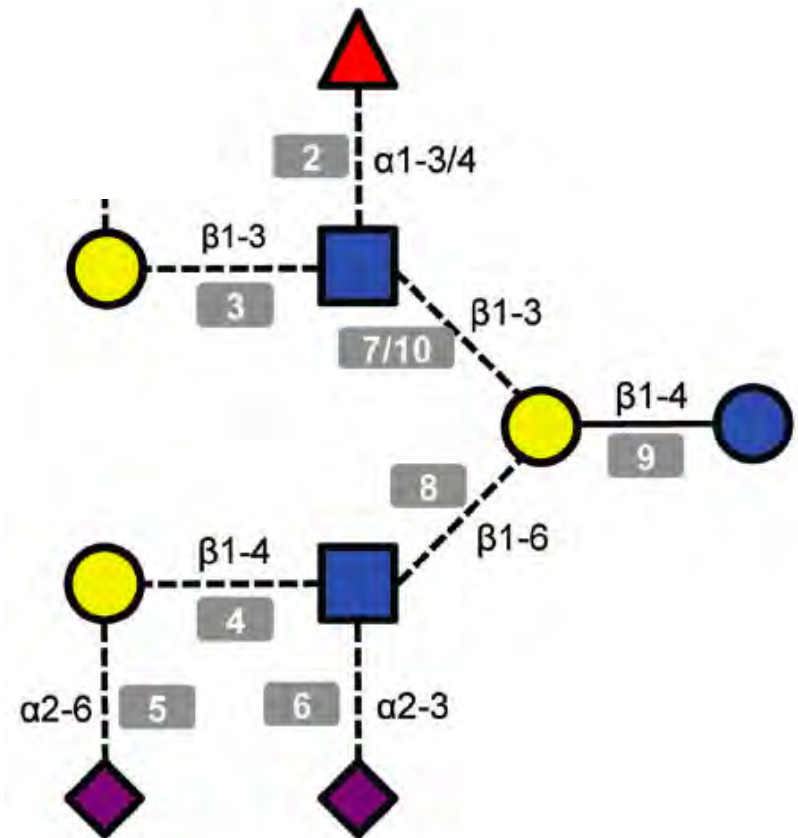


Do different mother's milk glycan types influence different microbiota populations?

Secretor vs non-secretor

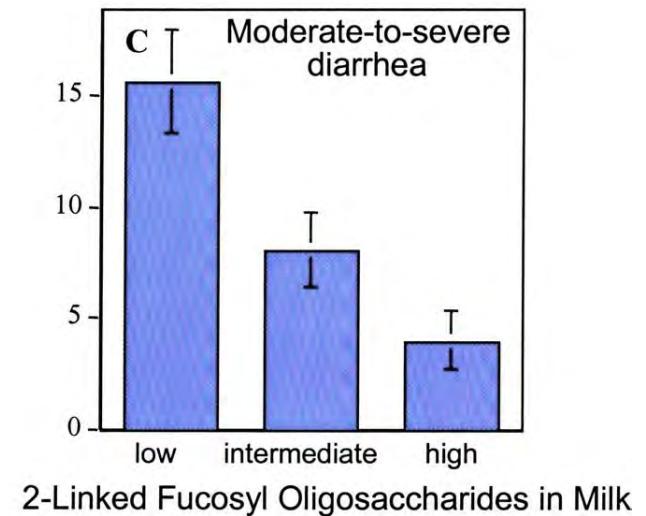
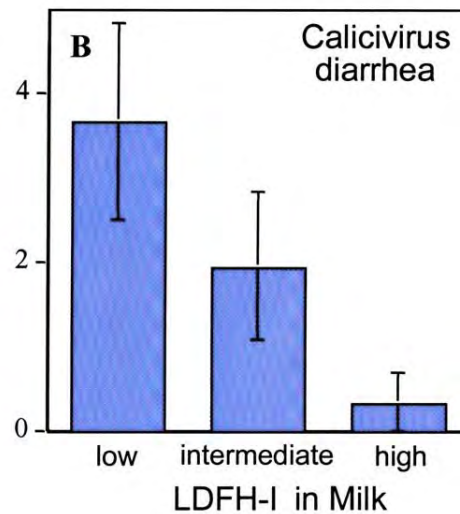
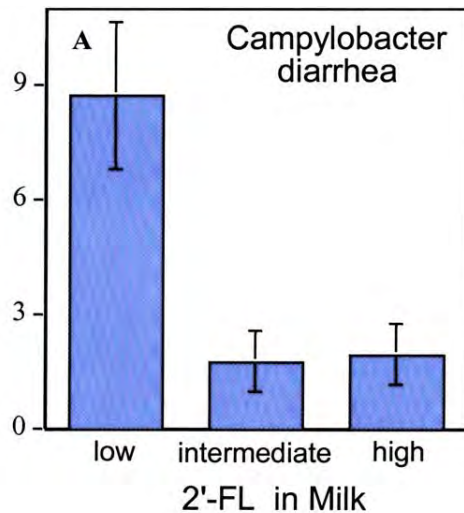
FUT2 ("Secretor" gene)

- Produces the 2' fucosylated precursor to the A, B, H, and Lewis b antigens in secretions, including breast milk
- 20% of U.S. population are non-secretors



Secretor vs. non-secretor milk protects differently

Incidence of diarrhea per 100 child-months





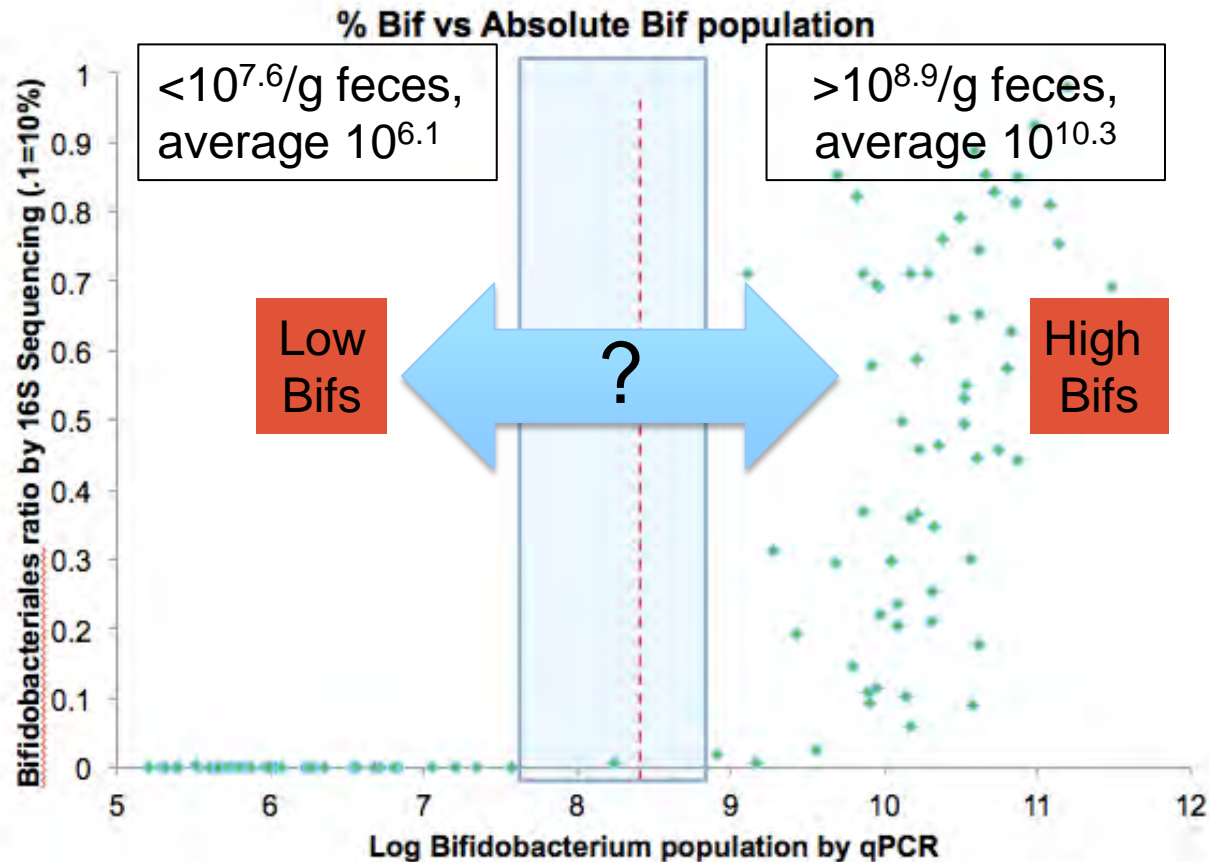
Who is conducting this study?

This clinical study led by J. Bruce German, Ph.D., and colleagues at the Health Institute, and Food Science and Technology, Viticulture and Enology departments is part of the Milk Bioactives and the Functional Components study. This study has been reviewed and approved by the UC Davis Committee on Human Subjects.



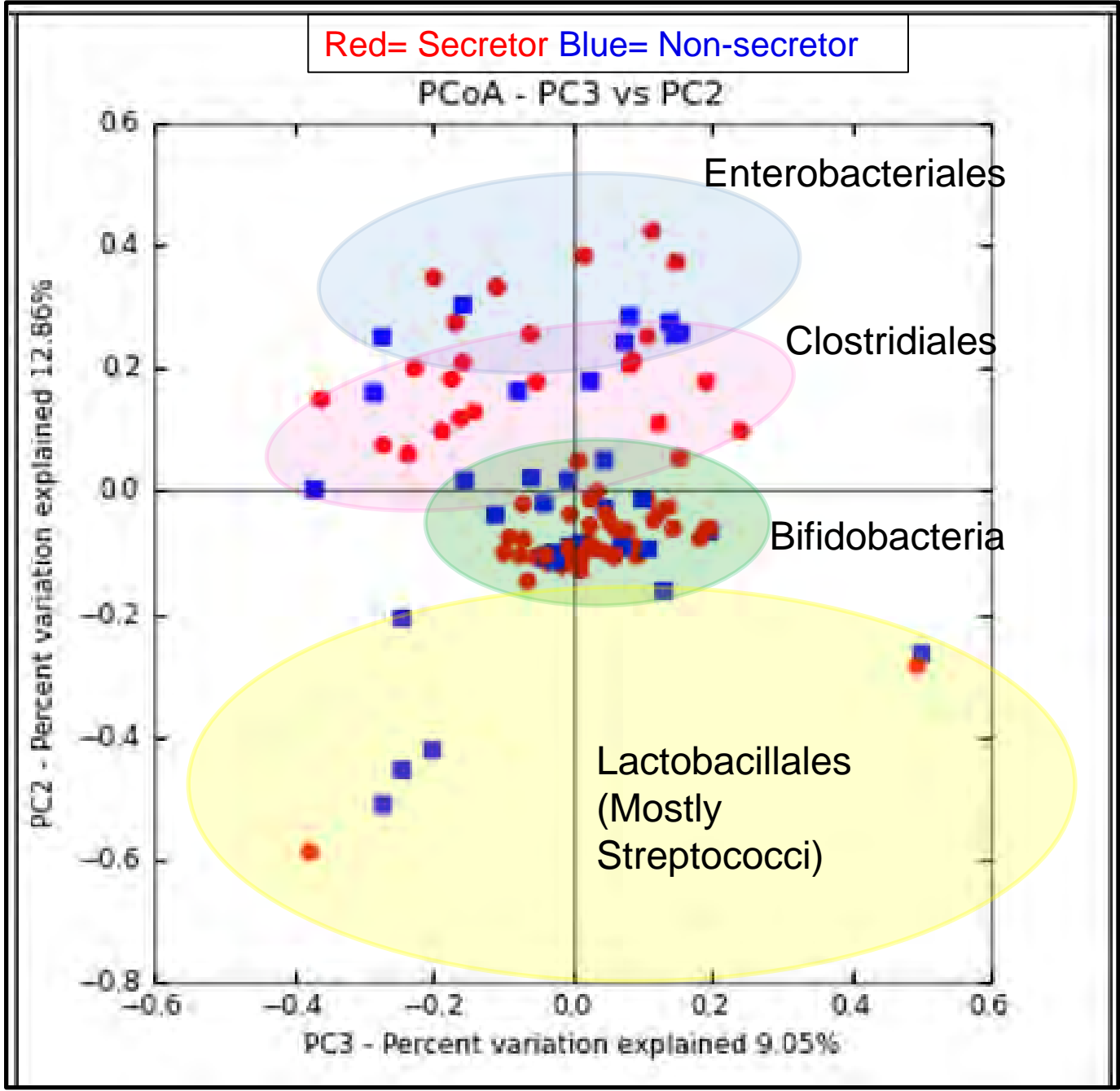
Zach Lewis

48 breast fed infants
4 time points
(Day 6, 21, 71, 120)



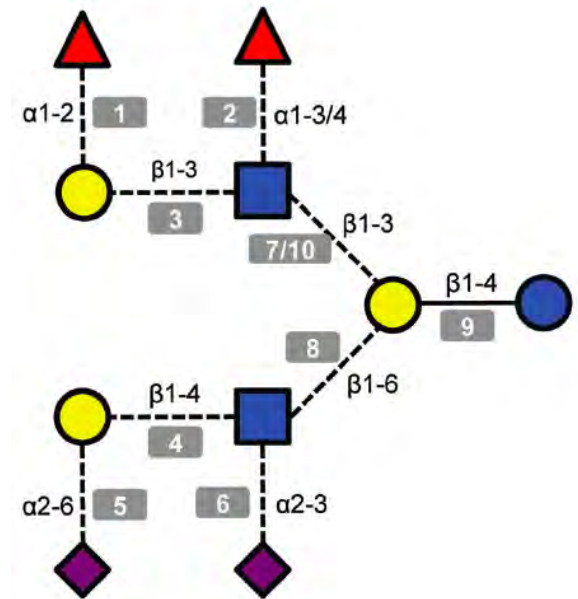
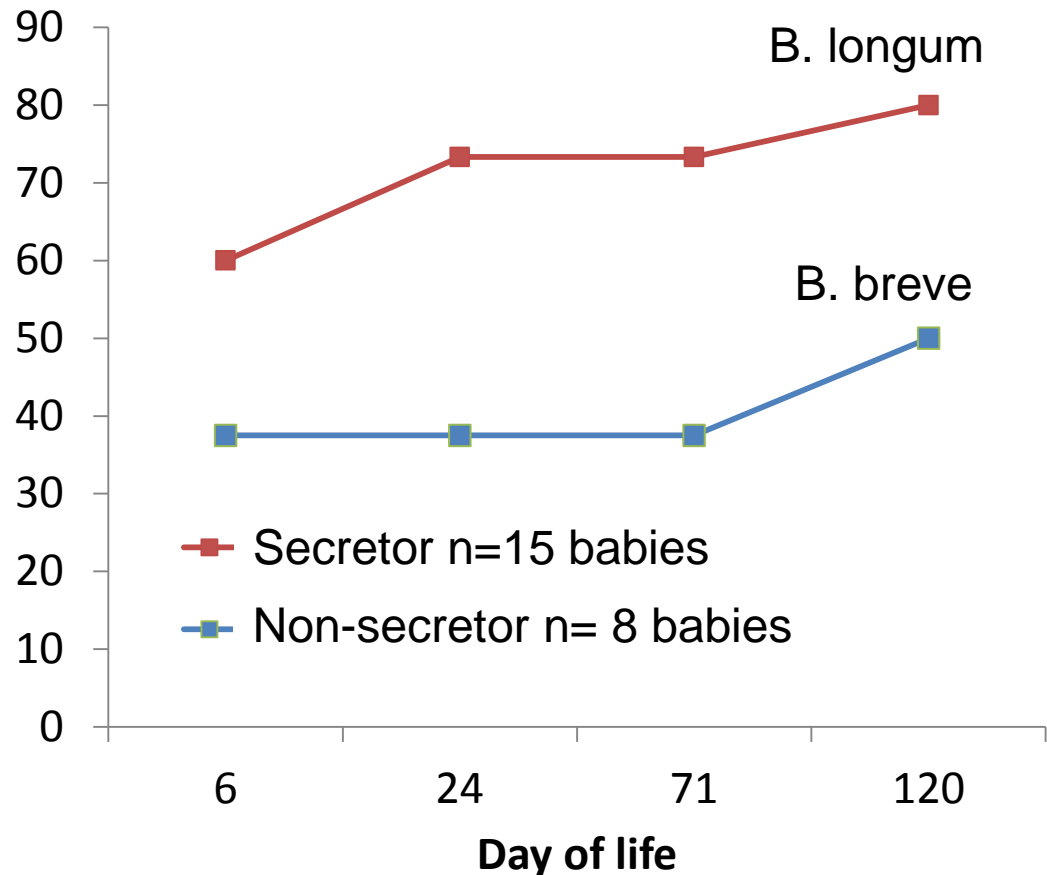


- 48 breast fed infants
- 4 time points
- (Day 6, 21, 71, 120)



Do different mothers milk glycan types influence different microbiota populations?

% Babies with bifidobacteria established ($>10^9$)



Lewis et al (in prep)

Are specific glycans consumed by bifidobacteria *in situ*?

