

# **The Integration of ENCODE into the Study of the Complexity of Cancer Susceptibility**

**Stephen J. Chanock, M.D.**

**Director, Division of Cancer Epidemiology & Genetics**

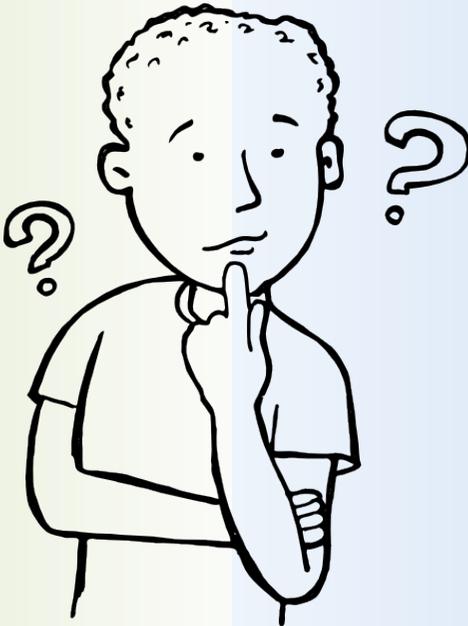
**July 1, 2015**

# Etiology of Cancer

## Environment



## Lifestyle



## Genetic Susceptibility



Mutations

SNPs

STR's

Chromosomes

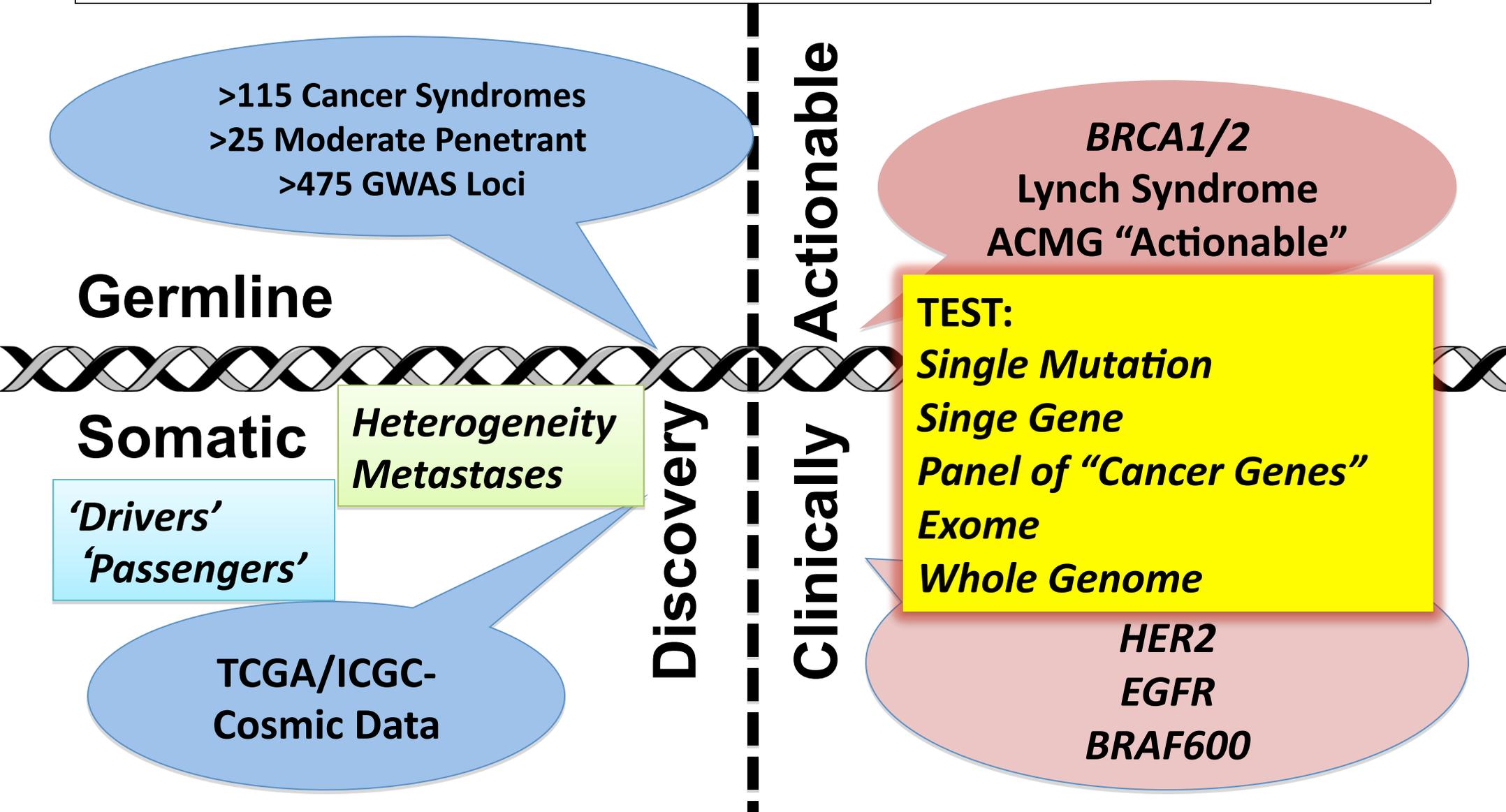
(Epigenetics)

100%  
"Triggers"

"Chance"

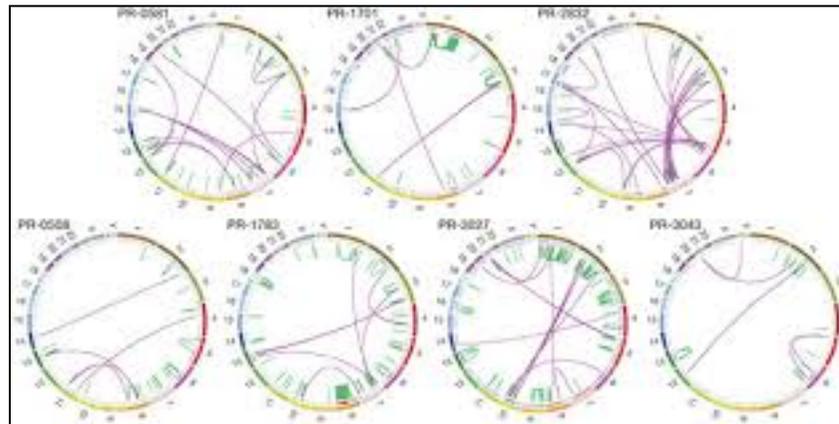
100%  
"Set Point"

# Cancer Genomics: 4 Spaces



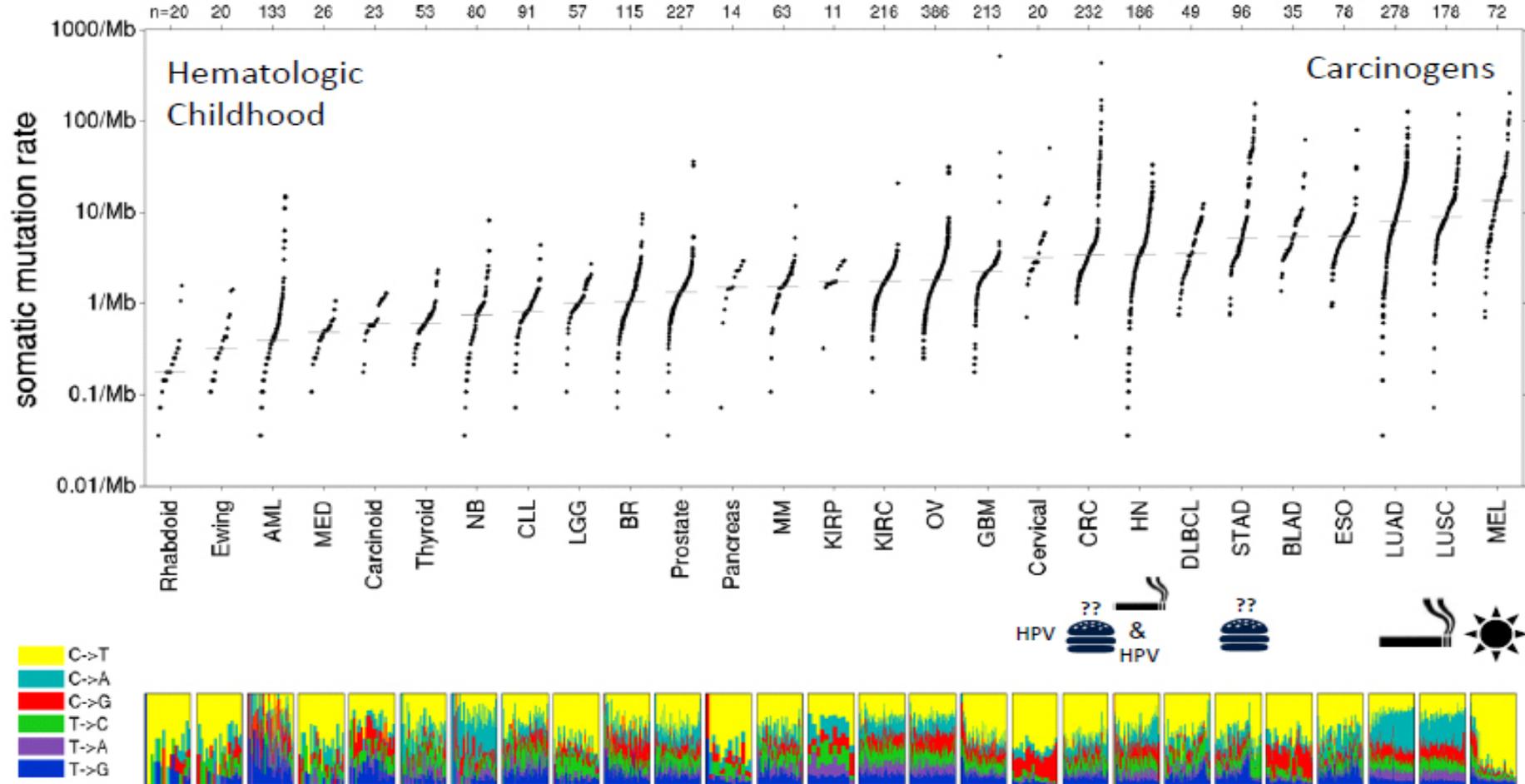
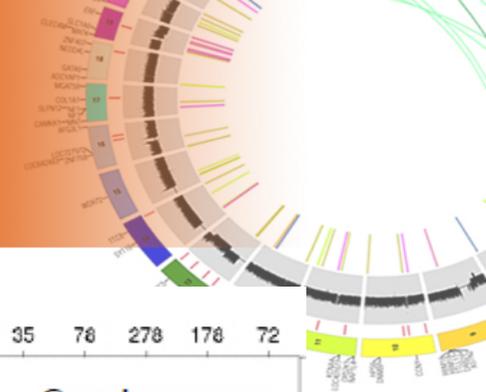
# What Happens When There is More than One Genome?

## Challenge of Cancer Genomics



# TCGA: Driven by the Numbers

*But not yet validated in the laboratory....*



Value of Frequency in Generating Hypotheses  
 But, *further* laboratory work is needed...

# Evidence for Heritability of Cancer

**1866**

Broca observed heritability based on familial breast cancer



**Interim**

Twin/Family/Sibling studies...