Feces from breast fed infant

```
graph TD; A[Feces from breast fed infant] --> B[Glycoprofile]; A --> C[Microbial ecology];
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Glycoprofile

Microbial
ecology

Are oligos missing in feces in which bifidobacteria are dominant?

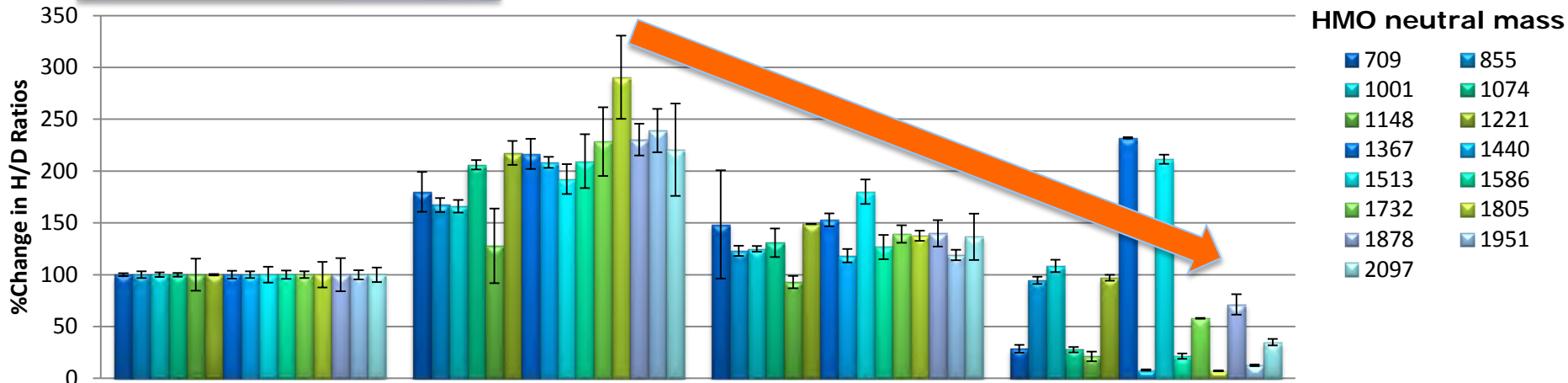
Feces Oligosaccharides of Term Infant Vary With Bacterial Population



Lorna de Leoz

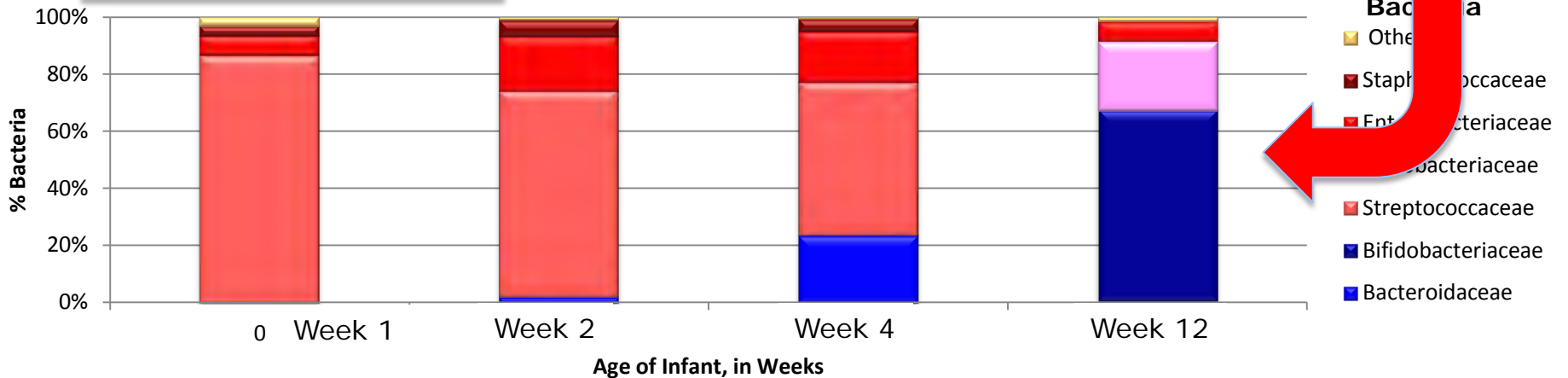
Fecal HMO Profile

FULL TERM INFANT



Fecal Bacterial Profile

B. longum/infantis

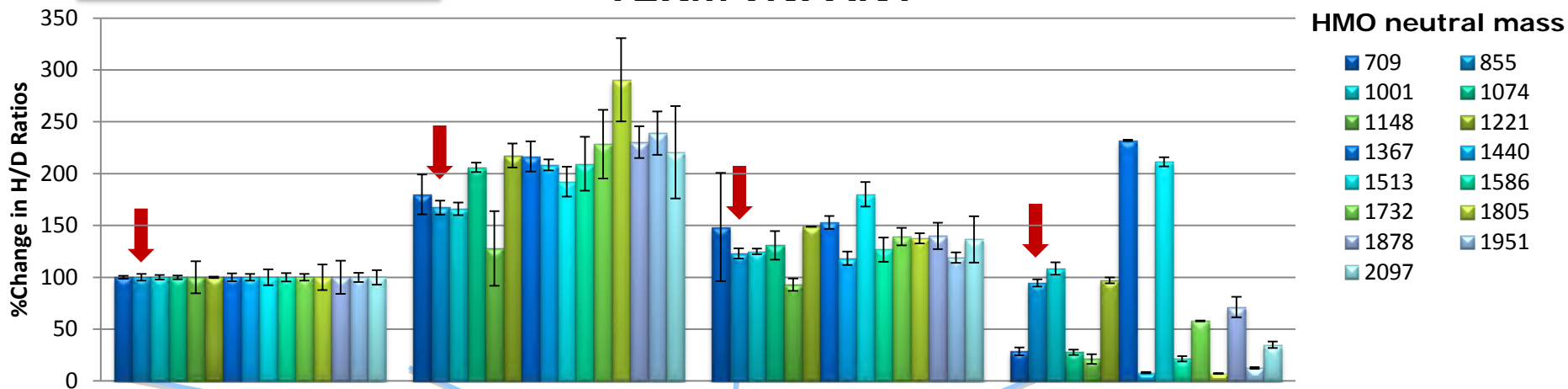


Specific oligosaccharides are consumed in the gut

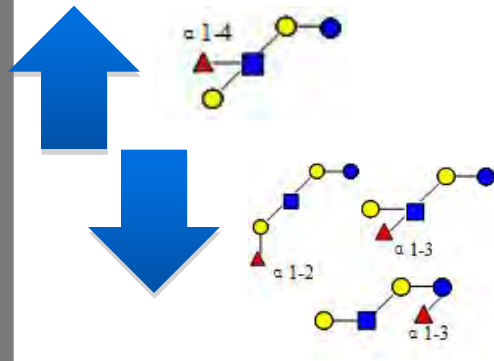
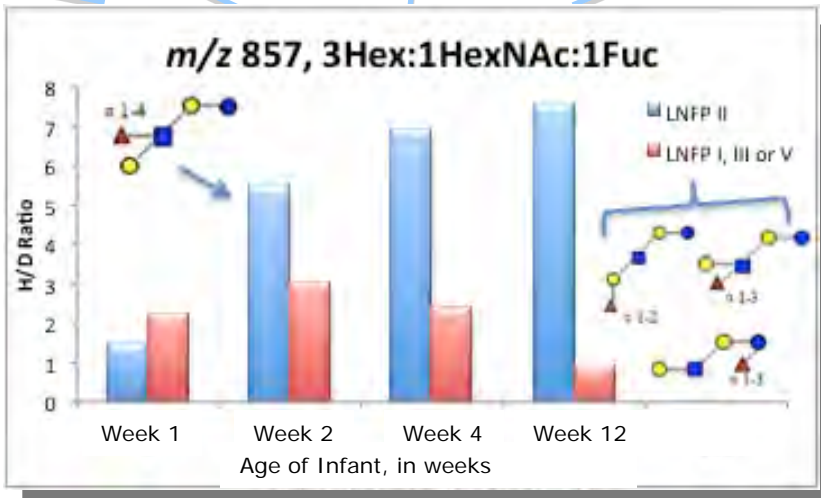


Fecal HMO Profile

TERM INFANT

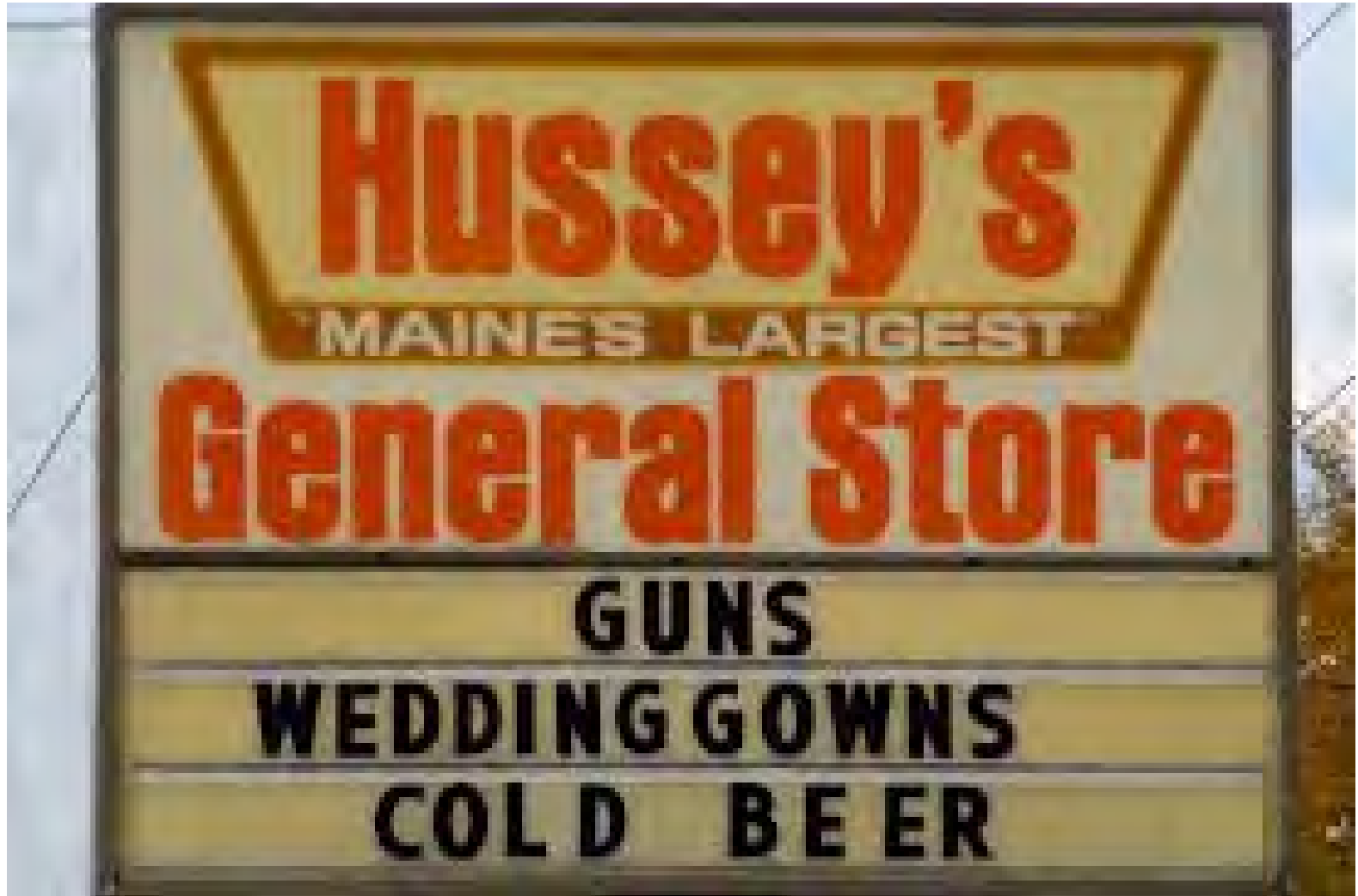


Glycan Specific Changes



Carlito Lebrilla
Lorna de Leoz
UCD Chemistry
(Submitted)

We need to be cautious on associations

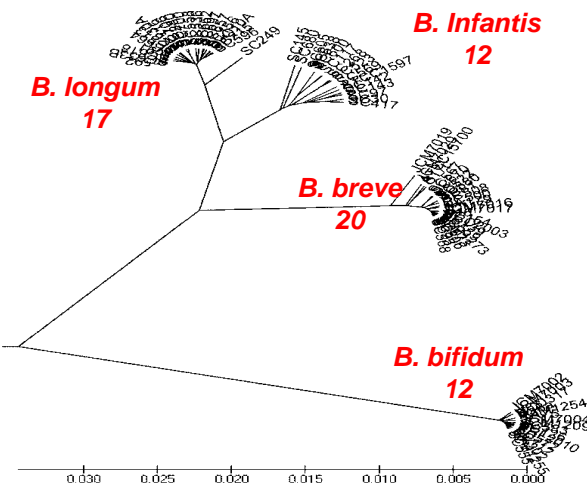


HMOs as “prebiotics” for Bifidobacteria

Which bifidobacterial species grow on HMO?

Which bifidobacteria grow on HMO sugars?

40 breast-fed infant (3-4 month old) stool samples
300 isolates → 74 characterized



Growth	Structure	<i>B. infantis</i>	<i>B. bifidum</i>	<i>B. longum</i>	<i>B. breve</i>
HMO		22/22	14/14	8/17	10/23
Lacto-N-tetraose		22/22	14/14	17/17	23/23
Lacto-N-neotetraose		22/22	14/14	2/17	23/23
2'-fucosyl lactose		22/22	13/14	1/17	2/23
3-fucosyl lactose		22/22	14/14	1/17	0
6-sialyl lactose		22/22	10/14	0	0

Multilocus
sequence tagging

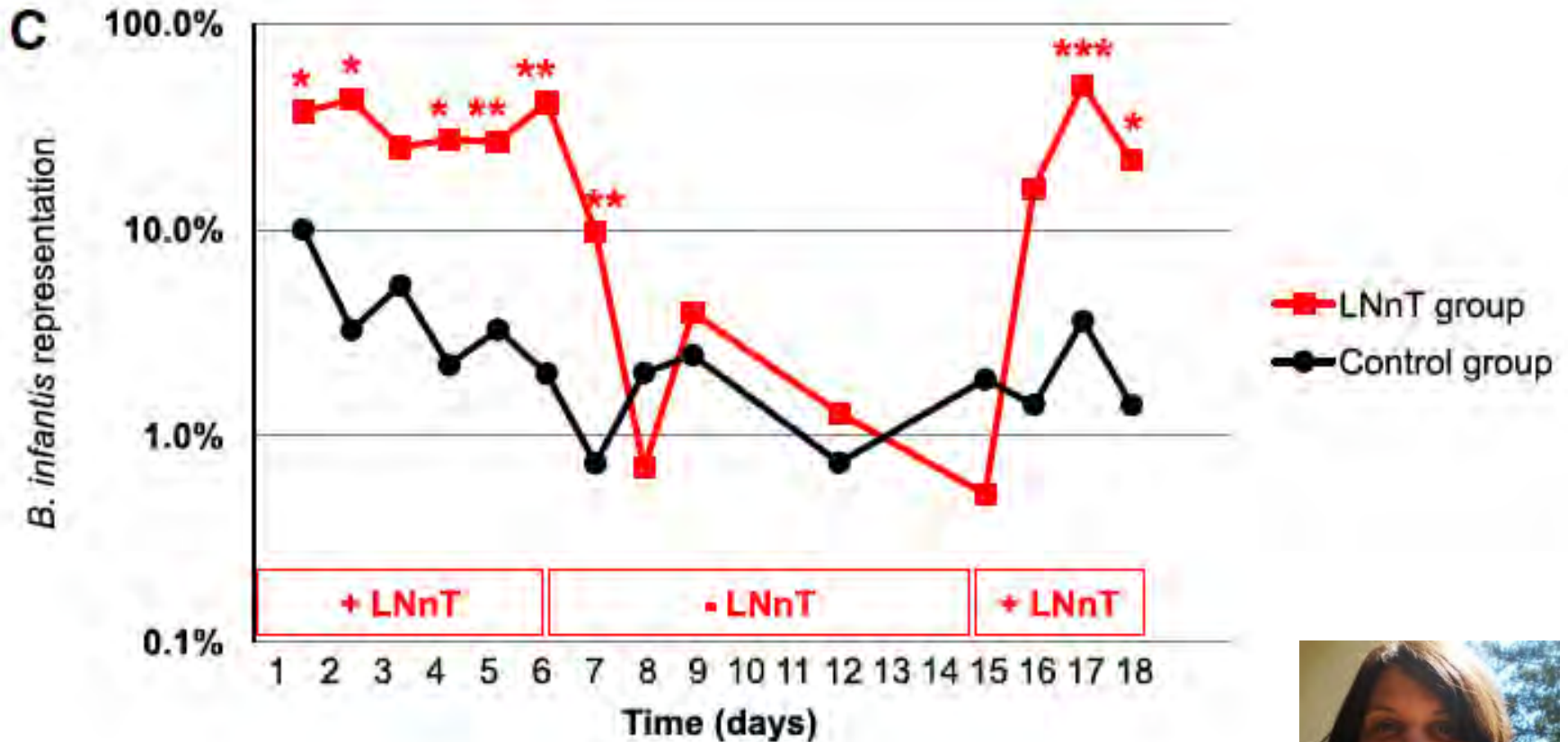


Santi Ruis Moyano

AEM 2006, MicroBiotech 2009, AEM 2013

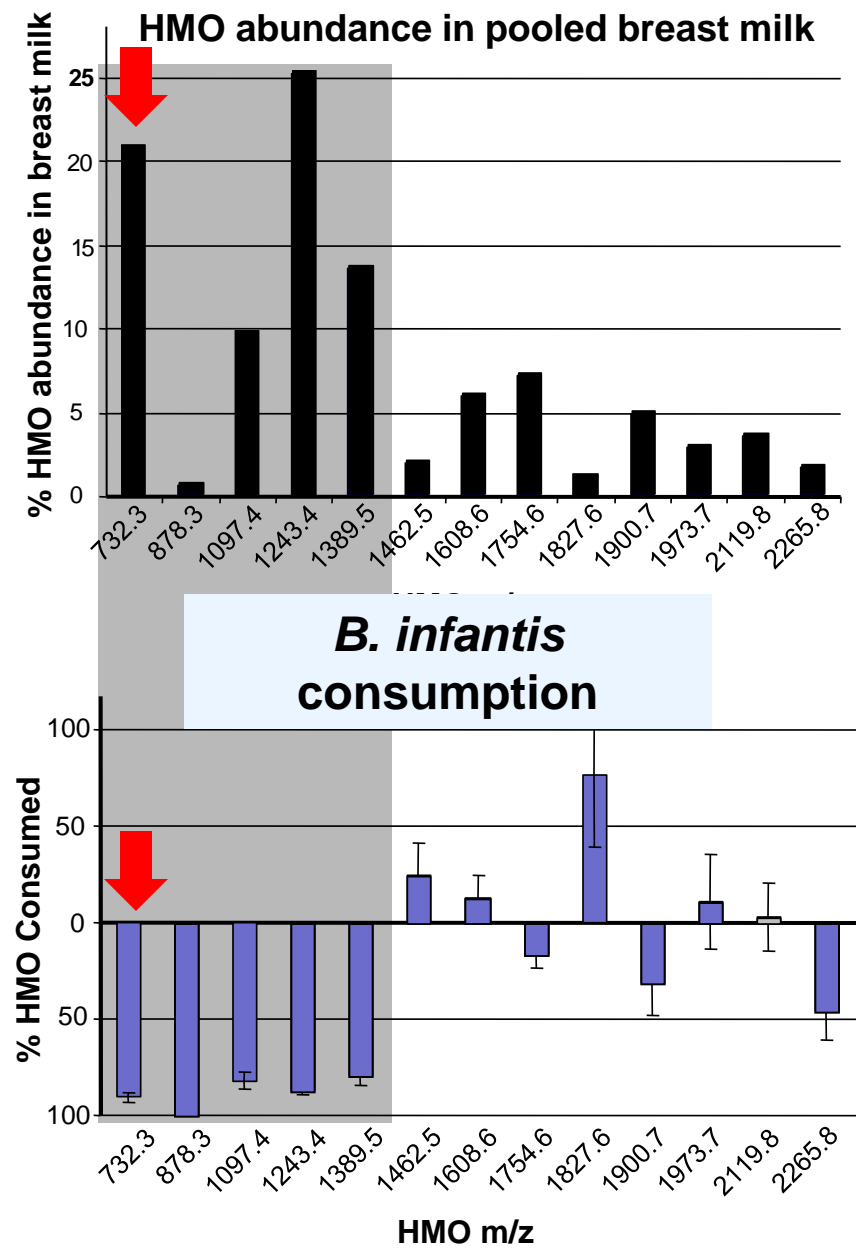
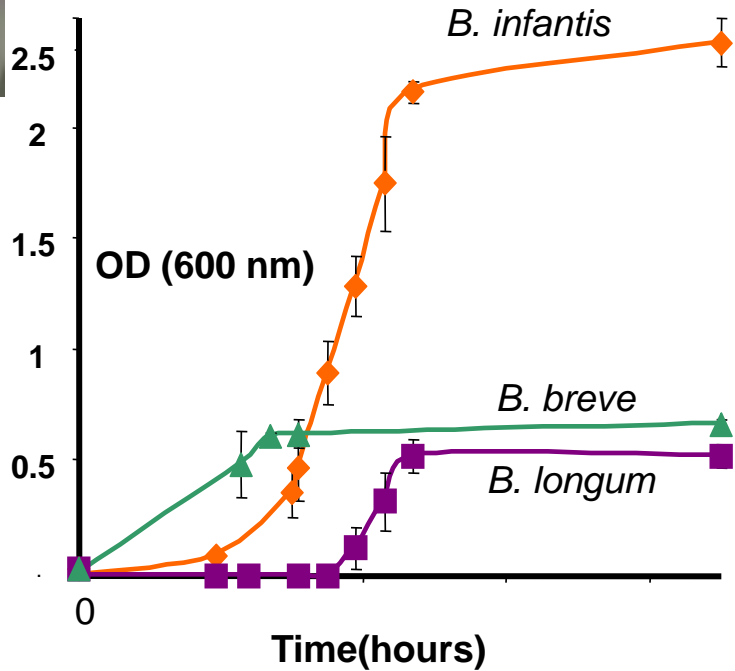
Bifidobacteria vs Bacteroides *in situ*

Lacto-N-neotetrose supplementation of gnotobiotic mice with *Bifidobacterium infantis* and *Bacteroides thetaiotaomicron*





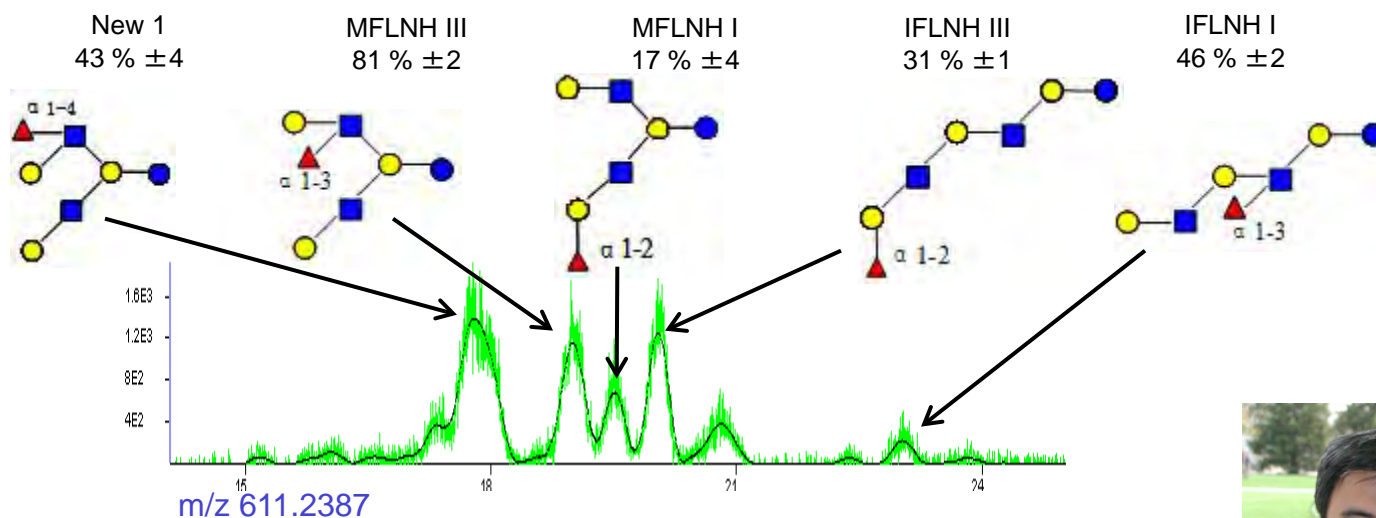
Bifidobacterial HMO Glycoprofiling



Several small MW oligosaccharides

Single HMO composition consumed by other bifidobacteria

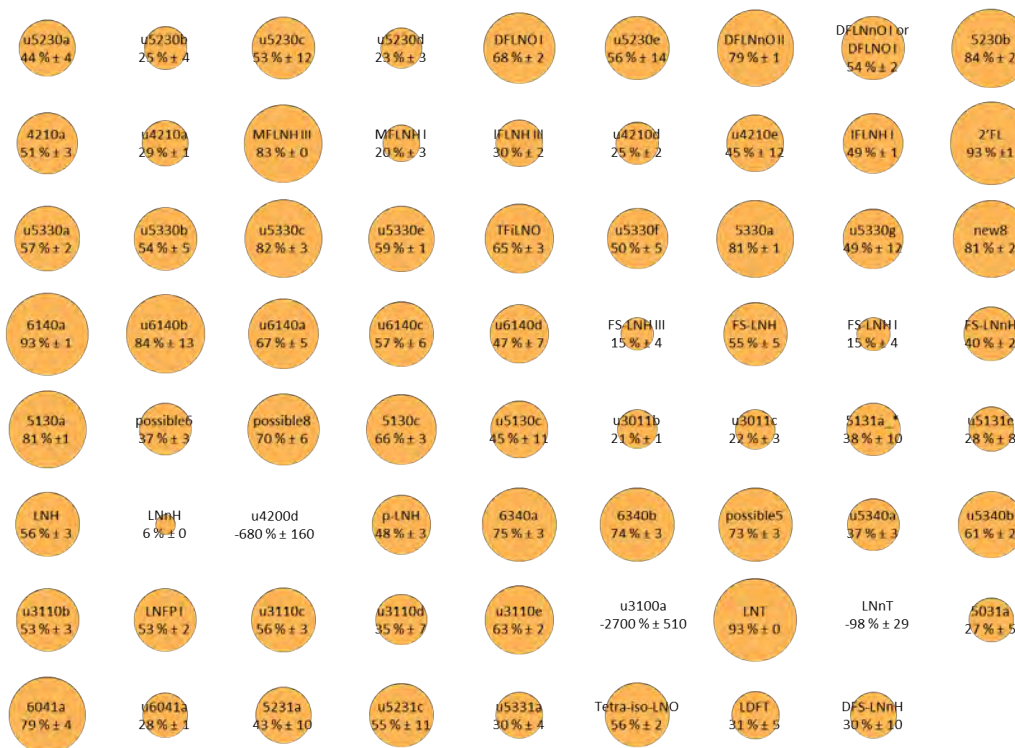
NanoLC separation of individual HMO compositions



Carlito Lebrilla
UCD Chemistry

Percent Consumption of HMO Structures

Consumption array of *B. infantis* ATCC 15697

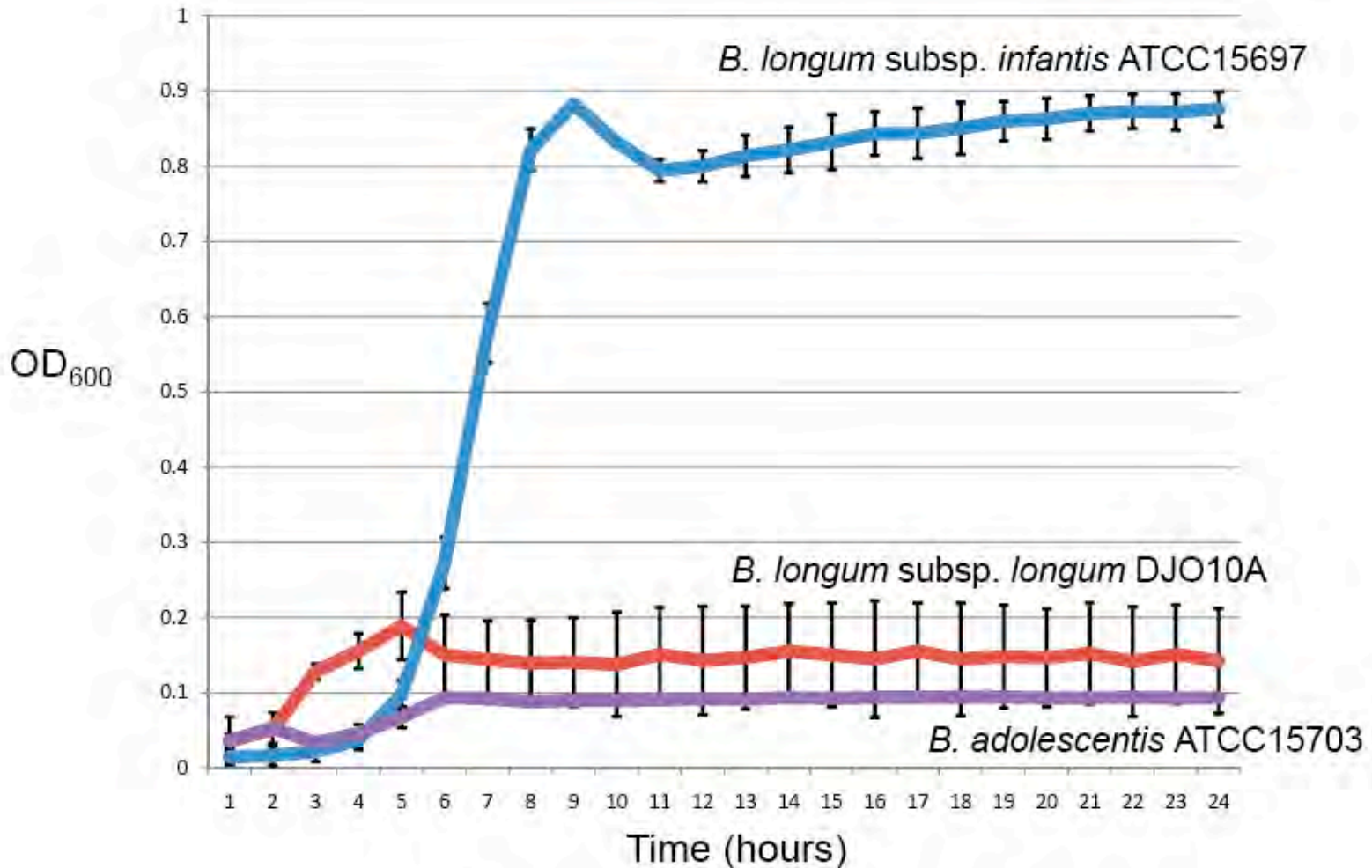


- Bacteria can be characterized by their HMO consumption profile.
- Orange circles are sized proportionally to the percent consumption.

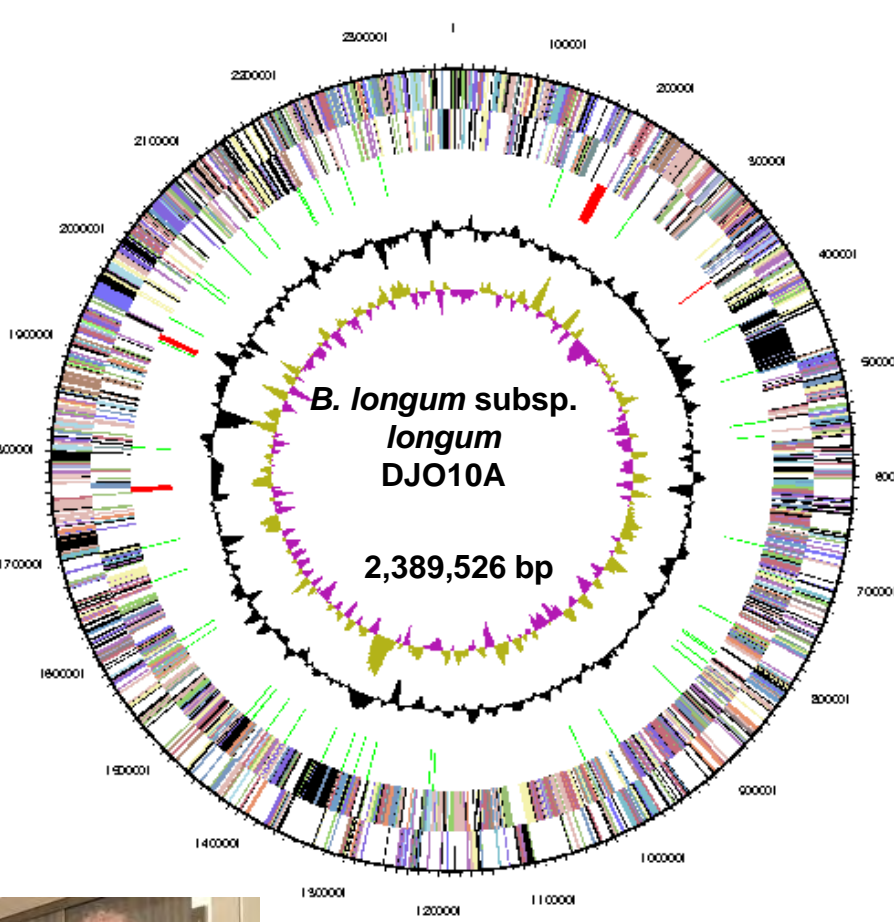


What genome features are required to utilize human milk oligosaccharides?

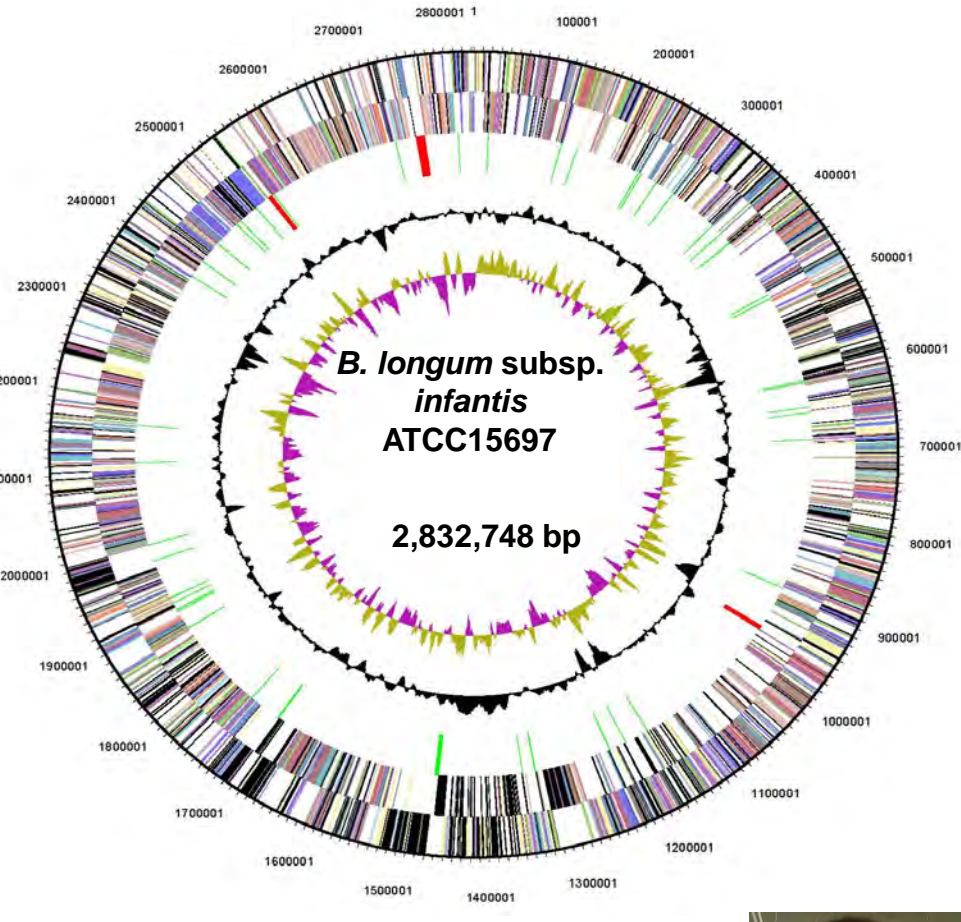
HMO utilization by Bifidobacteria



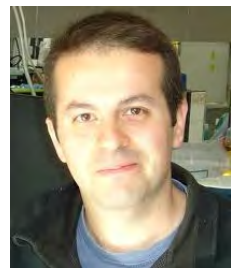
Comparative *Bifidobacterium* Genomics



Adult derived strain
BMC Genomics 2008

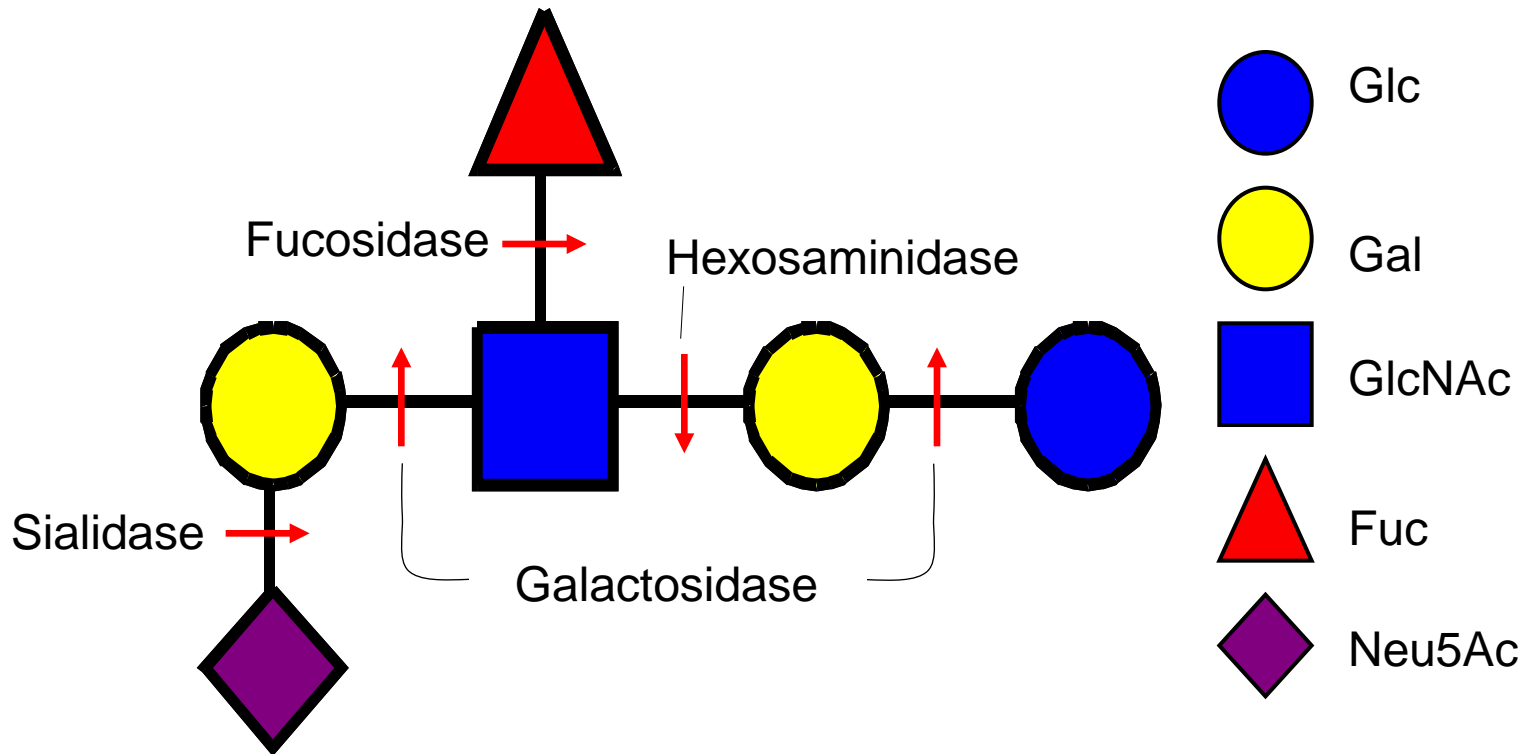


Infant derived strain
PNAS 2008



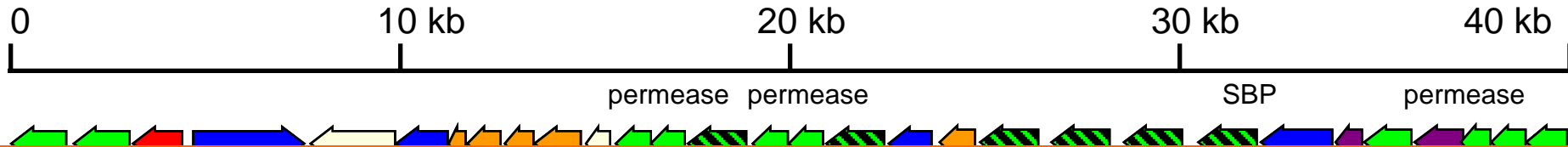
What's Needed to Deconstruct HMOs?

- Transport systems for oligo & monosaccharides
- Glycosyl hydrolases



B. infantis HMO cluster

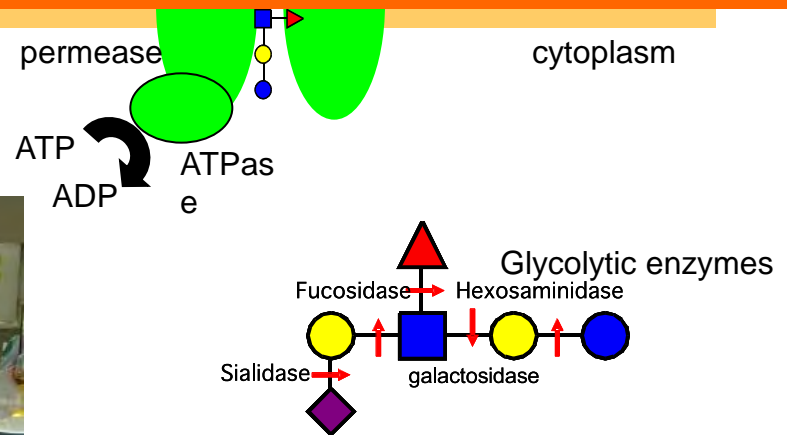
All 4 glycosyl hydrolases Array of oligosaccharide transporters



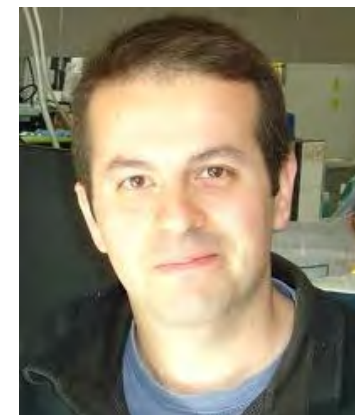
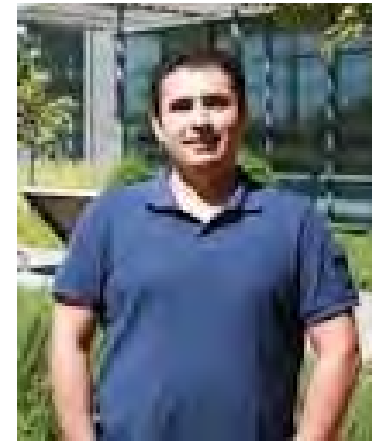
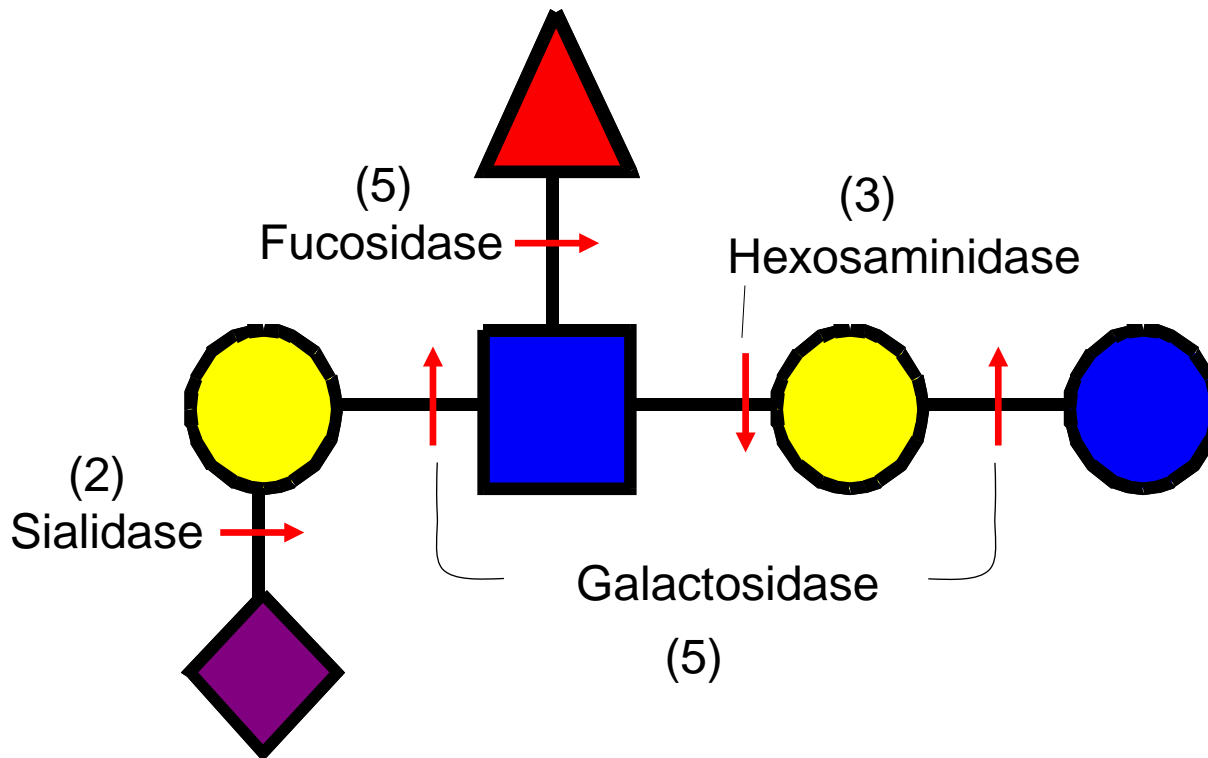
Genes unique to milk-associated bifidobacteria are uniquely expressed during growth on milk sugars

(PLoS One 2013, unpublished)

- ATP hydrolysis prompts transport of oligosaccharides across membrane
- Intracellular glycolytic enzymes deconstruct oligosaccharide



Characterization of the glycosidases and transporters from *B. infantis*?

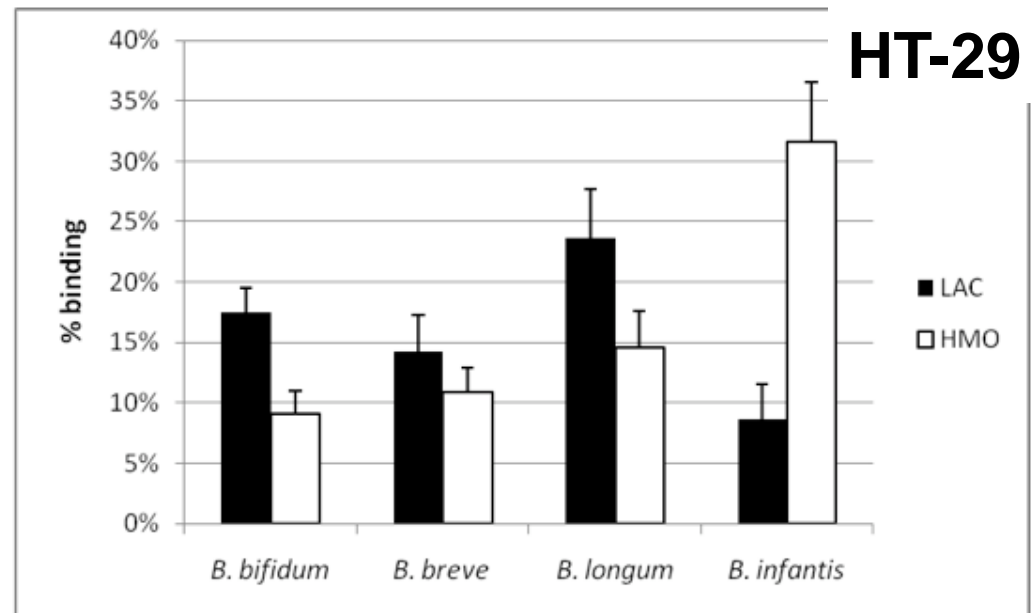
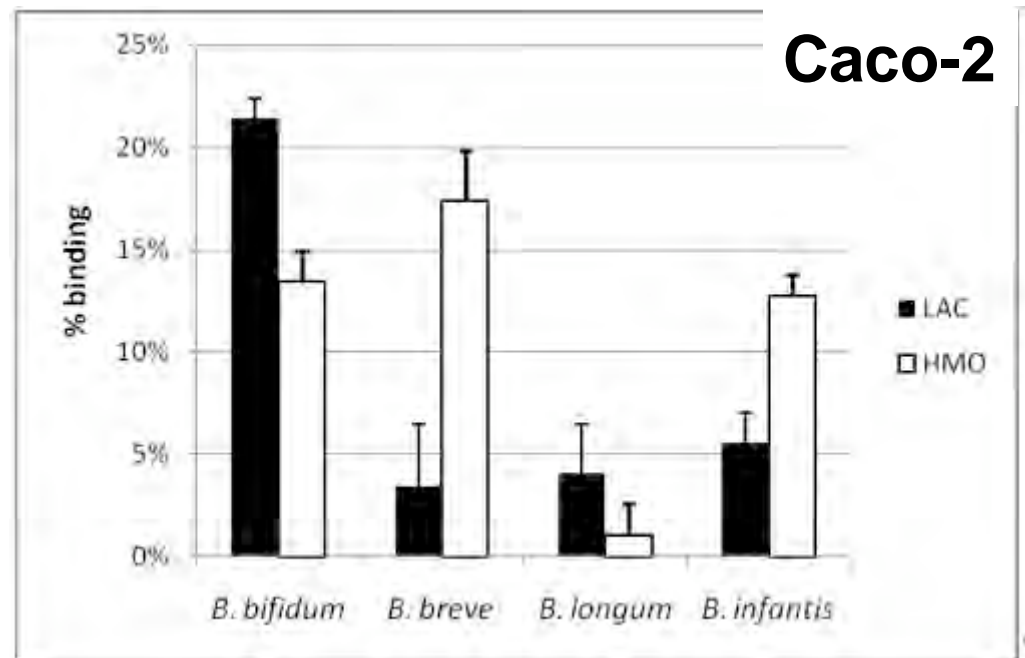


Sialidases - -----
 Fucosidases - -----
 Hexosaminidases - -----
 Galactosidases - -----
 Surface Binding Proteins

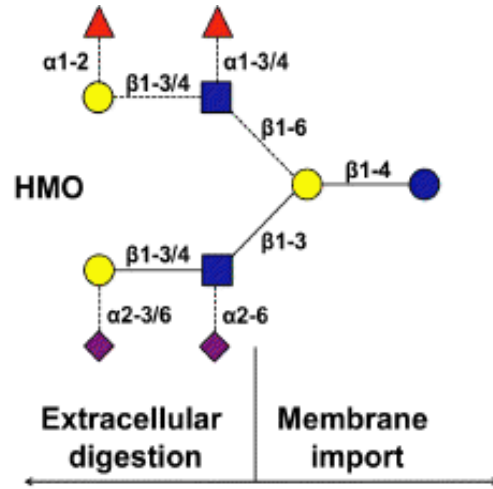
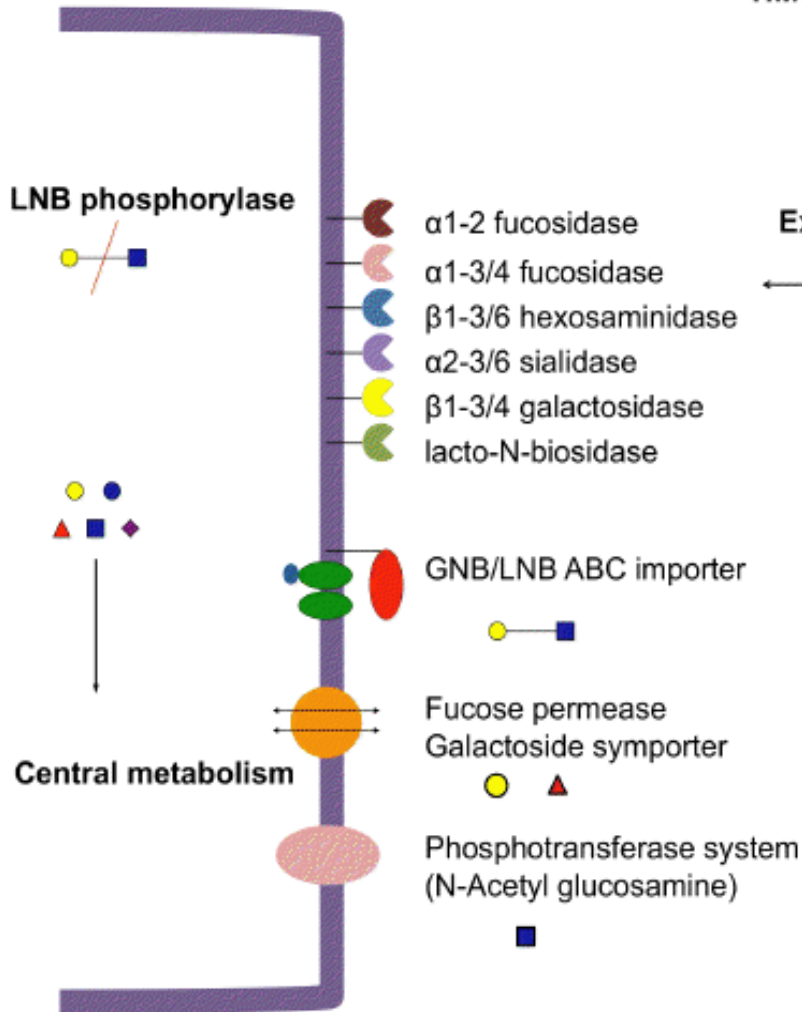
Sela et al JBC 2011
 Sela et al AEM 2012
 Garrido et al Anaerobe 2012
 Garrido et al Food Micro 2012
 Garrido et al PLoS One 2011

Growth on milk oligosaccharides helps **some** bifidobacteria bind intestinal cells

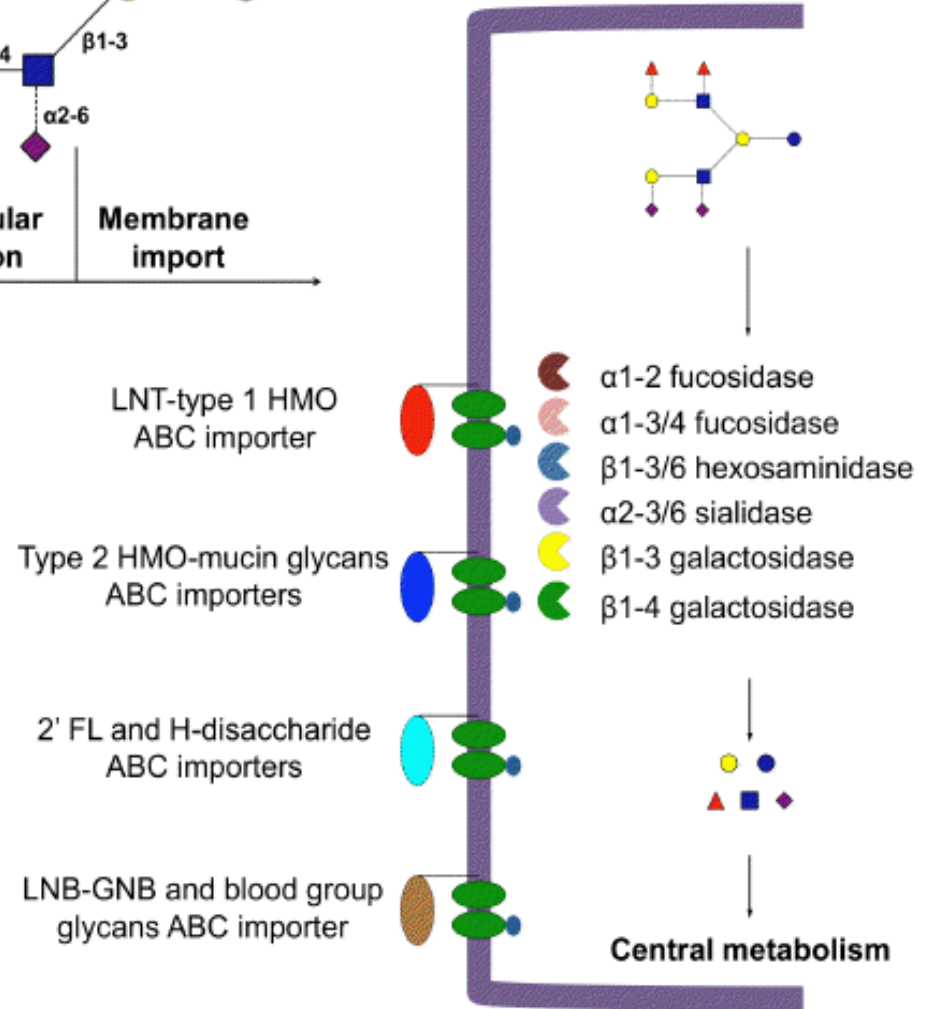
- HMO vs Lac grown cells:
- Induce TJ proteins
 - Induce anti-inflammatory cytokines (IL-10)



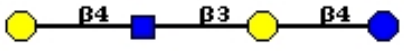
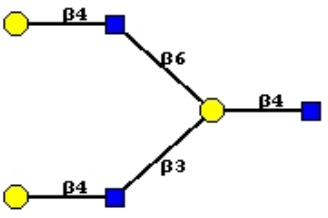