**1969**

Li-Fraumeni observed familial clustering (*TP53*)



**1971**

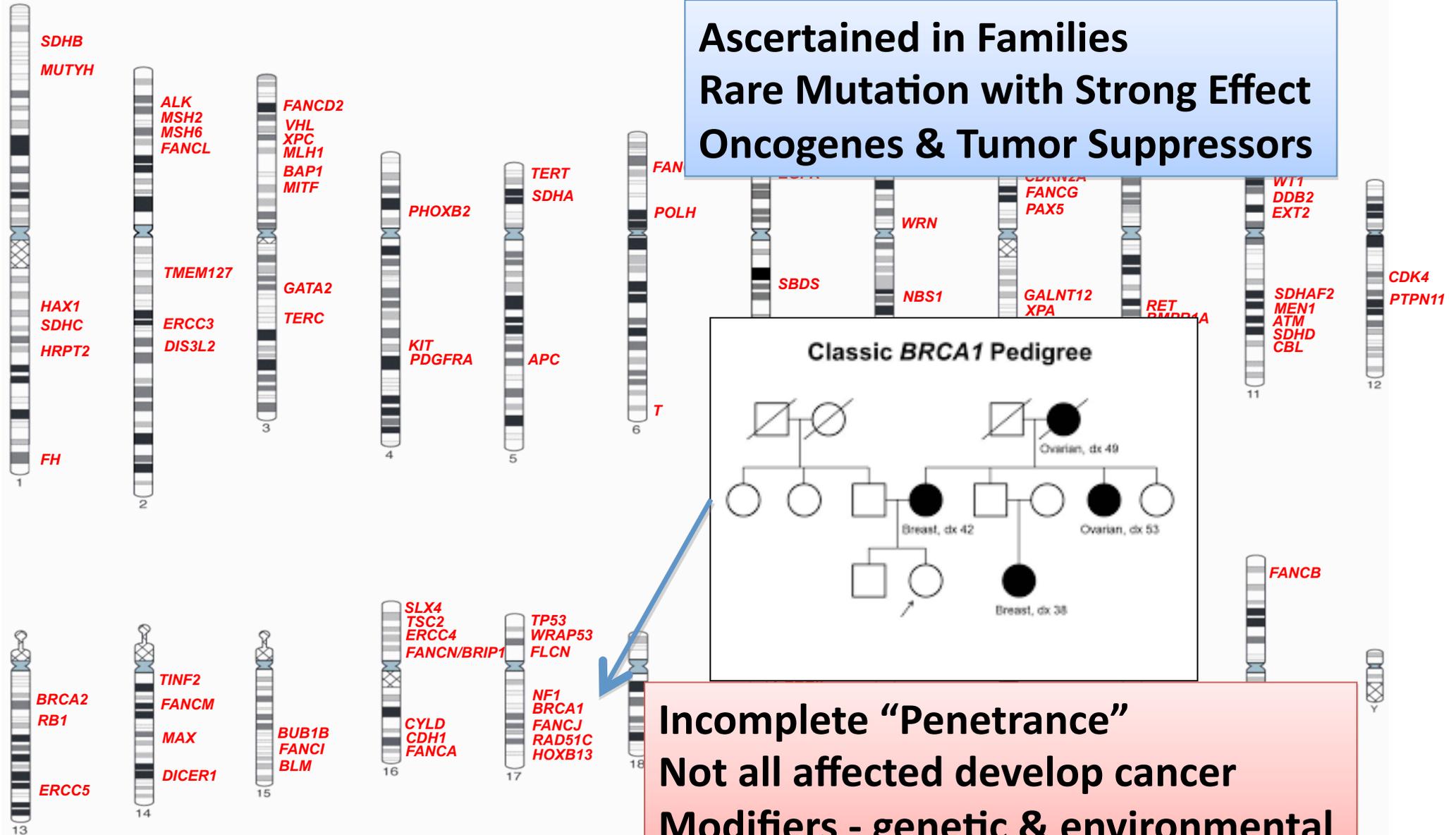
Knudson postulated “two-hit” hypothesis for retinoblastoma

**1991**

Positional cloning of a familial breast cancer gene (*BRCA1*)



# >115 Genes Mutated in Cancer Susceptibility Syndromes



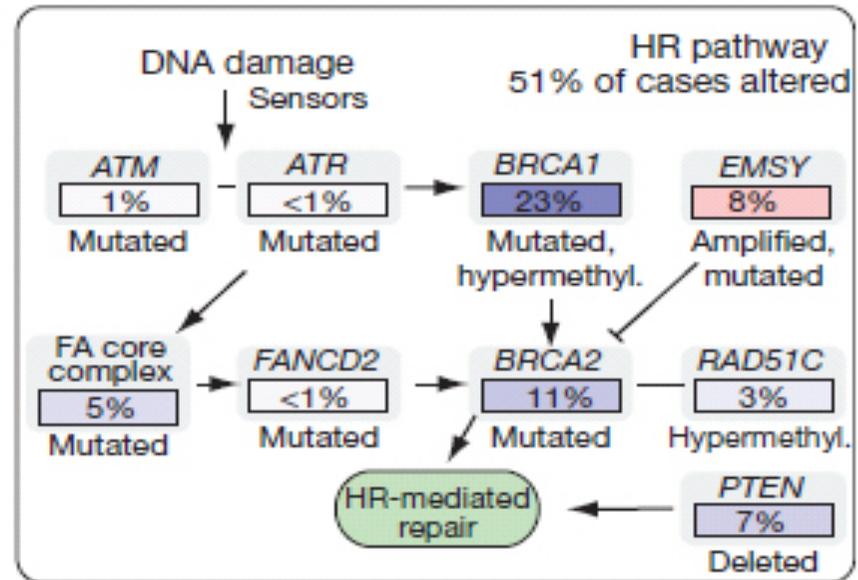
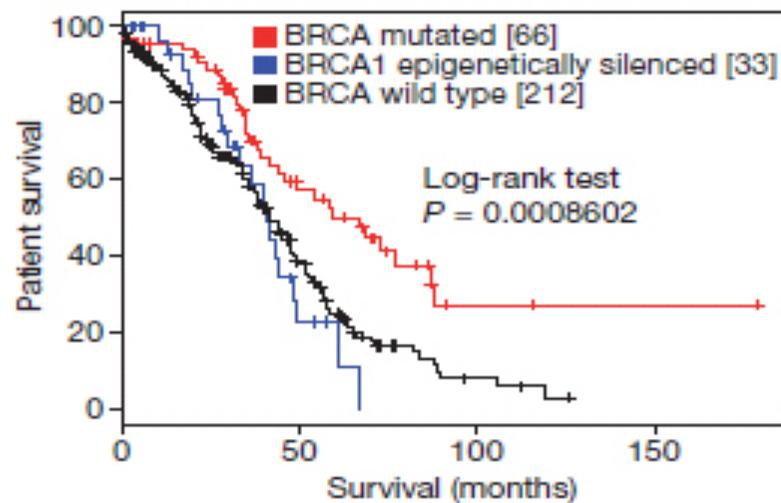
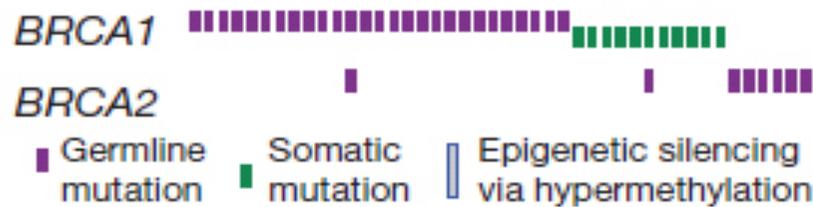
# TCGA: Lessons Learned from the Data

## Survival Analyses

### Impact of germline or somatic mutations



BRCA altered cases,  $N = 103$  (33%)



# High Penetrance Mutations & Somatic Alterations

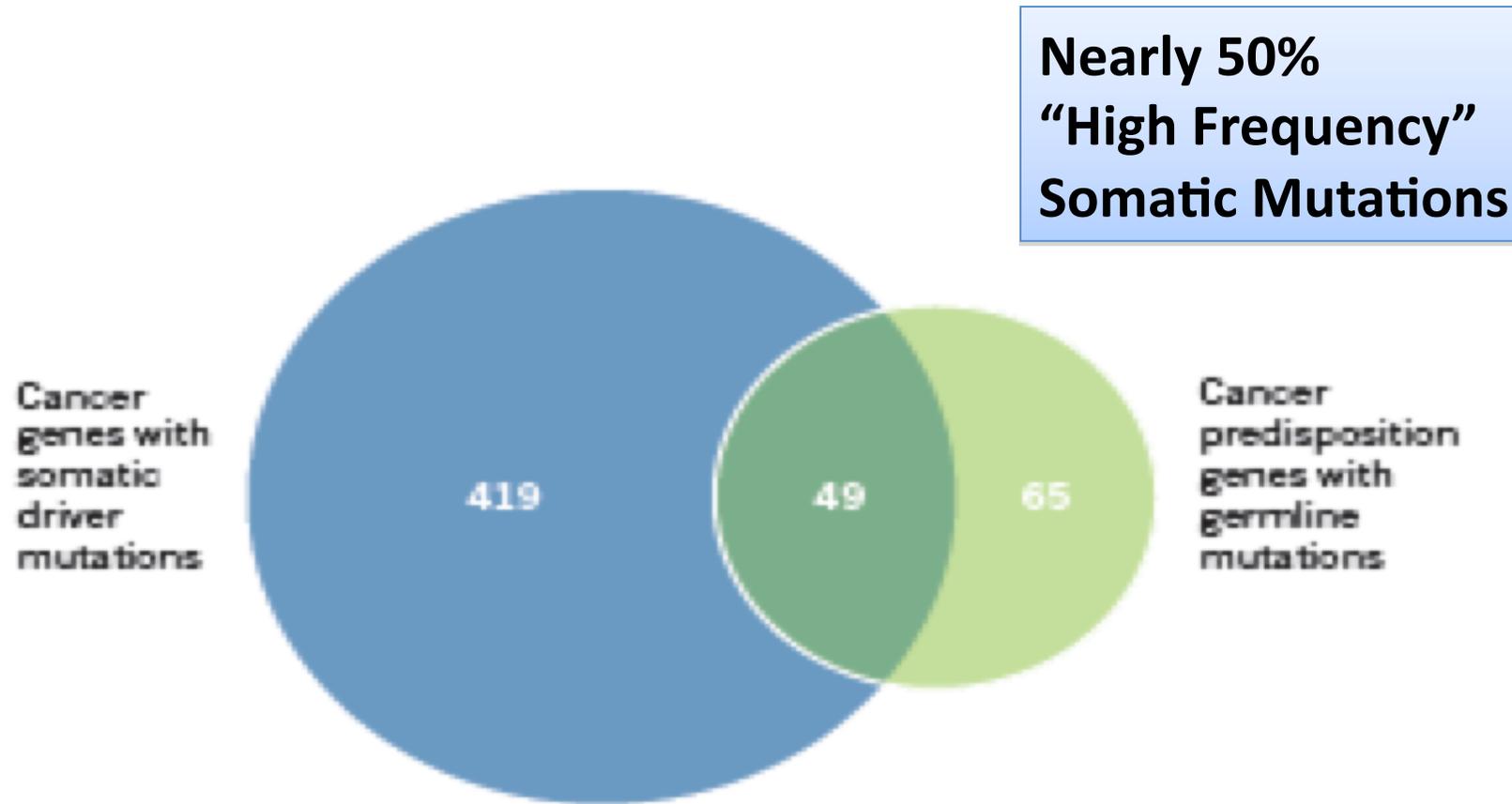


Figure 3 | Overlap between somatically mutated cancer genes and cancer predisposition genes (CPGs). 468 genes with somatic driver mutations in cancers are recorded in the COSMIC database of which 49 are also included within the 114 CPGs.