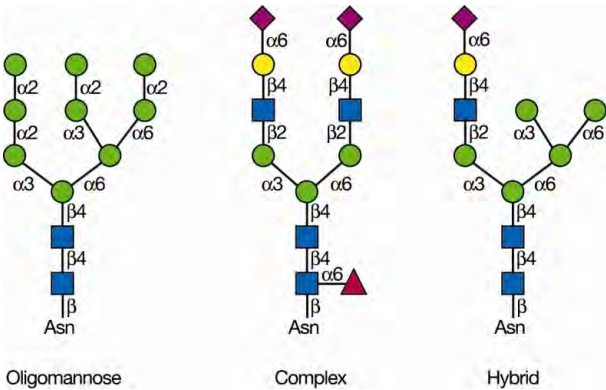
Bifidobacterium bifidum



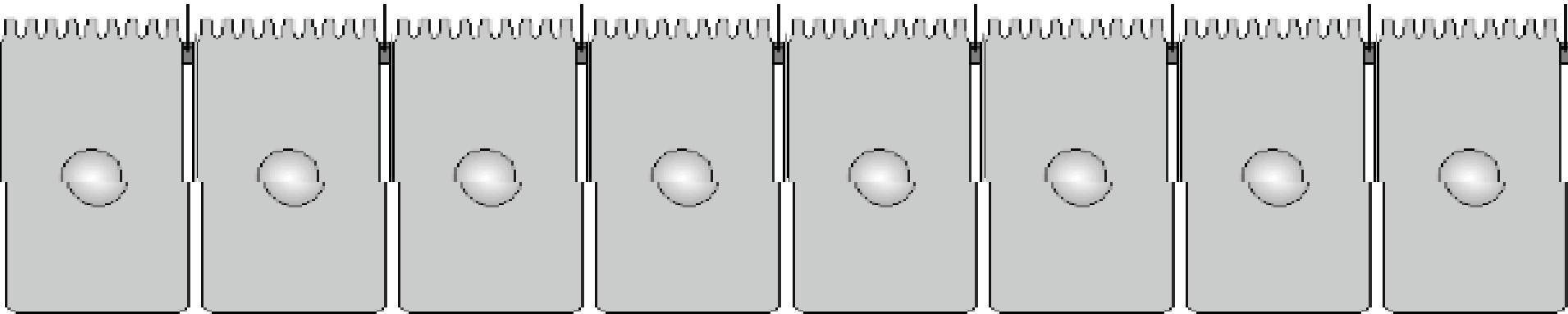
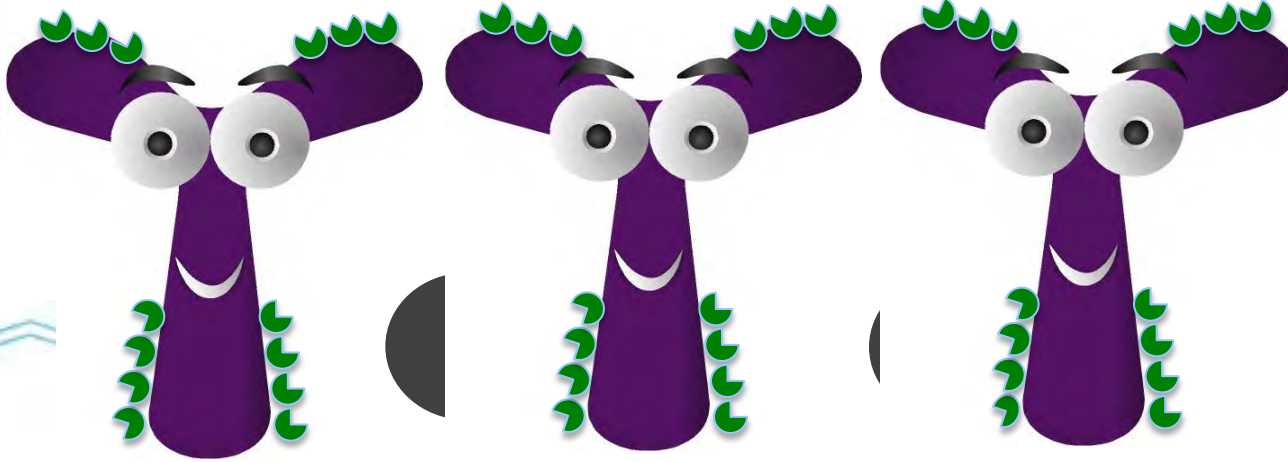
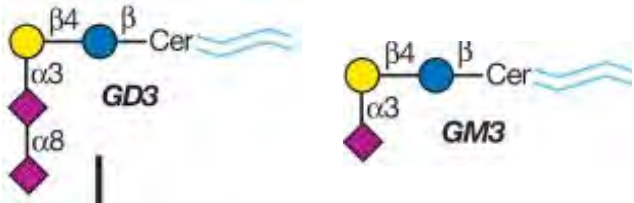
Bifidobacterium infantis



Model for bifidobacterial enrichment in the infant GIT

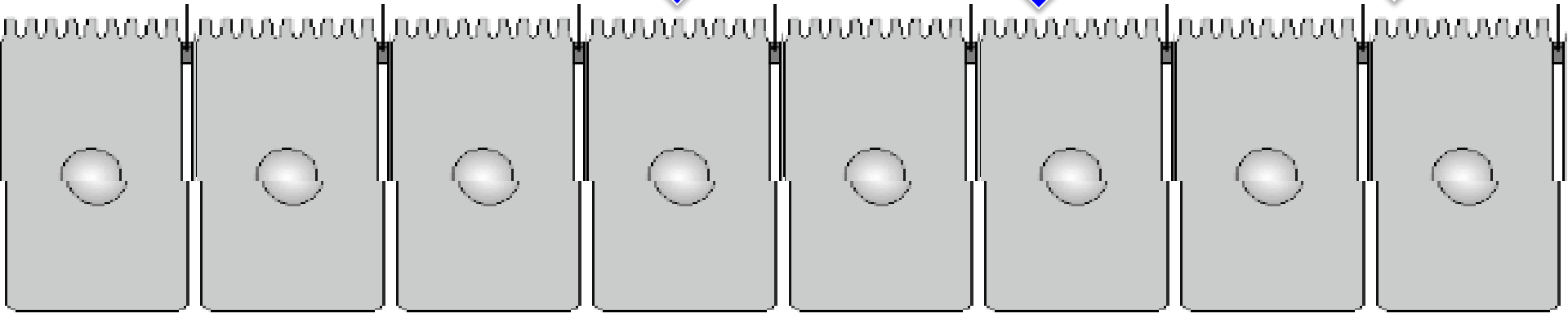
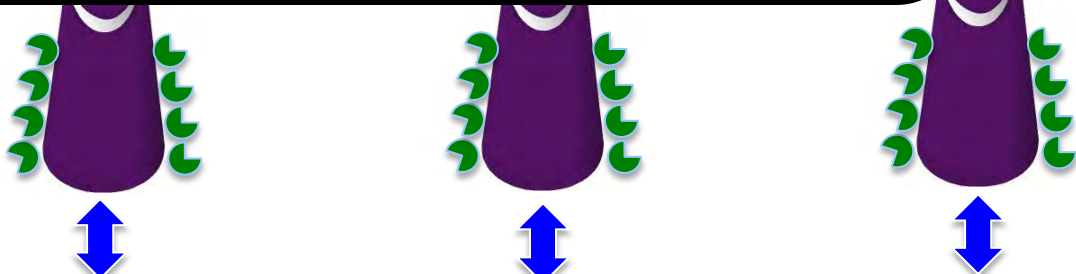
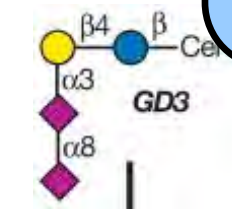
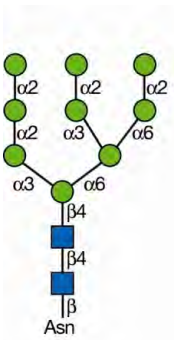


Oligomannose Complex Hybrid



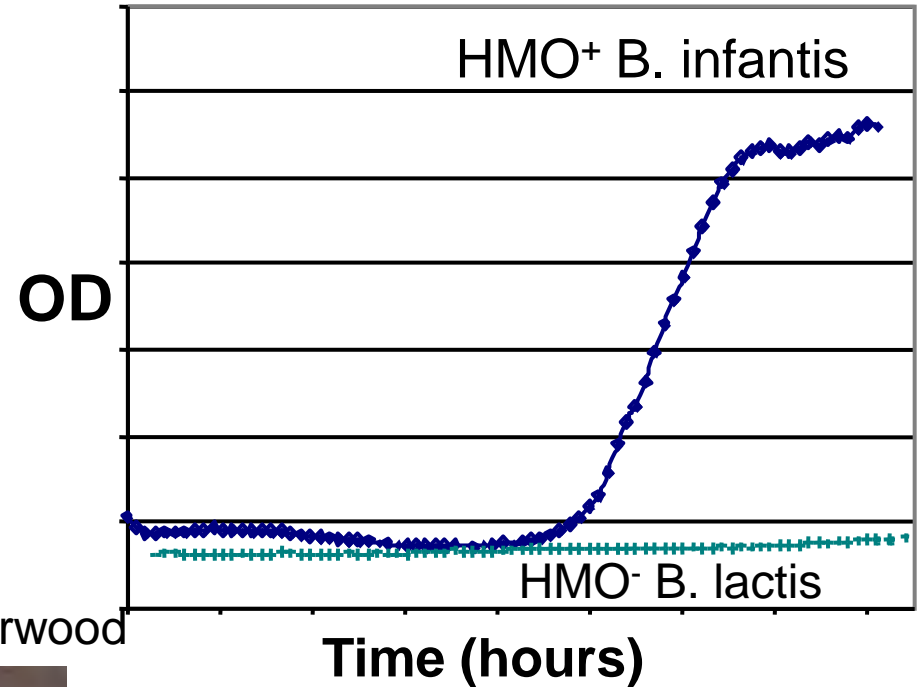
Model for bifidobacteria enrichment in the infant GIT

Complex milk glycans enhance efficacy of specific bifidobacteria

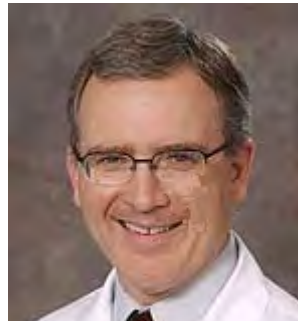


Can this knowledge be
translated?

Will synbiotic feeding HMO+ *B. infantis* with HMOs help establish bifidobacteria?



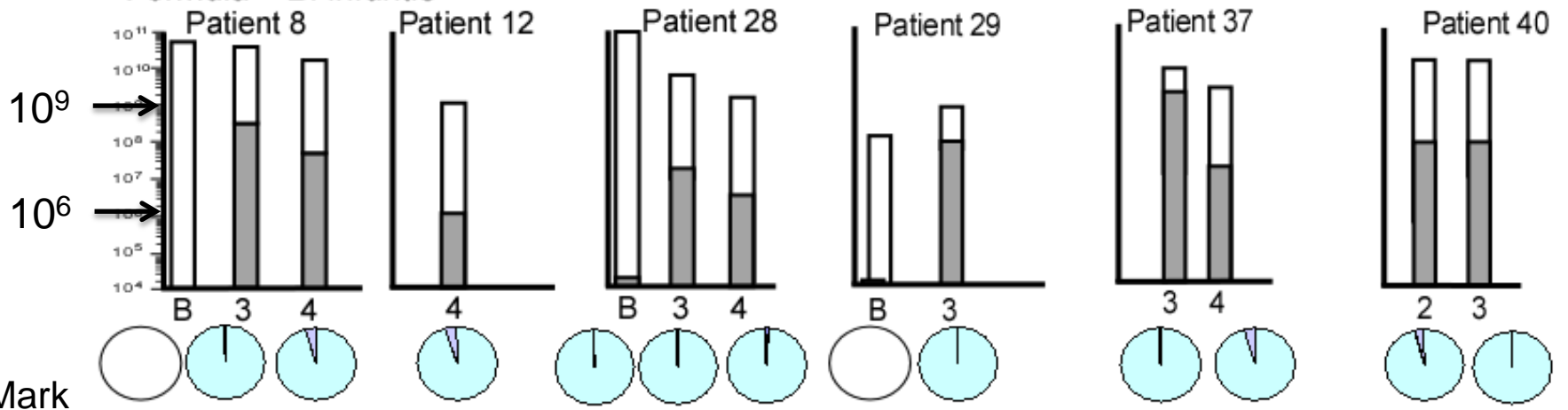
Mark Underwood



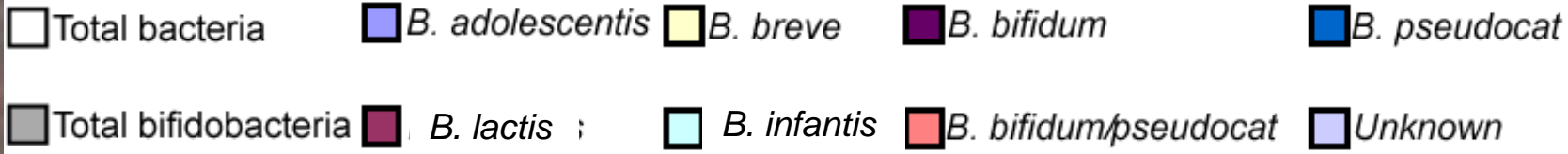
UCD Med School
Neonatology



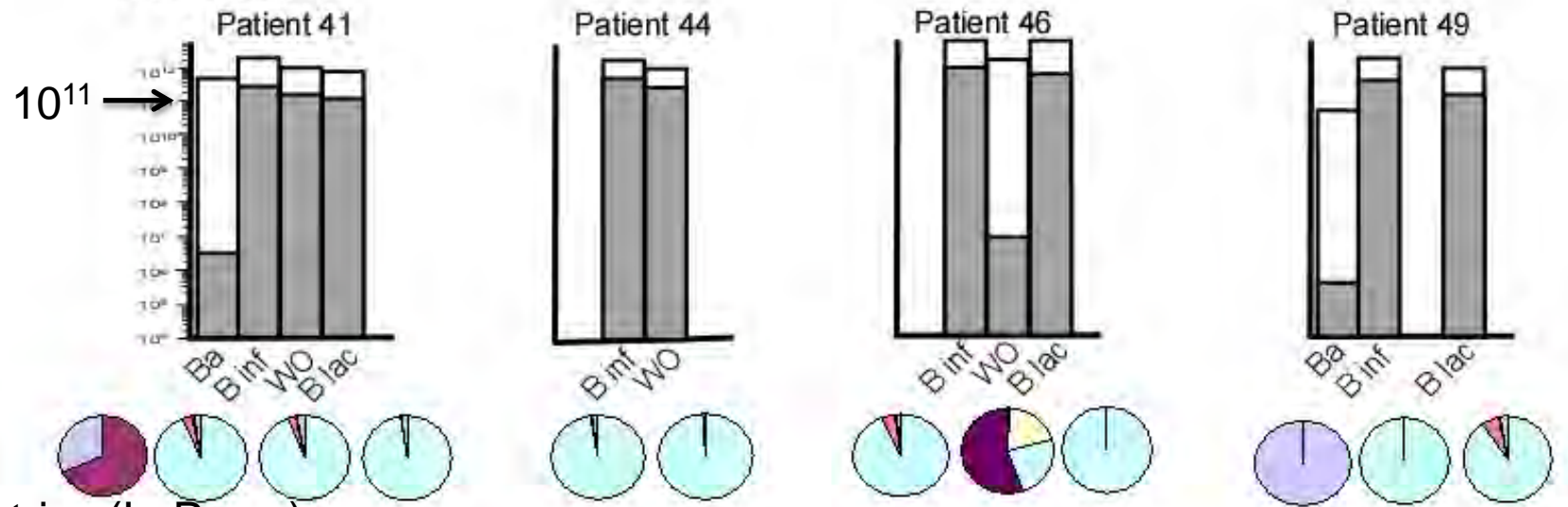
Formula + *B. infantis*



Mark Underwood



H+Binf/Blac



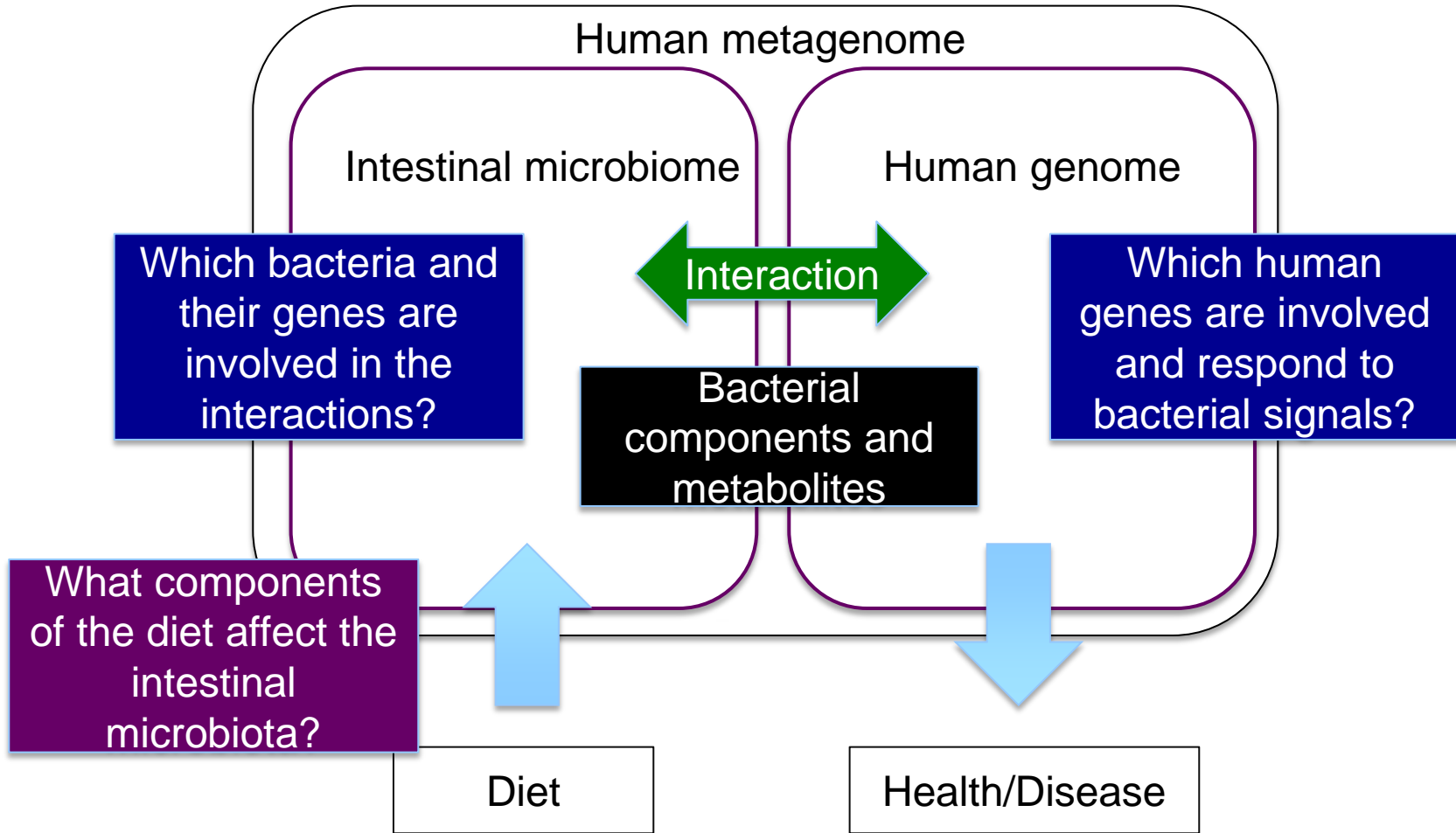
Take home points....

- Milk provides a model of establishing (modulating) a microbiota
- Specificity of that modulation is driven in part by glycan complexity and cognate bacterial catabolism
- Exploiting that knowledge to partner specific glycans with specific cognate bifidobacteria can enable more persistent colonization in humans

...this took detailed mechanistic research....

...mechanism leads to translational diagnostics...

Gaps/Needs/Challenges

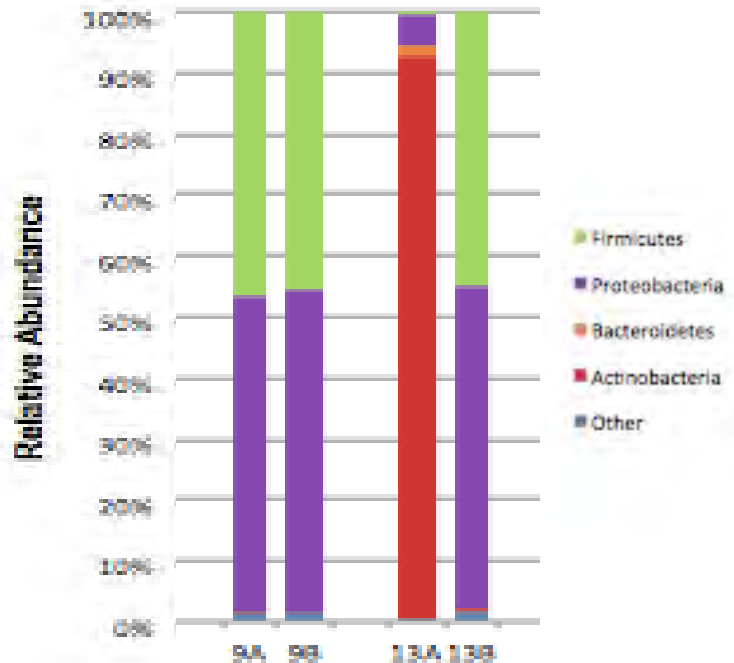


Gaps/Needs/Challenges

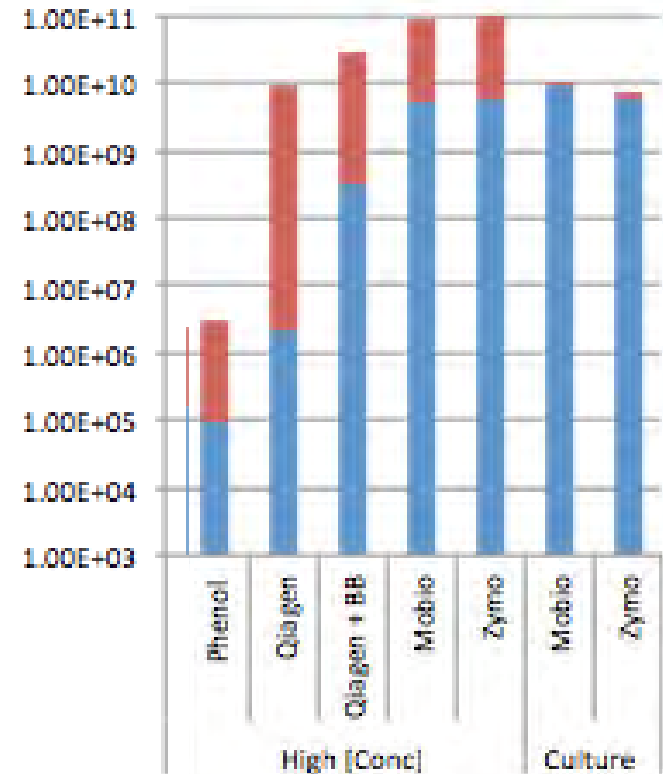
- Mechanistic research needed (systems biology + strain level examination)
- Interdisciplinary teams (thanks Vince)
- Better (supported) animal models (thanks Gary)
- Continued tool development...
 - Metabolomic/Metagenomic
 - Genetic tools!!
 - Glycomics
- Ability to stratify clinical populations

Needs/Challenges

Harmonizing of protocols – are we settled on a DNA prep?



8F forward primer misses bifidobacteria (need to spike in a 8Fbif primer)



Lack of bead beating lowers bifidobacteria

Lack of accurate annotation/function will impede or misdirect microbiome work

[Home](#) > Find Genomes

Loaded.

Bifidobacterium longum infantis ATCC 15697

[Add to Genome Cart](#)

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[Download Data](#)

About Genome

- [Overview](#)
- [Statistics](#)
- [Genes](#)

Genes total number	2577	100.00%
Protein coding genes	2486	96.47%
Pseudo Genes	72	2.79% ²
RNA genes	91	3.53%
rRNA genes	12	0.47%
5S rRNA	4	0.16%
16S rRNA	4	0.16%
23S rRNA	4	0.16%
tRNA genes	79	3.07%
Protein coding genes with function prediction	1567	60.81%
without function prediction	919	35.66%



Media/Public/Physician Confusion on Probiotics

Table 1. Key genera and species of microbes studied and used as probiotics

Genus	Species
<i>Lactobacillus</i>	<i>acidophilus</i> <i>brevis</i> <i>delbrueckii</i> ^a <i>fermentum</i> <i>gasseri</i> <i>johnsonii</i> <i>paracasei</i> <i>plantarum</i> <i>reuteri</i> <i>rhamnosus</i> <i>salivarius</i>
<i>Bifidobacterium</i>	<i>adolescentis</i> <i>animalis</i> ^b <i>bifidum</i> <i>breve</i> <i>infantis</i> <i>longum</i>
<i>Streptococcus</i>	<i>thermophilus</i> <i>salivarius</i>
<i>Enterococcus</i>	<i>faecium</i>
<i>Escherichia</i>	<i>coli</i>
<i>Bacillus</i>	<i>coagulans</i> ^c <i>clausii</i>
<i>Saccharomyces</i>	<i>cerevisiae</i> ^d

Reality of the research so far →
Probiotic action occurs at the level of
strains

But the public perception is not at the strain level



Our kombucha is raw meaning that it has never been pasteurized nor heat treated. As a living product, new cultures will continue to form even once bottled. Sometimes they are clear, like egg whites, and other times they are brownish in color. These cultures are harmless and indicate that the beverage is live and rich in probiotics. It's completely edible (go ahead, be bold) but if you prefer, please strain.

What's that floating
in my kombucha?

Are you delivering the probiotic species and strain you think you are?

Product	Microorganisms listed on the product label	T-RFLP	Species-specific PCR	Additional microbe T-RFLP patterns detected