# Search for Common Variants in Complex Diseases

## Reproducible Technology



## SNP Microarray Chip

>5 M genotyped SNPs across genome

>30 M imputed SNPs across genome

High Concordance > 99.5%/assay

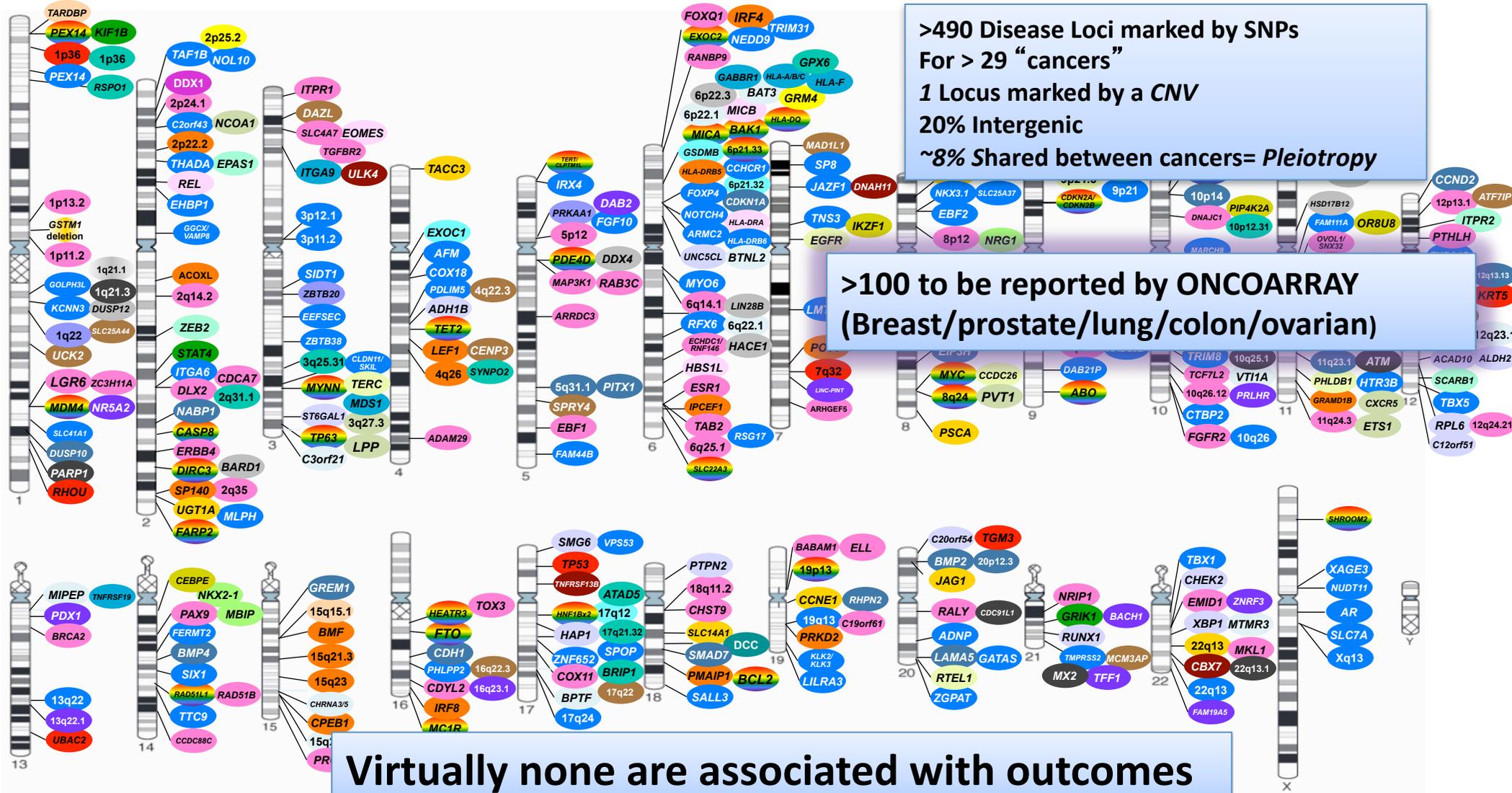
'Markers' across the genome

Commitment to Mapping

***Creates a multiple testing problem***



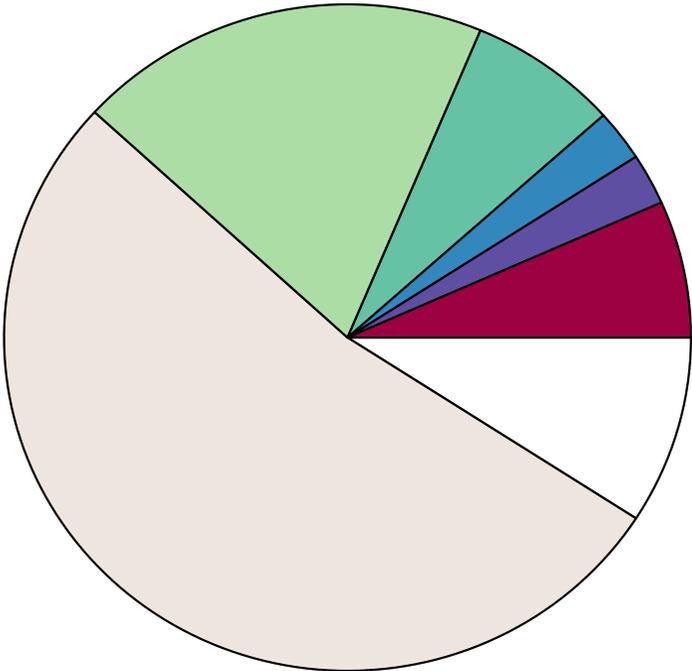
# Published Cancer GWAS Etiology Hits: July 2015



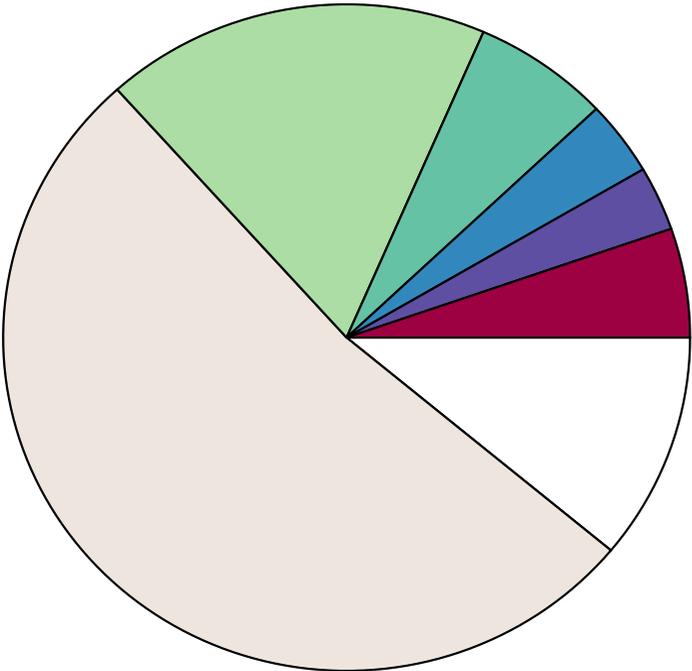
- 13 Basal Cell
- 14 Bladder
- 85 Breast
- 7 Cervical
- 31 CLL
- 31 Colorectal
- 1 Endometrial
- 17 Esophageal Sq
- 3 Ewing Sarcoma
- 1 Gallbladder
- 4 Gastric
- 10 Glioma
- 8 Hodgkins
- 5 Kidney
- 7 Liver
- 20 Lung
- 14 Melanoma
- 31 Multiple
- 6 Multiple Myeloma
- 11 Nasopharyngeal
- 9 Neuroblastoma
- 13 Non-Hodgkin
- 2 Osteosarcoma
- 16 Ovary
- 14 Pancreas
- 9 Pediatric ALL
- 99 Prostate
- 21 Testicular
- 5 Thyroid
- 3 Wilms

# GWAS Signals & Somatic Mutations: No Strong Correlation Redundant Pathways- 'NOT Drivers'

GWAS Genes



Permutation Genes



Number of Mutations (per 100 individuals)

□ 0	□ 0-1	□ 1-2	□ 2-3	□ 3-4	□ 4-5	□ 5<
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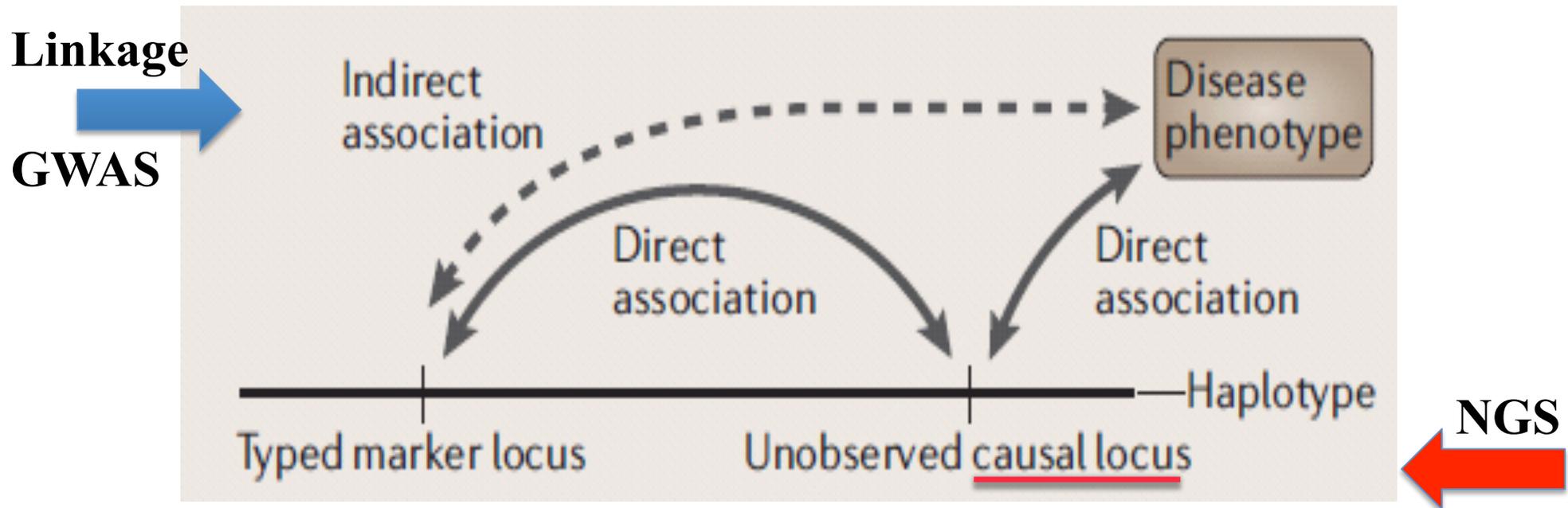


*M Machiela in Revision*

# Interpretation:

## Correlation does not imply causation

GWAS designs provide no mechanism to distinguish statistical association from causation



# Can we use ENCODE to prioritize SNPs for follow-up?

OPEN ACCESS Freely available online



## GPA: A Statistical Approach to Prioritizing GWAS Results by Integrating Pleiotropy and Annotation



Dongjun Chung<sup>1,2\*</sup>, Can Yang<sup>1,3,4\*</sup>, Cong Li<sup>5</sup>, Joel Gelernter<sup>3,6,7,8</sup>, Hongyu Zhao<sup>1,5,7,9\*</sup>

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### Abstract

Results from Genome-Wide Association Studies (GWAS) have shown that complex diseases are often affected by many genetic variants with small or moderate effects. Identifications of these risk variants remain a very challenging problem. There is a need to develop more powerful statistical methods to leverage available information to improve upon traditional approaches that focus on a single GWAS dataset without incorporating additional data. In this paper, we propose a novel statistical approach, GPA (Genetic analysis incorporating Pleiotropy and Annotation), to increase statistical power to identify risk variants through joint analysis of multiple GWAS data sets and annotation information because: (1) accumulating evidence suggests that different complex diseases share common risk bases, i.e., pleiotropy; and (2) functionally annotated variants have been consistently demonstrated to be enriched among GWAS hits. GPA can integrate multiple GWAS datasets and functional annotations to seek association signals, and it can also perform hypothesis testing to test the presence of pleiotropy and enrichment of functional annotation. Statistical inference of the model parameters and SNP ranking is achieved through an EM algorithm that can handle genome-wide markers efficiently. When we applied GPA to jointly analyze five psychiatric disorders with annotation information, not only did GPA identify many weak signals missed by the traditional single phenotype analysis, but it also revealed relationships in the genetic architecture of these disorders. Using our hypothesis testing framework, statistically significant pleiotropic effects were detected among these psychiatric disorders, and the markers annotated in the central nervous system genes and eQTLs from the Genotype-Tissue Expression (GTEx) database were significantly enriched. We also applied GPA to a bladder cancer GWAS data set with the ENCODE database data from T5 cell lines. GPA was able to detect cell lines that are biologically more relevant to bladder cancer. The R implementation of GPA is currently available at <http://dongjunchung.github.io/GPA/>.