1	<i>Lb acidophilus</i>	+	+	<i>Lb brevis</i> , <i>Lb plantarum</i> [*] , <i>Lb jonhsonii</i> [†] , <i>Lb amylolyticus</i> , <i>Lactobacillus</i> sp [‡]
	<i>B bifidum</i>	–	+	
	<i>L helveticus</i>	+	+	
	<i>S thermophilus</i>	+	+	
2	<i>Lb acidophilus</i>	+	+	<i>Lb brevis</i> , <i>Lb plantarum</i> [*] , <i>Lb amylolyticus</i> , <i>Lactobacillus</i> sp [‡]
	<i>B bifidum</i>	+	+	
	<i>Lb helveticus</i>	+	+	
	<i>Lb rhamnosus</i>	+	+	
5	<i>Lb acidophilus</i>	+	+	<i>Lb bif fermentus</i> , <i>Lactobacillus</i> sp [‡] , <i>L lactis</i> , <i>Lb rhamnosus</i> [§]
	<i>B bifidum</i>	–	+	
	<i>Lb helveticus</i>	+	+	
6	<i>Lb acidophilus</i>	+	+	<i>Lb bif fermentus</i> , <i>Lb sanfranciscensis</i> , <i>Lactobacillus</i> sp [‡] , <i>Lb plantarum</i> [*] , <i>Lb fructivorans</i>
	<i>B bifidum</i>	–	+	
	<i>Lb helveticus</i>	+	+	
7	<i>Lb acidophilus</i>	+	+	<i>Lb plantarum</i> [*]
	<i>B bifidum</i>	–	+	
	<i>B longum</i>	+	+	
8	<i>Lb acidophilus</i>	+	+	<i>Lb bif fermentus</i> , <i>Lactobacillus</i> sp [‡] , <i>Lb helveticus</i> , <i>Lb plantarum</i> [*] , <i>Lb rhamnosus</i> [§]
	<i>B longum</i>	+	+	
9	<i>B longum</i>	+	+	
	<i>B bifidum</i>	+	+	
10	<i>Lb acidophilus</i>	+	+	<i>L lactis</i> , <i>Lb rhamnosus</i> , <i>Lb plantarum</i> [*] , <i>Lb jonhsonii</i> [†]
11	<i>Lb acidophilus</i>	+	+	<i>Lb bif fermentus</i> , <i>Lactobacillus</i> sp [‡] , <i>Lb helveticus</i> , <i>Lb plantarum</i> [*] , <i>Lb rhamnosus</i> [§]
12	<i>Lb acidophilus</i>	–	+	<i>Lb plantarum</i> [*] , <i>Lb sanfranciscensis</i> , <i>Lb helveticus</i> , <i>Lactobacillus</i> sp [‡]
13	<i>Lb casei</i>	+	+	<i>Lb casei</i> , <i>Lb rhamnosus</i> [§]
14	<i>Lb acidophilus</i>	+	+	<i>Lb rhamnosus</i> [§] , <i>Lb plantarum</i> [*] , <i>Lactobacillus</i> sp [‡] , <i>B animalis</i> , <i>Lb amylolyticus</i> , <i>Lactobacillus</i> sp [§]
	<i>B longum</i>	+	+	
	<i>B bifidum</i>	+	+	

Understanding Responder Non-Responder Issues

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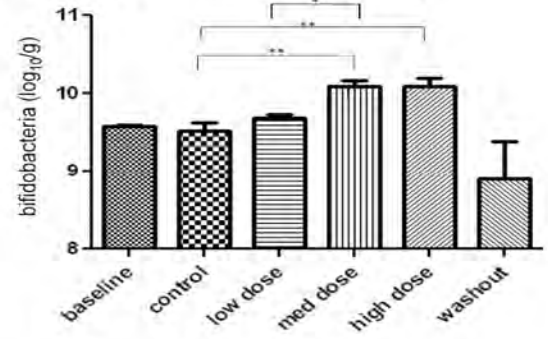


Barcoded Pyrosequencing Reveals That Consumption of Galactooligosaccharides Results in a Highly Specific Bifidogenic Response in Humans

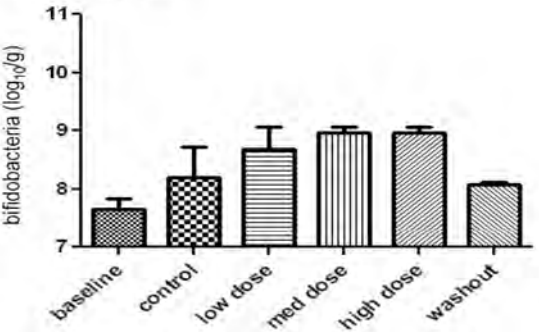
Lauren M. G. Davis¹, Inés Martínez¹, Jens Walter¹, Caitlin Goin², Robert W. Hutkins^{1*}

¹Department of Food Science and Technology, University of Nebraska, Lincoln, Nebraska, United States of America, ²School of Biological Sciences, University of Nebraska, Lincoln, Nebraska, United States of America

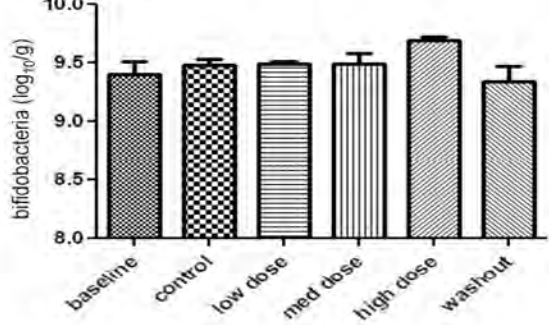
Subject 2



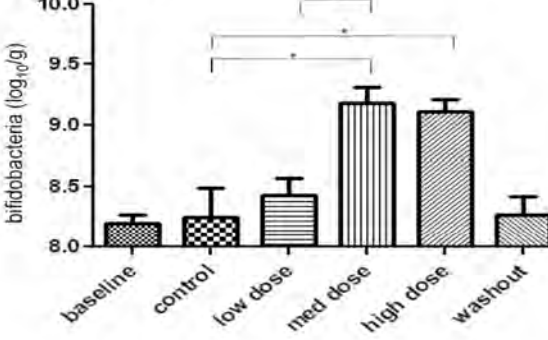
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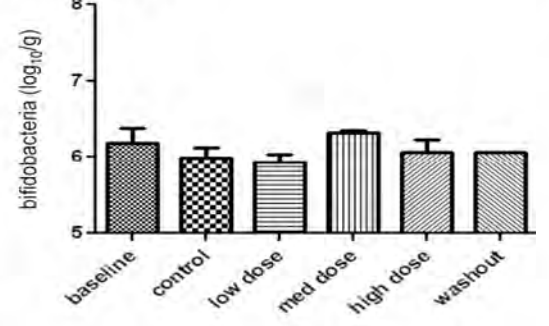
Subject 11



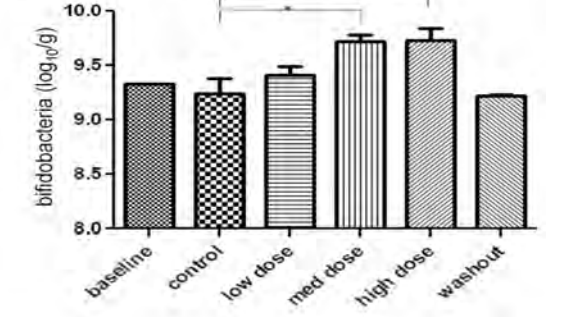
Subject 12



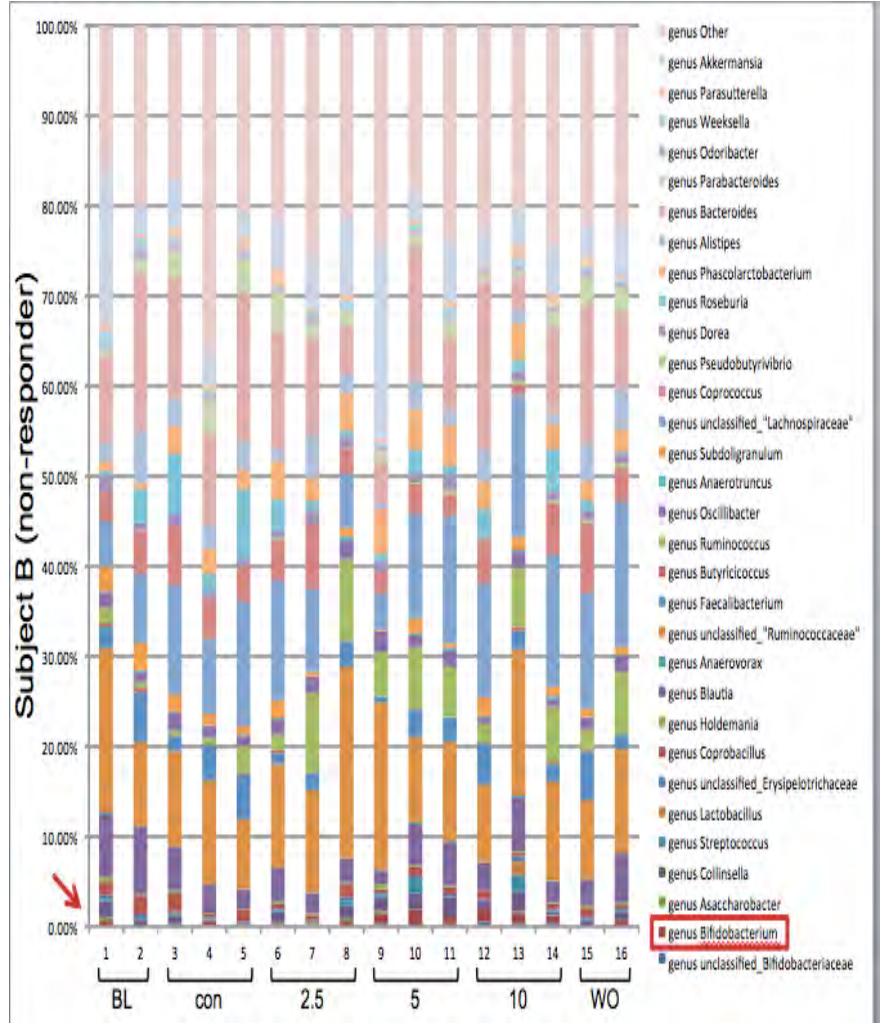
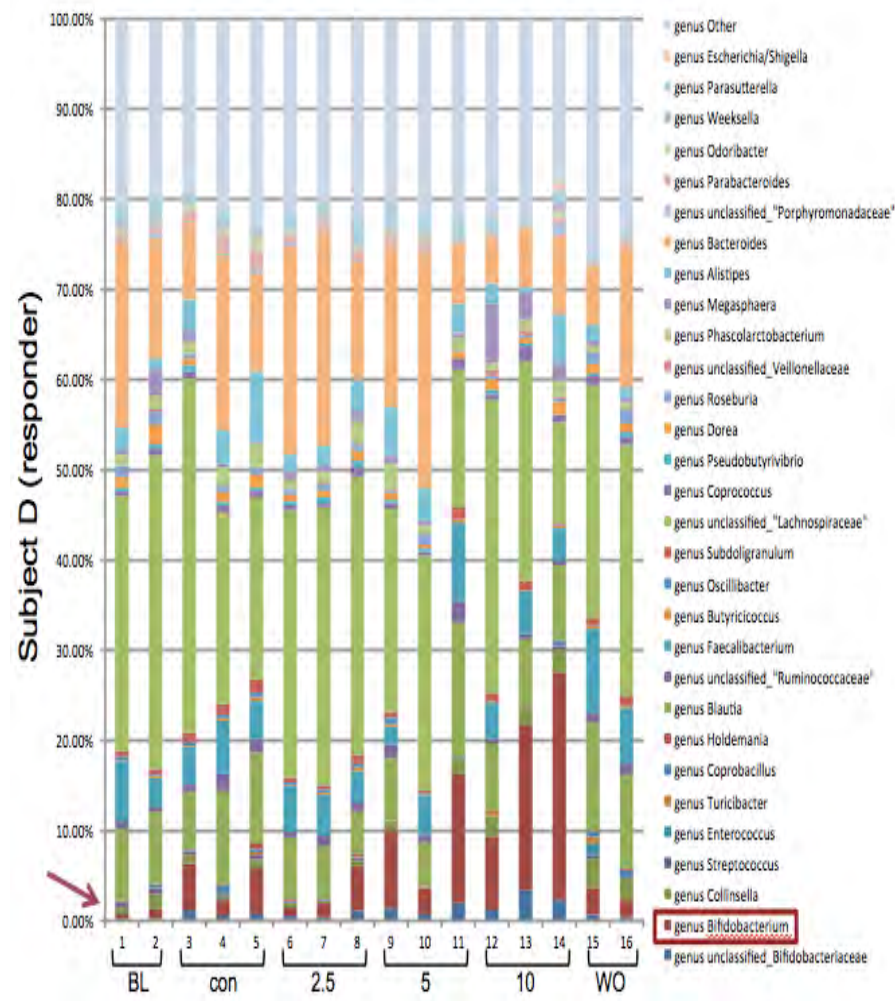
Subject 16



Subject 17



Understanding Responder Non-Responder Issues



PIs: **Carlito Lebrilla, J. Bruce German, Xi Chen, Mark Underwood, Chuck Bevins, Helen Raybould**

Students/Postdocs: **David Sela, Maciej Chichlowski, Karen Kalanetra, Santiago Ruiz-Moyano, Milady Ninonuevo, Riccardo LoCascio, Yanhong Lin, Larry Lerno, Jae Han Kim, Mariana Barboza, Scott Kronewitter, Richard Siepert, Aaron Adamson, Daniel Garrido, Angela Marcobal, Robert Ward and Samara Freeman**



Acknowledgements



the UC
Discovery Grant

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and Alternative Medicine

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Conflict of Interest Statement

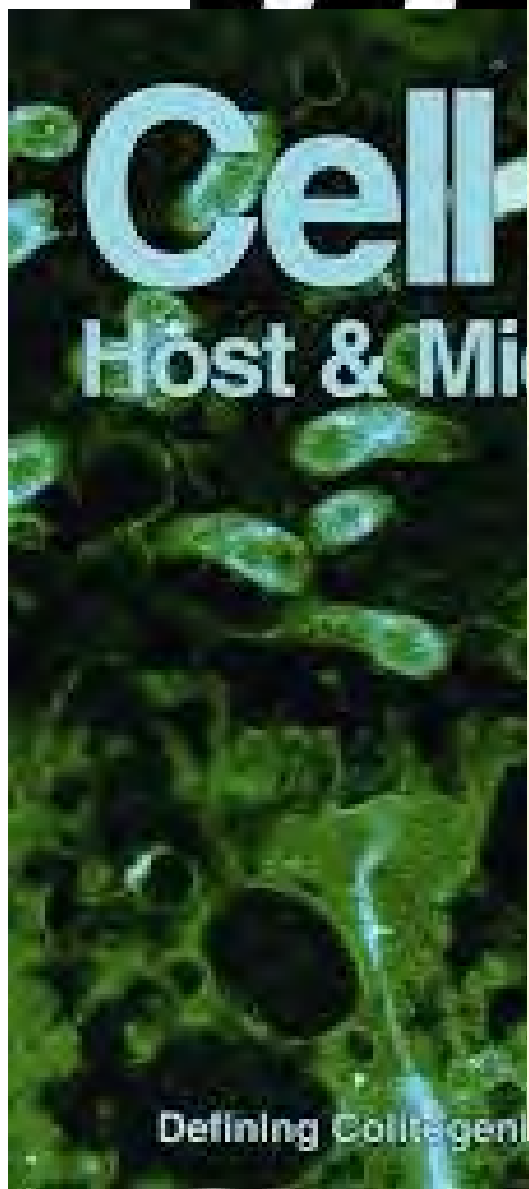
Co-Founder – Evolve Biosystems Inc.

Co-Founder – MicroTrek Inc.



PNAS

Probiotic
Lacto

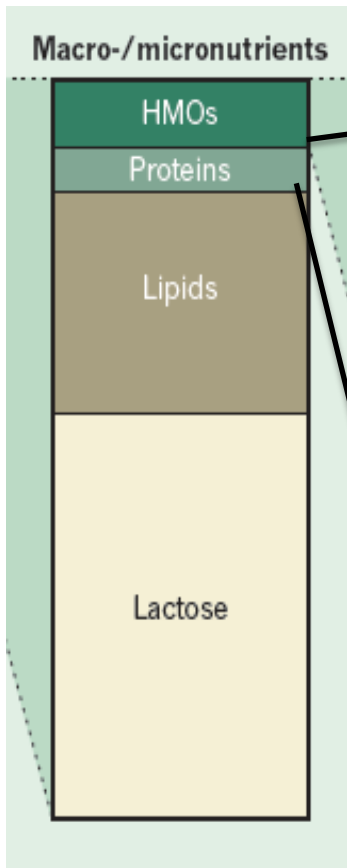


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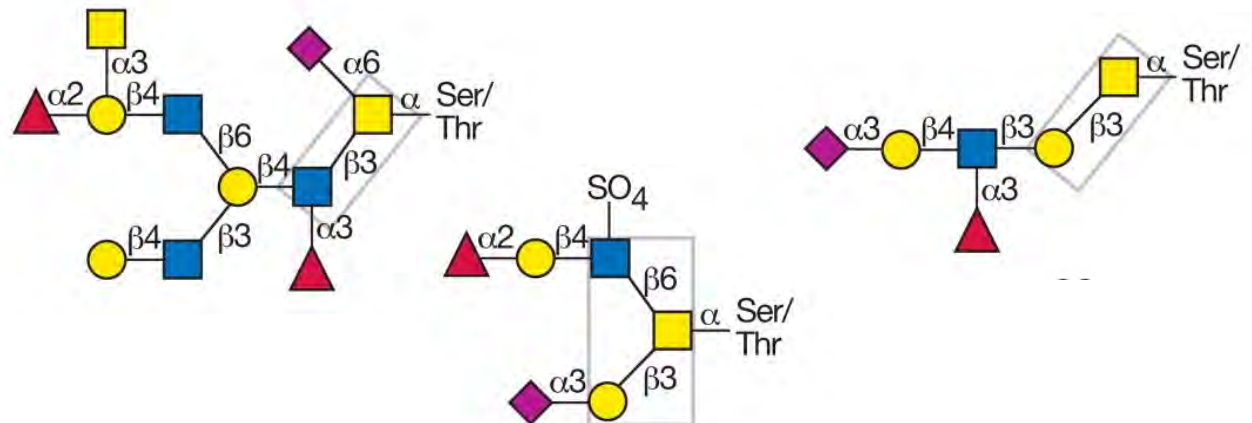
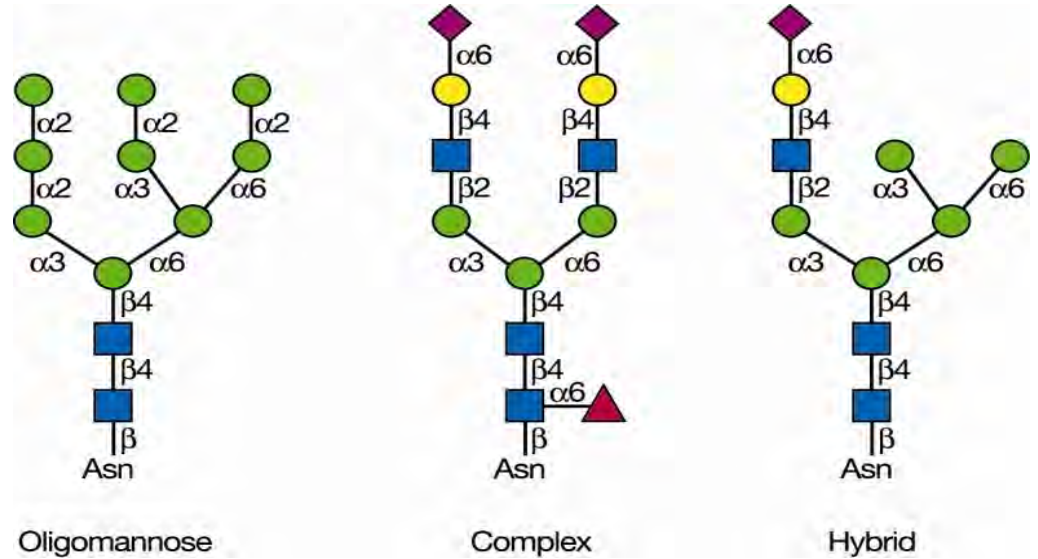


What about bifidobacterial growth on milk glycoproteins?



N-linked exp.
Lactoferrin
Immunoglobulins

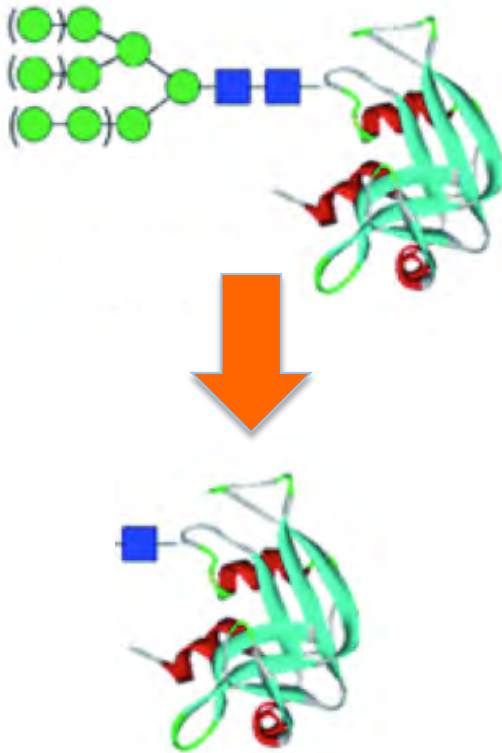
O-linked exp.
Caseins (K)



What about bifidobacterial growth on milk glycoproteins?



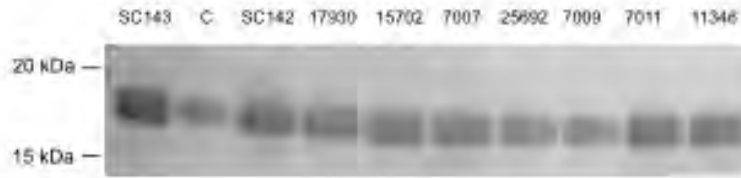
RNaseB as proxy
N-linked glycan



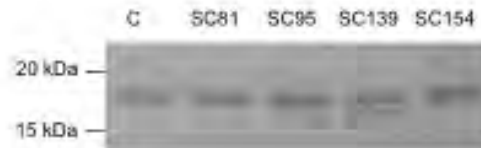
B. longum



B. infantis



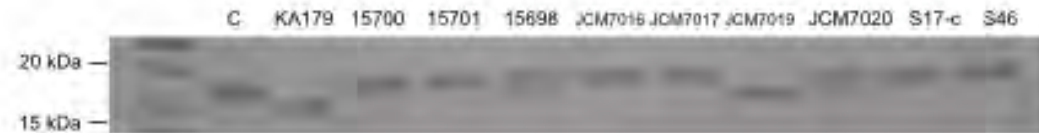
B. breve



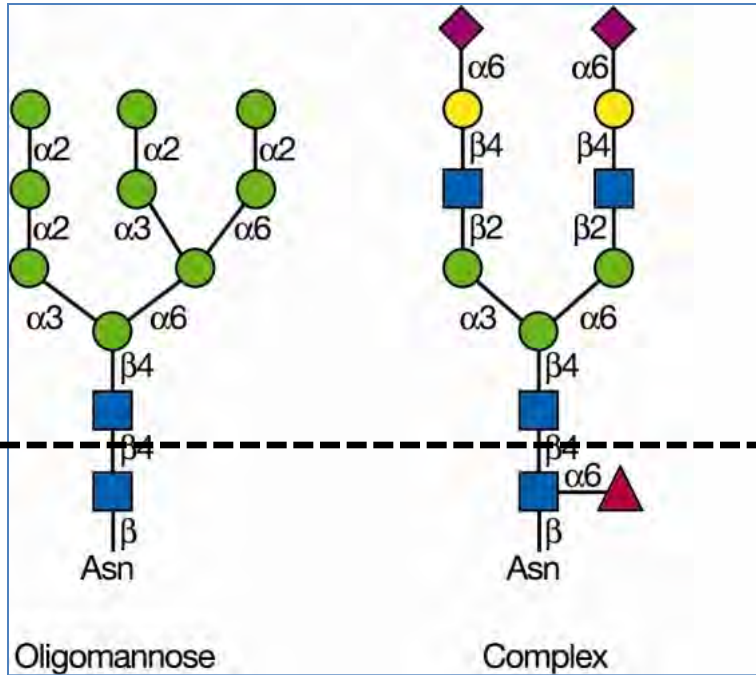
B. breve