**Citation:** Chung D, Yang C, Li C, Gelernter J, Zhao H (2014) GPA: A Statistical Approach to Prioritizing GWAS Results by Integrating Pleiotropy and Annotation. *PLoS Genet* 10(11): e1004787. doi:10.1371/journal.pgen.1004787

**Editor:** Hua Tang, Stanford University, United States of America

**Received:** February 17, 2014; **Accepted:** September 29, 2014; **Published:** November 13, 2014

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**Funding:** This study was supported by National Institutes of Health grants RC2 DA028909, R01 DA12690, R01 DA12849, R01 DA18432, R01 AA11330, R01 AA01755, P30 AA12870, R01 GM95007, R01 DA030976, UL1 RR024139, and the VA Cooperative Studies Program of the Department of Veterans Affairs, Office of Research and Development. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing Interests:** The authors have declared that no competing interests exist.

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† These authors contributed equally to this work.

### Introduction

Hundreds of genome-wide association studies (GWAS) have been conducted to study the genetic bases of complex human traits. As of January, 2014, more than 12,000 single-nucleotide polymorphisms (SNPs) have been reported to be significantly associated with at least one complex trait (see the web resource of GWAS catalog [1] <http://www.genome.gov/gwasstudies/>). Despite of these successes, these significantly associated SNPs can only explain a small portion of genetic contributions to complex traits/diseases [2]. For example, human height is a highly heritable trait whose heritability is estimated to be around 80%, i.e., 80% of variation in height within the same population can be attributed to genetic effects [3]. Based on large-scale GWAS, about 180 SNPs have been reported to be significantly associated with human height [4]. However, these loci together only explain about 5–10%

of variation in height [2,4,5]. This phenomenon is referred to as the “missing heritability” [2,6,7].

Identifying the source of this missing heritability has drawn much attention from researchers, and progress has been made towards explaining the apparent discrepancy. The role of a much greater-than-expected set of common variants (minor allele frequency (MAF)  $\geq 0.01$ ) has been shown to be critical in explaining the phenotypic variance [8]. Instead of only using genome-wide significant SNPs, Yang et al. [9] reported that, by using all genotyped common SNPs, 45% of the variance for human height can be explained. This result suggests that a large proportion of the heritability is not actually missing: given the limited sample size, many individual effects of genetic markers are too weak to pass the genome-wide significance, and thus those variants remain undiscovered. So far, people have found similar genetic architectures for many other complex traits [10], such as

YES  
&  
NO

OPEN ACCESS Freely available online



## Comprehensive Functional Annotation of 77 Prostate Cancer Risk Loci

Dennis J. Hazelett<sup>1\*</sup>, Suhm Kyong Rhee<sup>1</sup>, Malaina Gaddis<sup>2</sup>, Chunli Yan<sup>1</sup>, Daniel L. Lakeland<sup>3</sup>, Simon G. Coetzee<sup>4</sup>, Ellipse/GAME-ON consortium<sup>5\*</sup>, Practical consortium<sup>6\*</sup>, Brian E. Henderson<sup>5</sup>, Houtan Noushmeh<sup>4</sup>, Wendy Cozen<sup>7</sup>, Zsafia Kote-Jarai<sup>6</sup>, Rosalind A. Eeles<sup>6,8</sup>, Douglas F. Easton<sup>9</sup>, Christopher A. Haiman<sup>7</sup>, Wange Lu<sup>10</sup>, Peggy J. Farnham<sup>2</sup>, Gerhard A. Coetzee<sup>1\*</sup>

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### Abstract

Genome-wide association studies (GWAS) have revolutionized the field of cancer genetics, but the causal links between increased genetic risk and onset/progression of disease processes remain to be identified. Here we report the first step in such an endeavor for prostate cancer. We provide a comprehensive annotation of the 77 known risk loci, based upon highly correlated variants in biologically relevant chromatin annotations—we identified 727 such potentially functional SNPs. We also provide a detailed account of possible protein disruption, microRNA target sequence disruption and regulatory response element disruption of all correlated SNPs at  $r^2 \geq 0.5$ . 88% of the 727 SNPs fall within putative enhancers, and many alter critical residues in the response elements of transcription factors known to be involved in prostate biology. We define as risk enhancers those regions with enhancer chromatin biofeatures in prostate-derived cell lines with prostate-cancer correlated SNPs. To aid the identification of these enhancers, we performed genomewide ChIP-seq for H3K27-acetylation, a mark of actively engaged enhancers, as well as the transcription factor TCF7L2. We analyzed in depth three variants in risk enhancers, two of which show significantly altered androgen sensitivity in LNCaP cells. This includes rs4907792, that is in linkage disequilibrium ( $r^2 = 0.91$ ) with an eQTL for NUDT11 (on the X chromosome) in prostate tissue, and rs10486567, the index SNP in intron 3 of the JAZF1 gene on chromosome 7. Rs4907792 is within a critical residue of a strong consensus androgen response element that is interrupted in the protective allele, resulting in a 56% decrease in its androgen sensitivity, whereas rs10486567 affects both NKX3-1 and FOXA-AR motifs where the risk allele results in a 39% increase in basal activity and a 28% fold-increase in androgen stimulated enhancer activity. Identification of such enhancer variants and their potential target genes represents a preliminary step in connecting risk to disease process.

**Citation:** Hazelett DJ, Rhee SK, Gaddis M, Yan C, Lakeland DL, et al. (2014) Comprehensive Functional Annotation of 77 Prostate Cancer Risk Loci. *PLoS Genet* 10(11): e1004102. doi:10.1371/journal.pgen.1004102

**Editor:** Vivian G. Cheung, University of Michigan, United States of America

**Received:** October 1, 2013; **Accepted:** November 14, 2013; **Published:** January 30, 2014

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**Funding:** The work reported here was funded by the National Institutes of Health (NIH) [CA109147, U19CA148537 and U19CA148107 to GAC; ST32CA009320-27 to SGC and NIH/NIDDK USHG000696 (JPF) and Dan and Mazzeo Awards Program (GAC) and ST32CA009320-27 for MG]. The scientific development of this project were in part supported by the Genetic Associations and Mechanisms in Oncology (GAME-ON); a NCI Cancer Post-GWAS Initiative. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing Interests:** The authors have declared that no competing interests exist.

\* Email: hazelett@usc.edu (DJH); coetzee@usc.edu (GAC)

\* Membership of the Ellipse/GAME-ON consortium and the Practical consortium is provided in the Acknowledgments.

### Introduction

The basic goal of research into human genetics is to connect variation at the genetic level with variation in organismal and cellular phenotype. Until recently, inferences about such connections have been limited to the kind associated with heritable disorders and developmental syndromes. Such variations often turn out to be the result of disruptions to protein coding sequences of critical enzymes for an affected pathway. Recent advances in

genomics and medicine have begun to illuminate a sea of variation of a more subtle variety, not always the result of mutation of protein coding sequences. In particular, genome-wide association studies (GWAS) have identified thousands of variants associated with hundreds of disease traits [1]. These variants, typically encoded by single nucleotide polymorphisms (SNPs), are given landmark status and called “index-SNPs” (they are also frequently referred to in the literature as “tag-SNPs”) as the reference for disease or phenotype association in that region. The vast majority

**GWAS Signals**

**“One by One” Investigation**

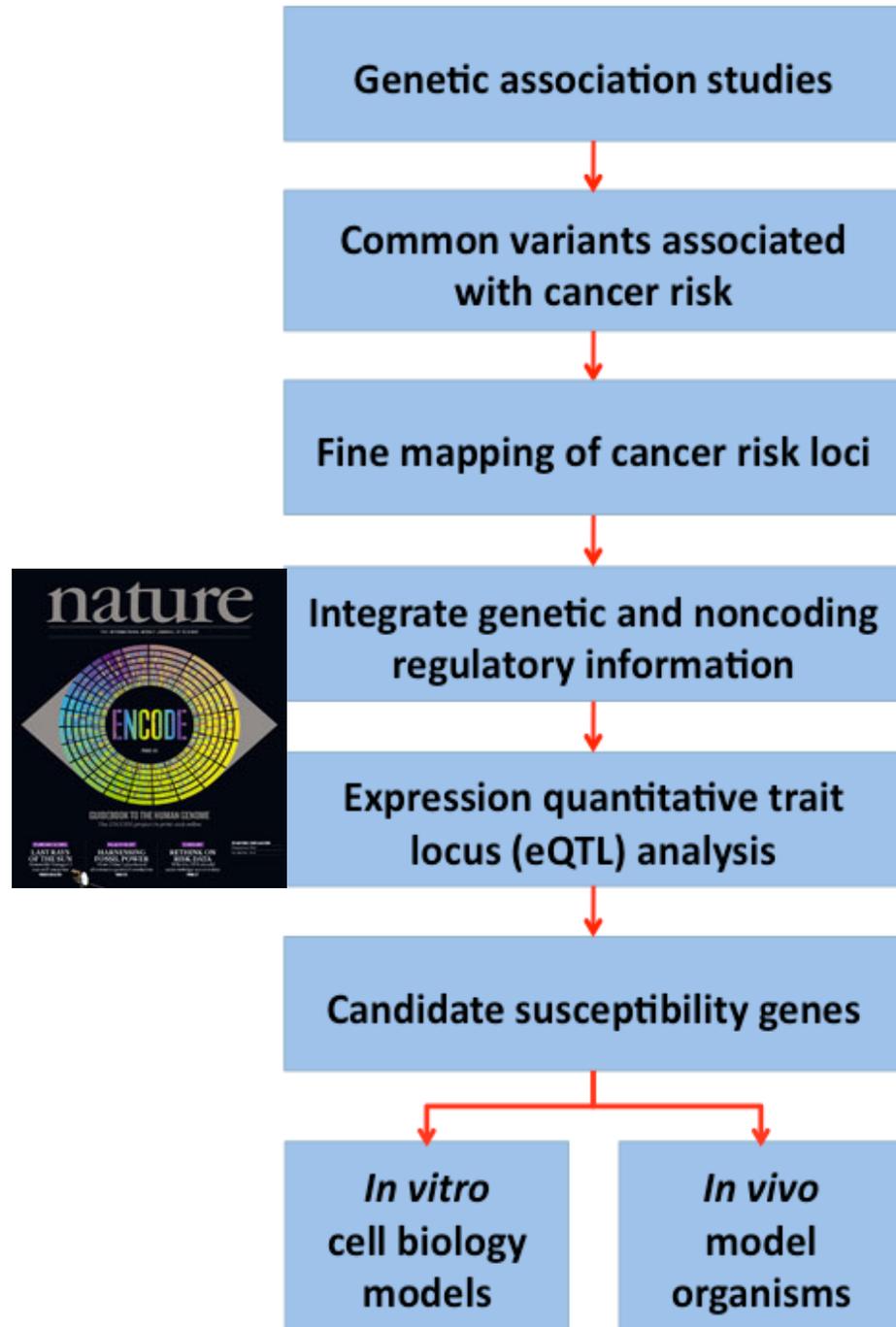
**Insights into Biology**

**Perturbations of Redundant Pathways/Processes**

**Not Causal**

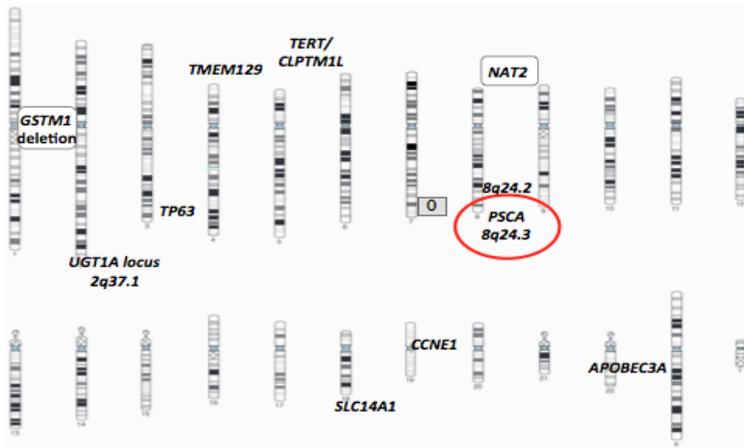
**Instead.....**

**Functional Contribution**

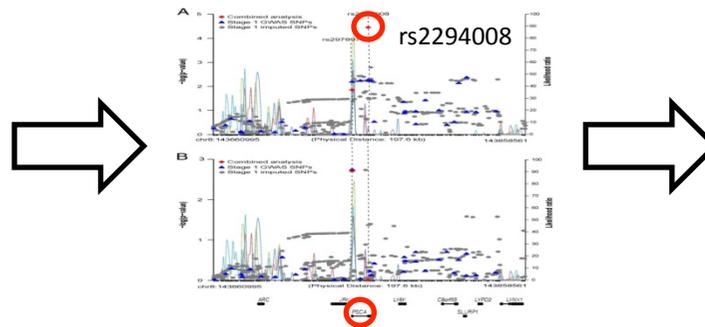


# Bladder Cancer GWAS Discovery → Clinical Trial Target Prostate Stem Cell Antigen (PSCA) PI: M Prokunina-Olsson

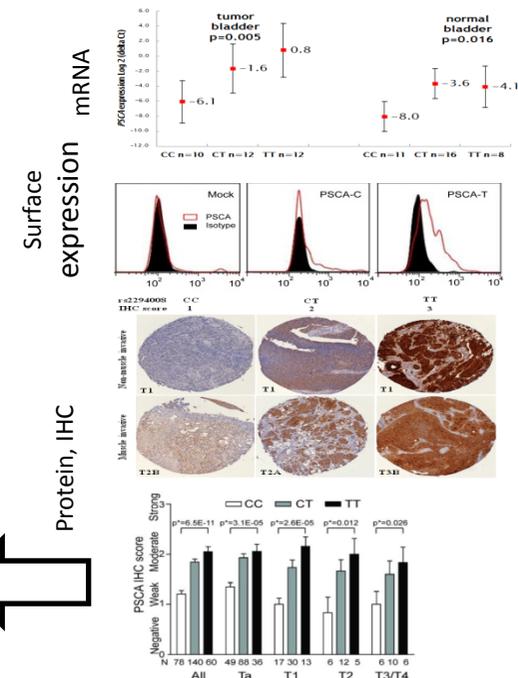
**PSCA, 8q24.3  
Discovered 2009**



**Fine Mapping  
Genotyping & Imputation**



**Functional Studies  
Risk allele T  
↑ mRNA expression**



**Possible Clinical  
Trial**

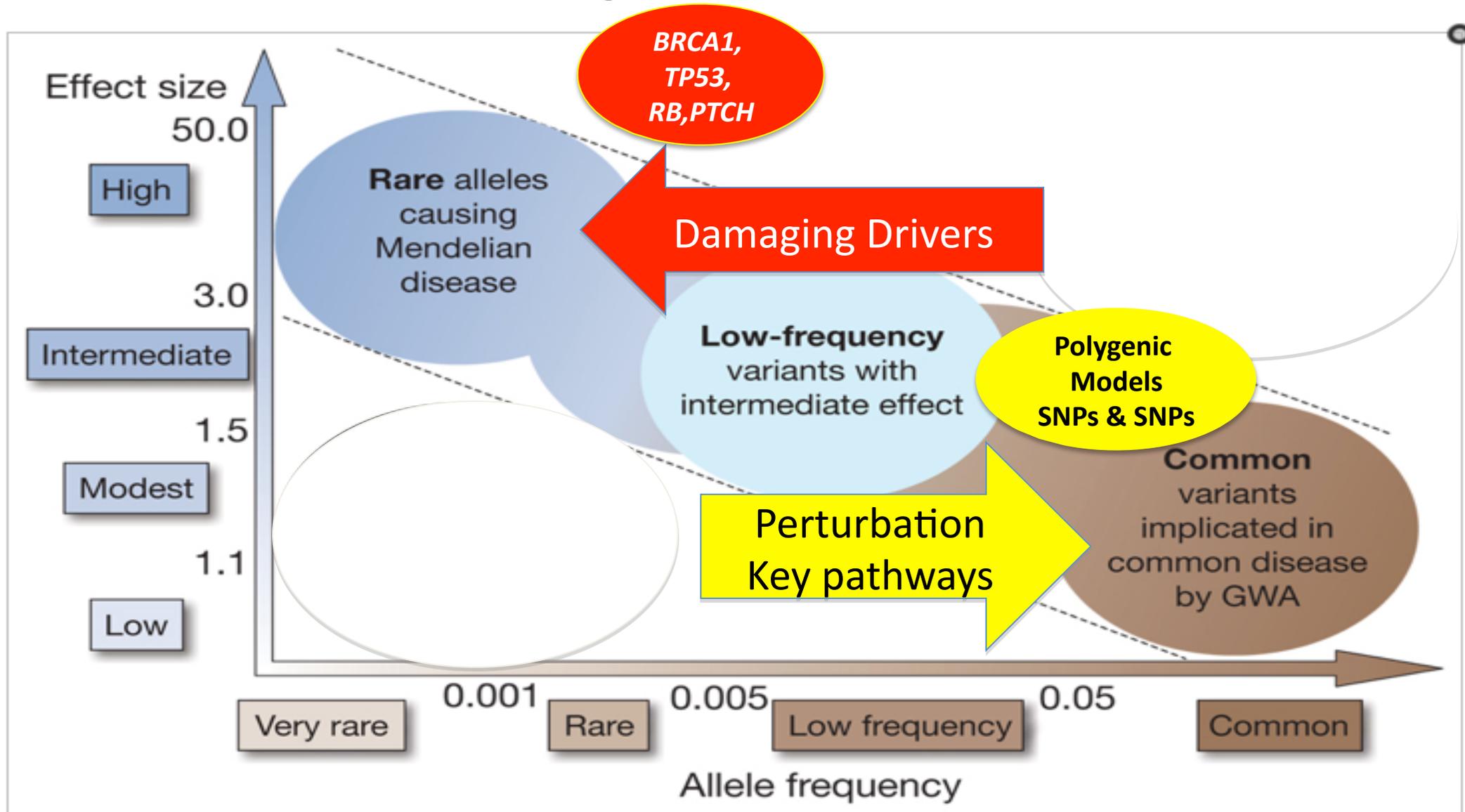
Therapeutic humanized  
anti-PSCA antibody for  
bladder cancer

**Translational application  
rs2294008 predicts PSCA  
expression in tumors  
Anti-PSCA therapy?**

Mila  
Prokunina-  
Olsson



# Architecture of Genetic Susceptibility of Cancer Defining Distinct Spaces



# What Fraction of the Polygenic Component Contributes to Each Cancer?

- 13 cancer GWAS
  - 49,492 cases
  - 34,131 controls (often used in  $> 1$  study)
- Use genotyped SNPs
- **Explains 10-50% of variability on the liability scale**

# Across Cancer Types

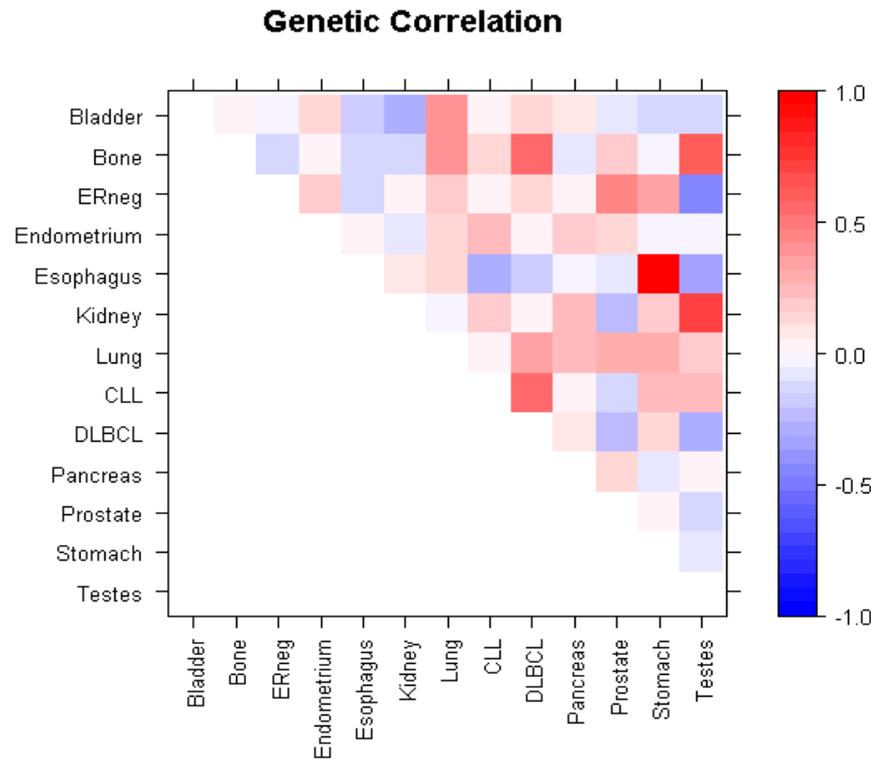
Table 2 Estimates of first-degree familial relative risk from familial registries and GWAS

Cancer	All 1 <sup>st</sup> Degree Relationships	Sweden	Sibling	Iceland <sup>A</sup>	Utah <sup>A</sup>	GWAS
		Parent/Child				
Bladder	1.69 (1.33-2.14)	1.53 (1.16–1.99)	3.30 (1.70–5.78)	1.68 (1.39-2.05)	1.8 (1.4–2.3)	1.37 (1.25-1.50)
Breast (ER-)						1.28 (0.98-1.63)
Endometrium	3.02 (2.33-3.92)	2.85 (2.08–3.82)	3.97 (1.97–7.13)	1.86 (1.31-2.62)	1.4 (1.1–1.8)	1.56 (1.25-1.92)
Esophagus		2.14 (0.77-4.70)		2.09 (1.30-3.31)	1.3 (0.2–10.0)	1.63 <sup>B</sup> (1.27-2.05)
Glioma	1.67 (1.43-1.94)		3.31 (2.08-5.02)	1.41 (0.74-2.40)	2.3 (0.99-4.5)	1.19 (0.91-1.54)
Kidney	1.78 (1.33-2.39)	1.52 (1.06–2.11)	4.52 (2.15–8.35)	2.30 (1.89-2.80)	2.1 (1.3–3.5)	1.54 (1.07-2.13)
Lung						
European	1.70 (1.42-2.05)	1.64 (1.34–2.00)	2.61 (1.29–4.68)	2.00 (1.83-2.16)	2.4 (1.9–3.0)	1.42 (1.28-1.57)
Asian						1.31 <sup>B</sup> (1.16-1.46)
Lymphoma						
CLL	8.5 (6.1-11.7)				6.1 (4.75-7.65)	2.28 (1.86-2.77)
DLBCL	9.8 (3.1-31.0)					1.40 (1.15-1.68)
Osteosarcoma						12.7 (8.27-19.1)
Pancreas		1.68 (1.16–2.35)		2.33 (1.83-2.96)	2.1 (1.3–3.2)	1.35 (1.12-1.62)
Prostate	2.75 (2.32-3.25)	2.71 (2.26–3.22)	4.91 (1.28–12.7)	1.89 (1.75-2.01)	2.1 (1.9–2.2)	1.51 (1.32-1.72)
Stomach	1.99 (1.47-2.71)	1.72 (1.19–2.40)	8.82 (3.50–18.3)	1.90 (1.74-2.05)	2.0 (1.1–3.7)	1.94 <sup>B</sup> (0.95-3.49)
Testes	7.07 (5.34-9.37)	4.31 (2.05–7.95)	8.50 (6.01–11.7)	3.52 (1.18-7.37)	1.8 (0.4–8.6)	3.09 (1.41-6.05)

# Shared Heritability from GWAS

## 13 Distinct Cancers

(49,492 cases and 34,131 shared controls)



### Shared factors:

Some expected

- Testes & Kidney
  - CLL & DLBCL
  - Bladder & Lung (smoking)
- Others not...
- DLBCL & Osteosarcoma