B. breve



Endoglycosidase genes in bifidobacteria



Endo-**beta**-N-acetylglucosaminidases

EndoBI-1 and EndoBI-2 active on all N-linked milk glycoproteins

GH18a (EndoBI-1)

- B. infantis JCM11346
- B. infantis JCM7007
- B. infantis ATCC17930
- B. infantis ATCC15702
- B. infantis ATCC15697
- B. infantis JCM7009
- B. infantis JCM7011

EndoE(alpha subunit)

GH18b (EndoBI-2)

- B. infantis 157F
- B. longum SC706
- B. longum SC116
- B. longum SC630
- B. breve SC559
- B. infantis SC142
- B. infantis SC143

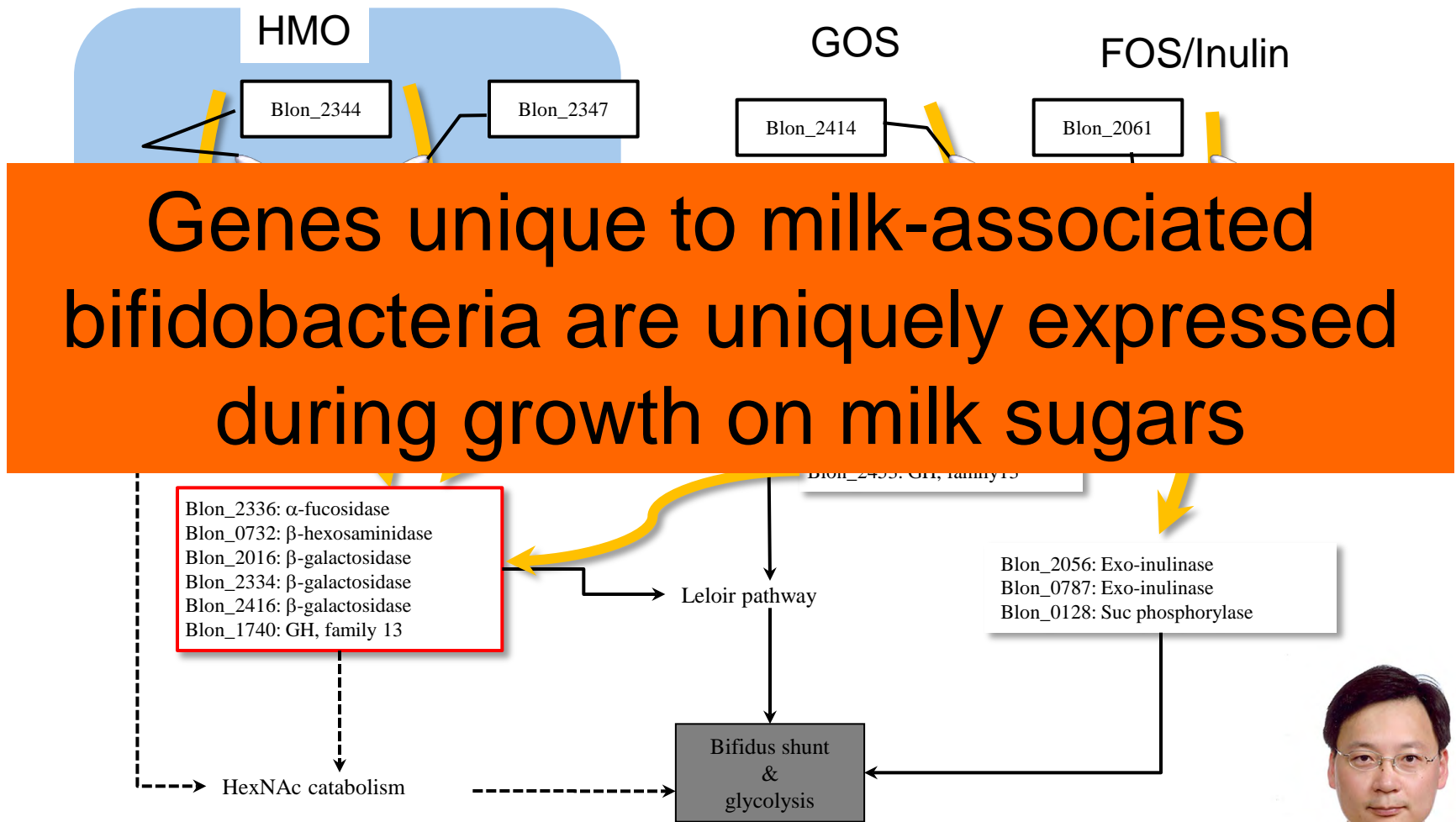
EndoD

GH85 (EndoBB)

- B. breve UCC2003
- B. breve JCM1273
- B. longum DJO10A
- B. breve JCM7019
- B. breve JCM7020
- B. breve KA179
- B. breve SC139
- B. breve SC506
- B. breve SC568
- B. breve SC95

0.2

Whole cell proteomics of *B. infantis* grown on different prebiotic sugars



Jae Han Kim

Milk Bioactives Project activities

Human milk research

Translation

Translation

Translation

Bovine milk research

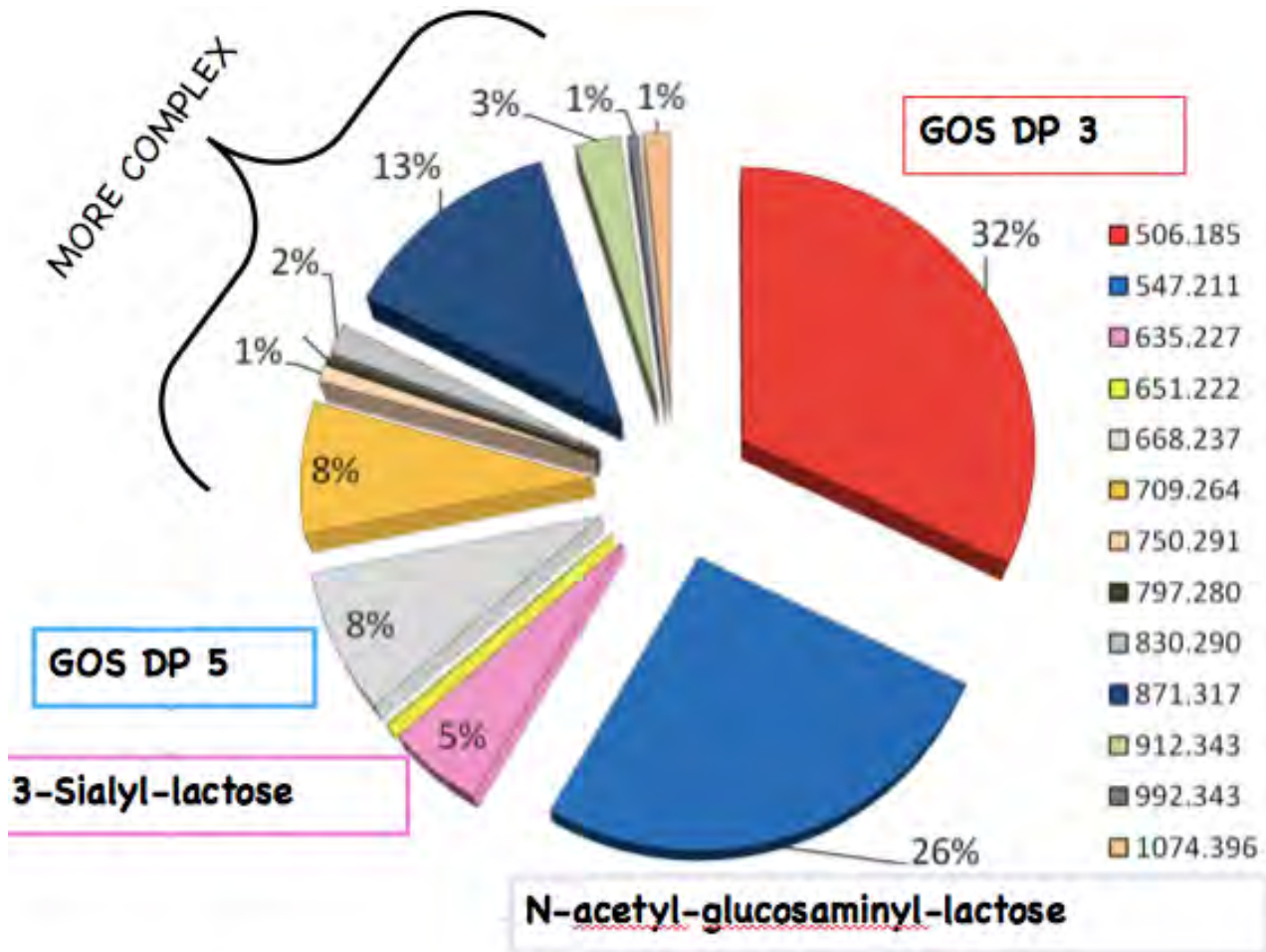
Prebiotic milk oligosaccharides

Glycoproteins,
glycopeptides,
glycolipids

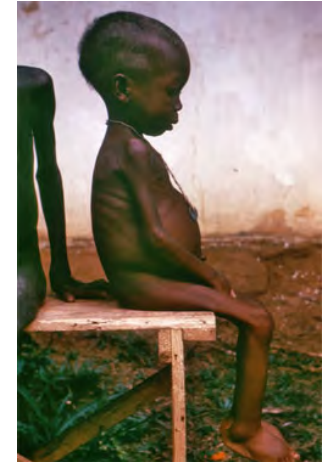
Milk-enhanced
Probiotics
(bifidobacteria)

Milk processing
enzymes

Why permeate oligosaccharides



BMMI Project



**BILL & MELINDA
GATES foundation**



Jeff Gordon Wash U

Nutrition Facts

Serving Size 1 cup (228g)
Servings Per Container 2

Amount Per Serving	
Calories 250	Calories from Fat 110
% Daily Value*	
Total Fat 12g	18%
Saturated Fat 3g	15%
<i>Trans</i> Fat 3g	
Cholesterol 30mg	10%
Sodium 470mg	20%
Total Carbohydrate 31g	10%
Dietary Fiber 0g	0%
Sugars 5g	
Protein 5g	
Vitamin A	4%
Vitamin C	2%
Calcium	20%
Iron	4%

* Percent Daily Values are based on a 2,000 calorie diet. Your Daily Values may be higher or lower depending on your calorie needs.

	Calories:	2,000	2,500
Total Fat	Less than	65g	80g
Sat Fat	Less than	20g	25g
Cholesterol	Less than	300mg	300mg
Sodium	Less than	2,400mg	2,400mg
Total Carbohydrate		300g	375g
Dietary Fiber		25g	30g

“In the 20th century we determined what food is...”



Bruce German
UCD Food Science

...but in the 21st century we need to determine what food does.”