Josh Sampson  
+ 280 co-authors



***Prediction is difficult,  
Especially about the future.***

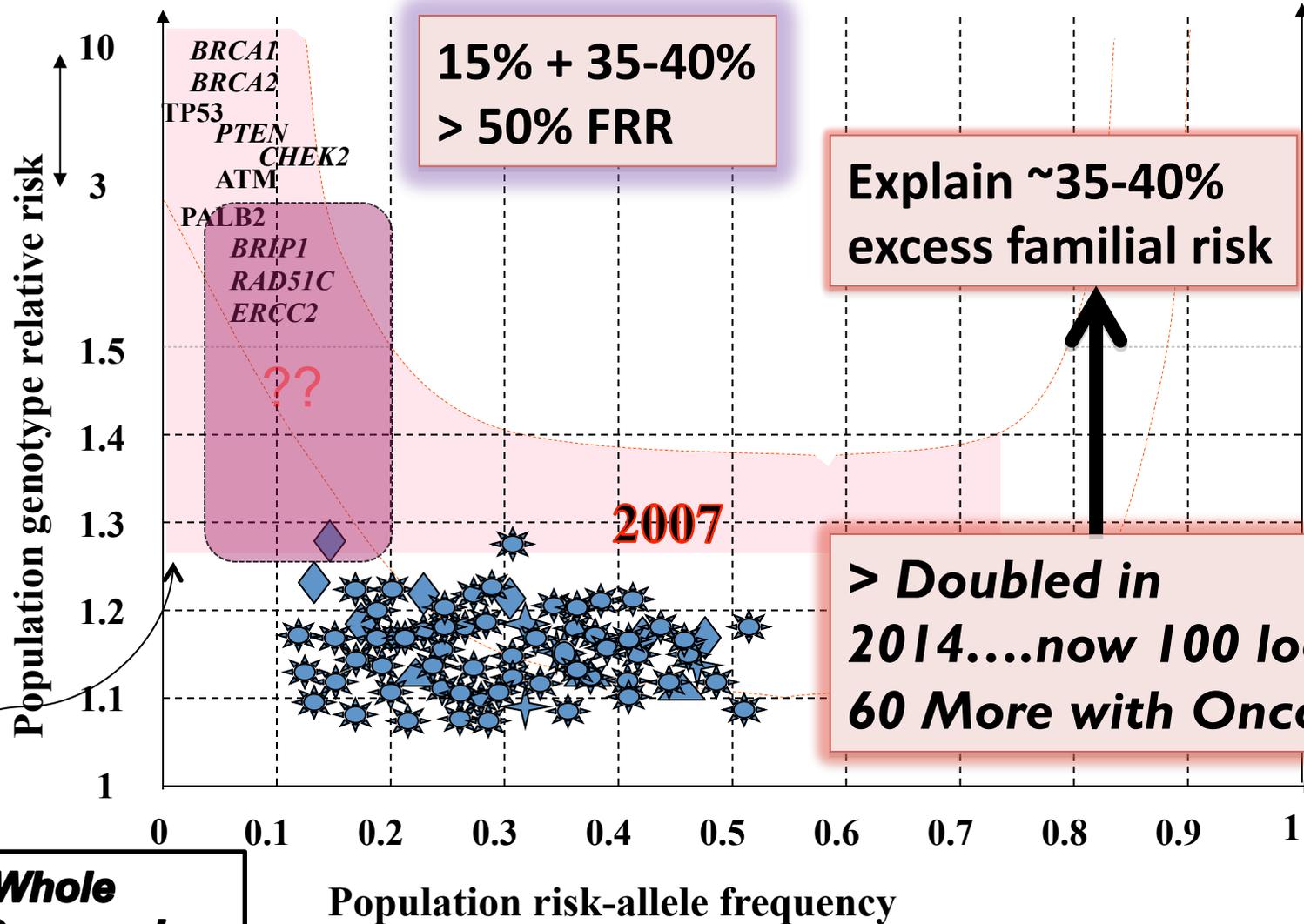
**Yogi Berra**

**Dan Quayle**

**Niels Bohr**

# Genetic Predisposition to Breast Cancer European Population

1994



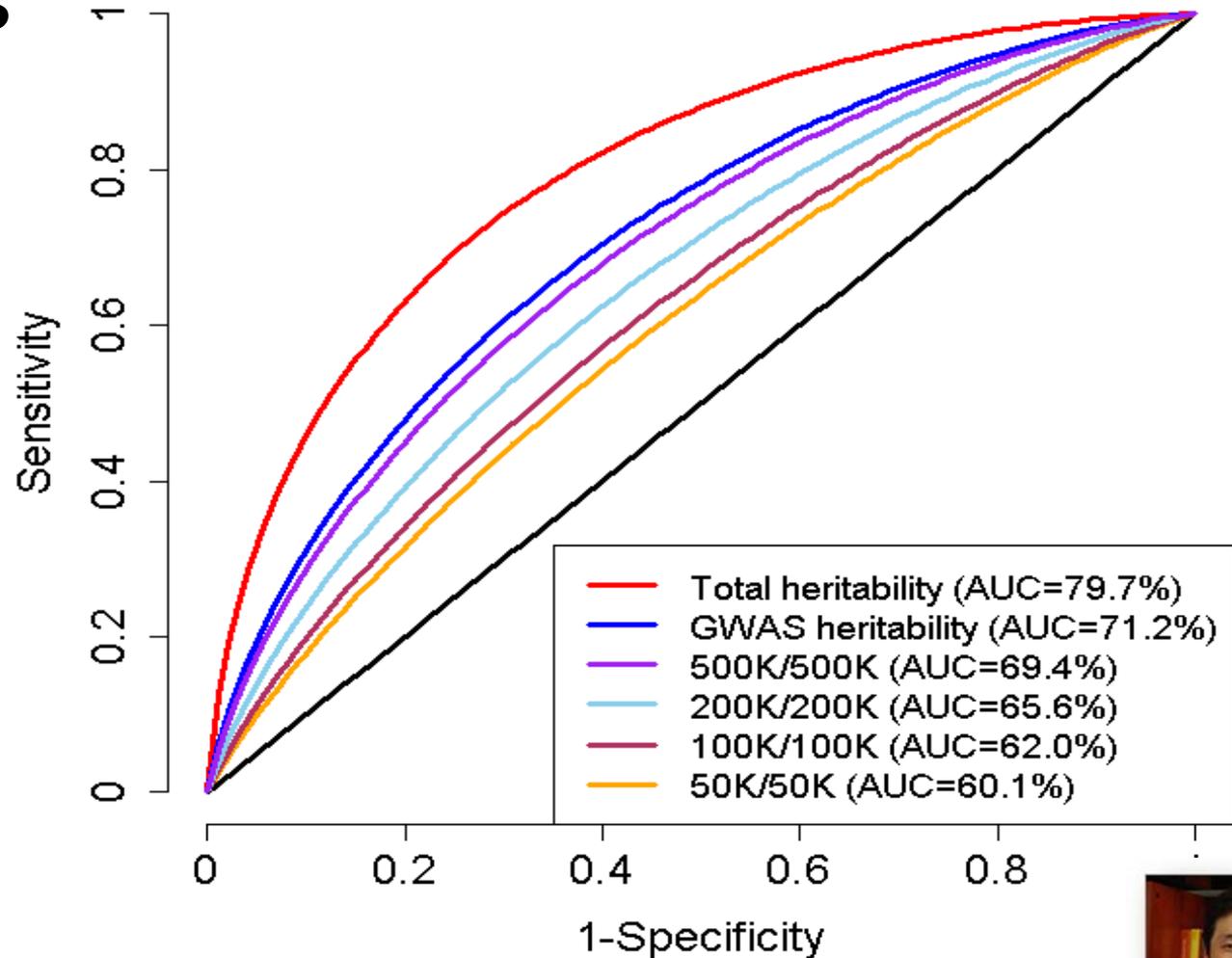
**Exome & Whole  
Genome Sequencing**

# 2015 Projections For General Population

So far, we explain  
~35-40% familial risk

Change in Absolute Risk  
For Screening

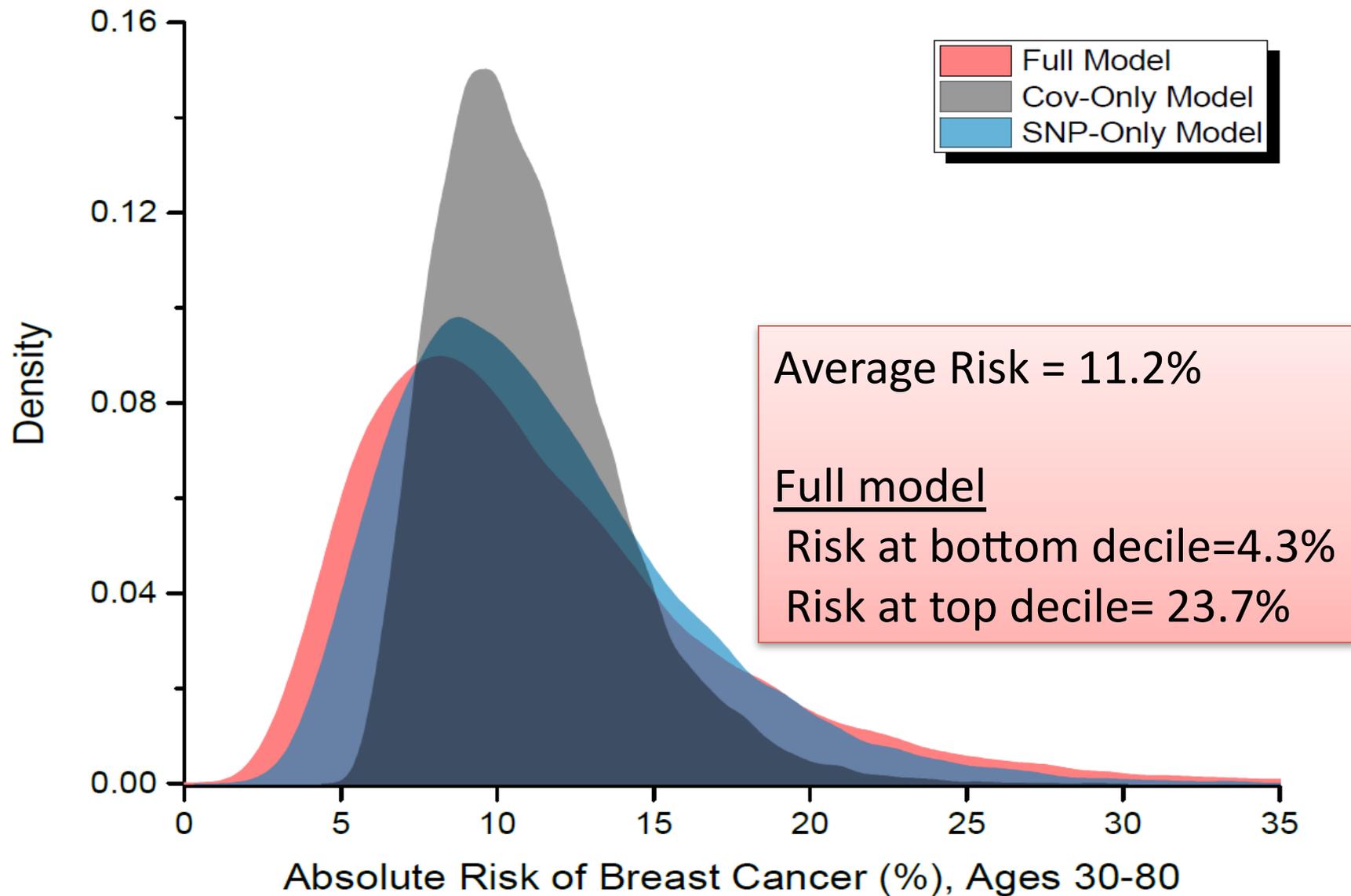
AUC for Breast Cancer



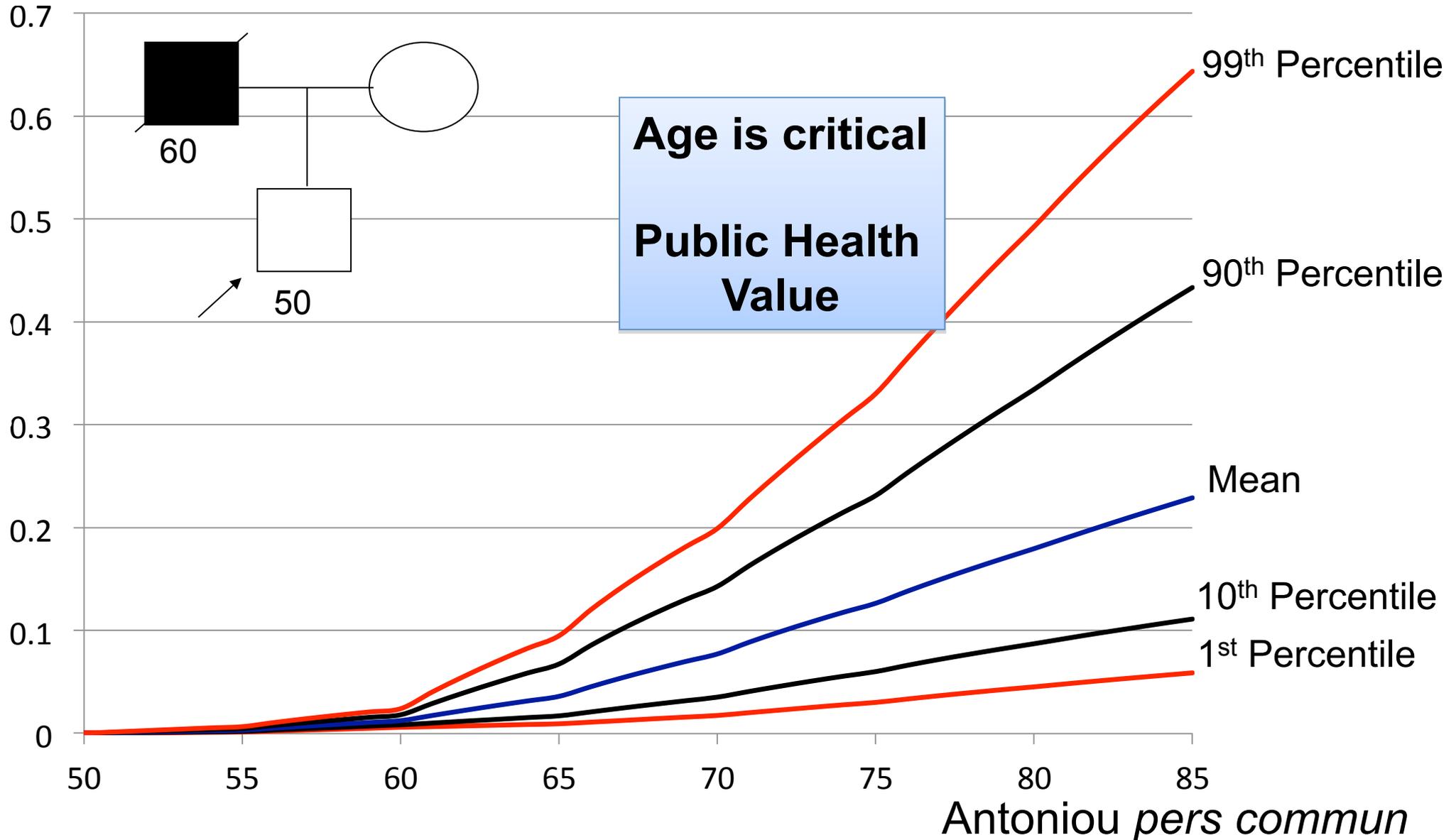
JuHyun Park

- Total heritability corresponds to 2-fold sibling relative risk.
- GWAS Heritability: ~3000 SNPs explain 1.4 fold sibling relative risk

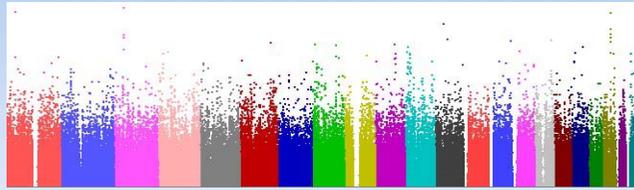
# Projected Distribution of Absolute Lifetime Risk (Age 30-80) of Breast Cancer for US Caucasian Women



# Predicted Prostate Cancer Risk by SNP Profile Distribution (76 SNPs)

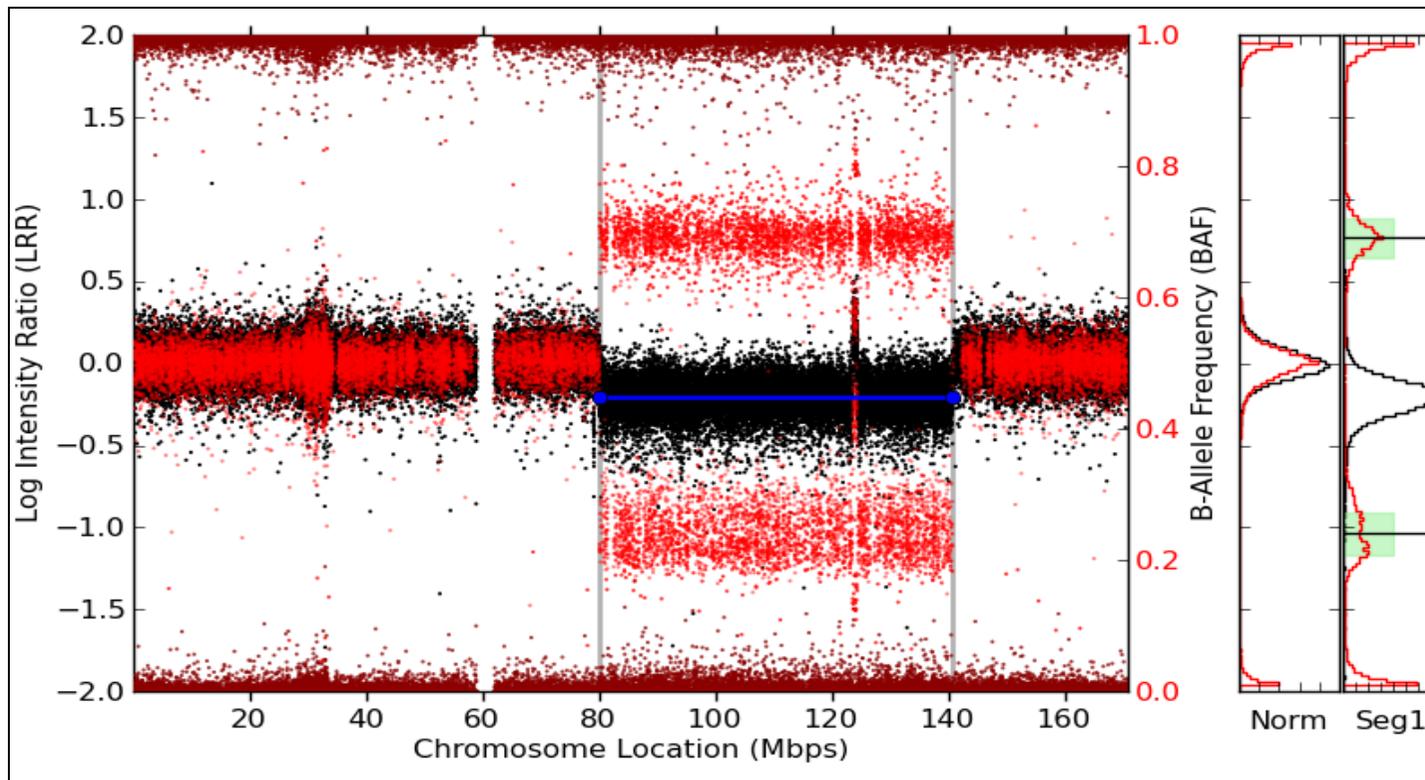


## Unexpected Findings



Genome-wide association studies

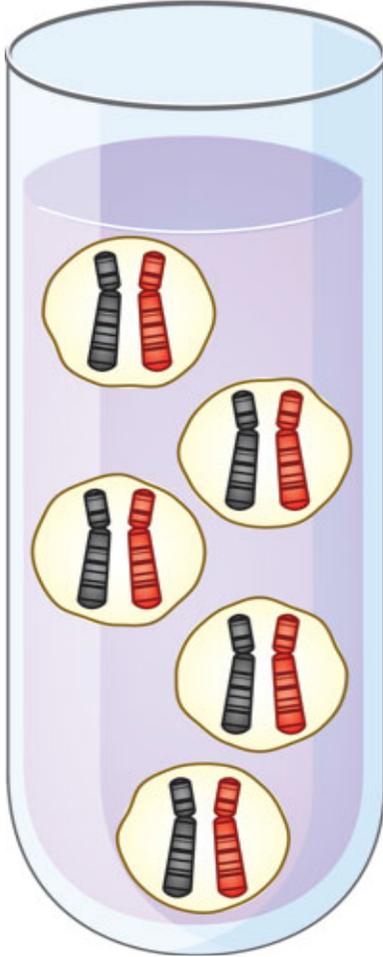
Large chromosomal abnormalities, structural variation, aneuploidy in germline DNA



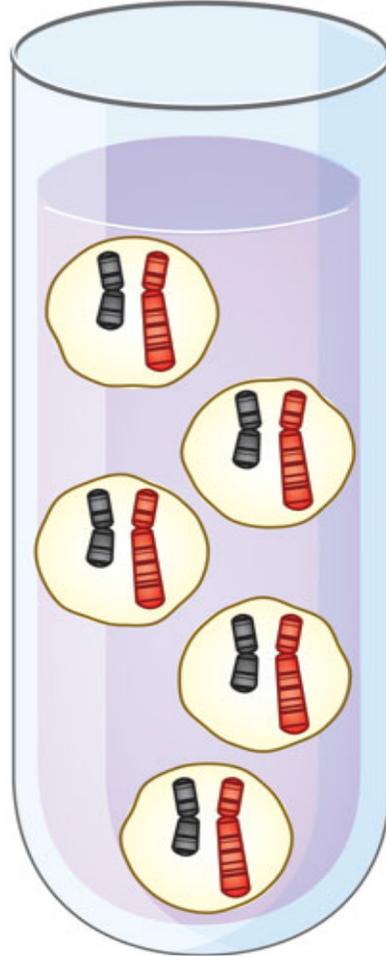
Rodriguez-Santiago *AJHG* 2010  
Jacobs et al *Nature Genetics* 2012  
Laurie et al *Nature Genetics* 2012

# *Somatic Mosaicism- the Dynamic Genome*

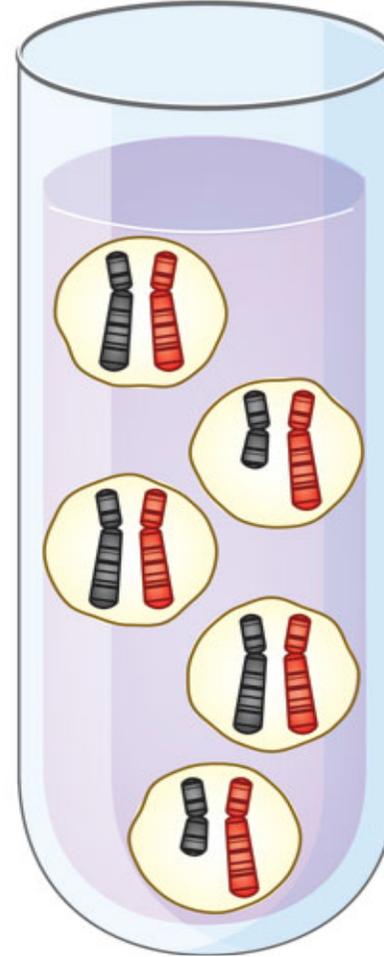
Normal



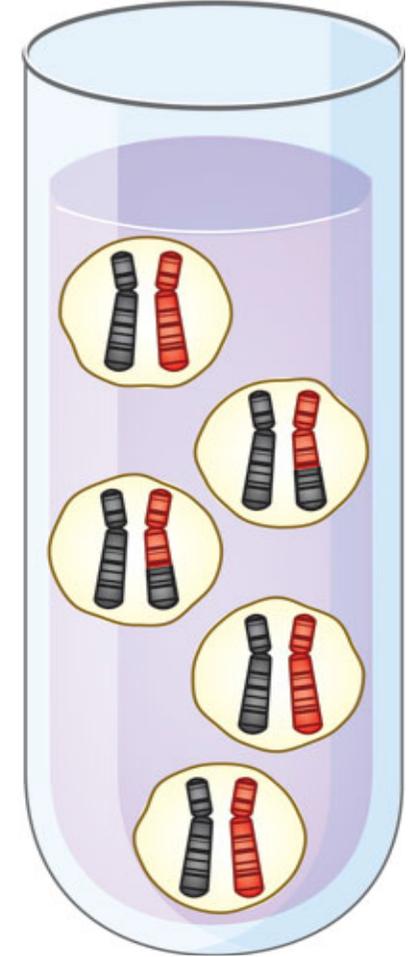
Constitutive copy-loss CNV



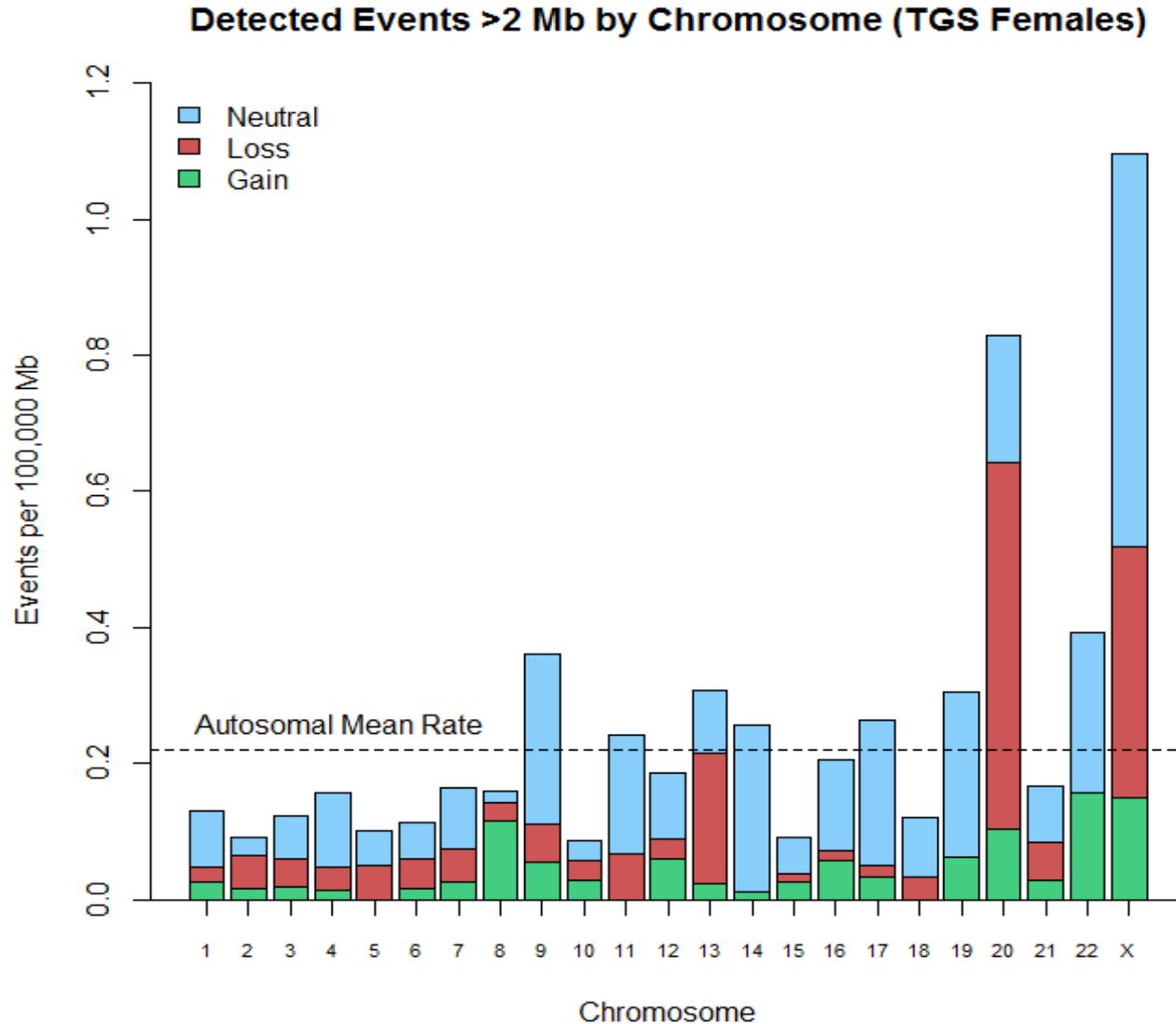
Mosaic copy-loss CNV



Mosaic UPD



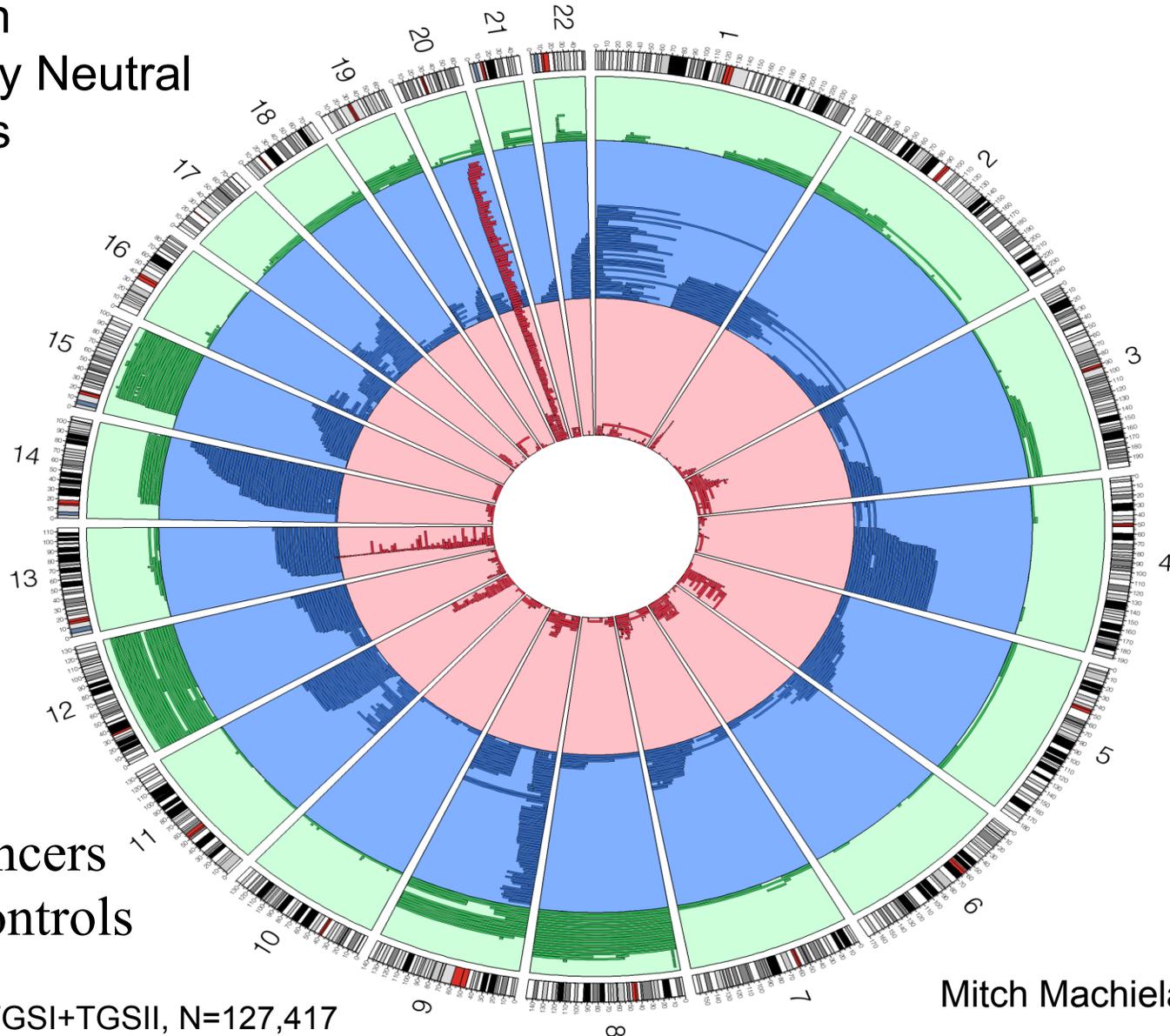
# Rate of Mosaicism by Chromosome: Adjusted for Chromosomal Size



# Combined Sample Detected Events >2Mb

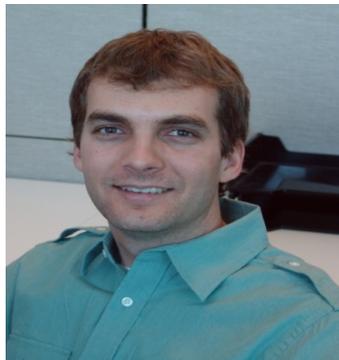
(N=1,330 events in 127,417 individuals)

- Mosaic Gain
- Mosaic Copy Neutral
- Mosaic Loss



Non-Heme Cancers  
Cancer-free Controls

AJHG 2015



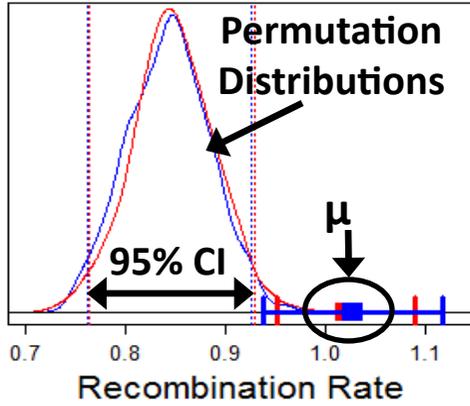
Mitch Machiela

\* Combined GENEVA+TGSI+TGSII, N=127,417

# Breakpoint Analysis of Large Mosaic Regions

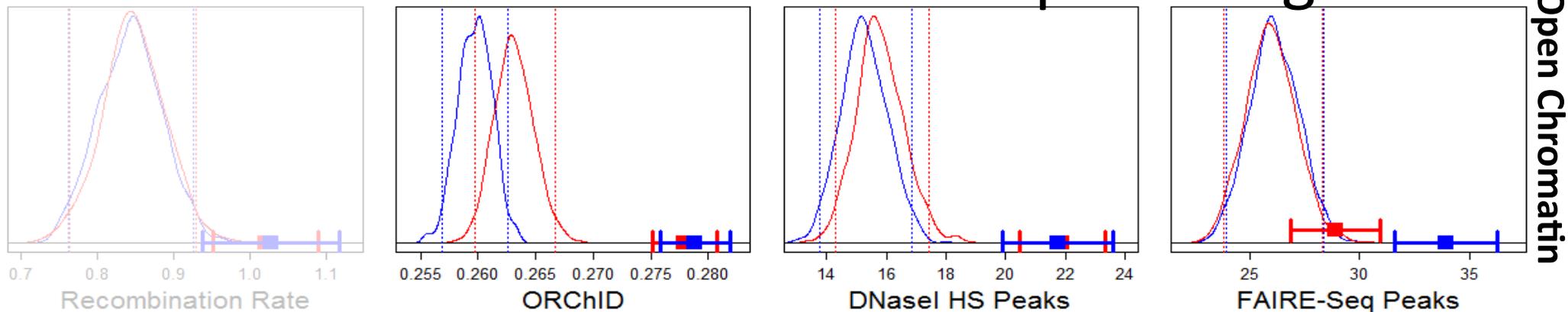
- 688 Interstitial Events
- 543 Telomeric Copy Neutral Events
- Examined
  - 200kb Windows
  - 500 Permutation
- Enrichment of ENCODE elements?

# ENCODE Features around Breakpoint Regions



- 549 Telomeric Copy Neu
- 688 Interstitial Loss

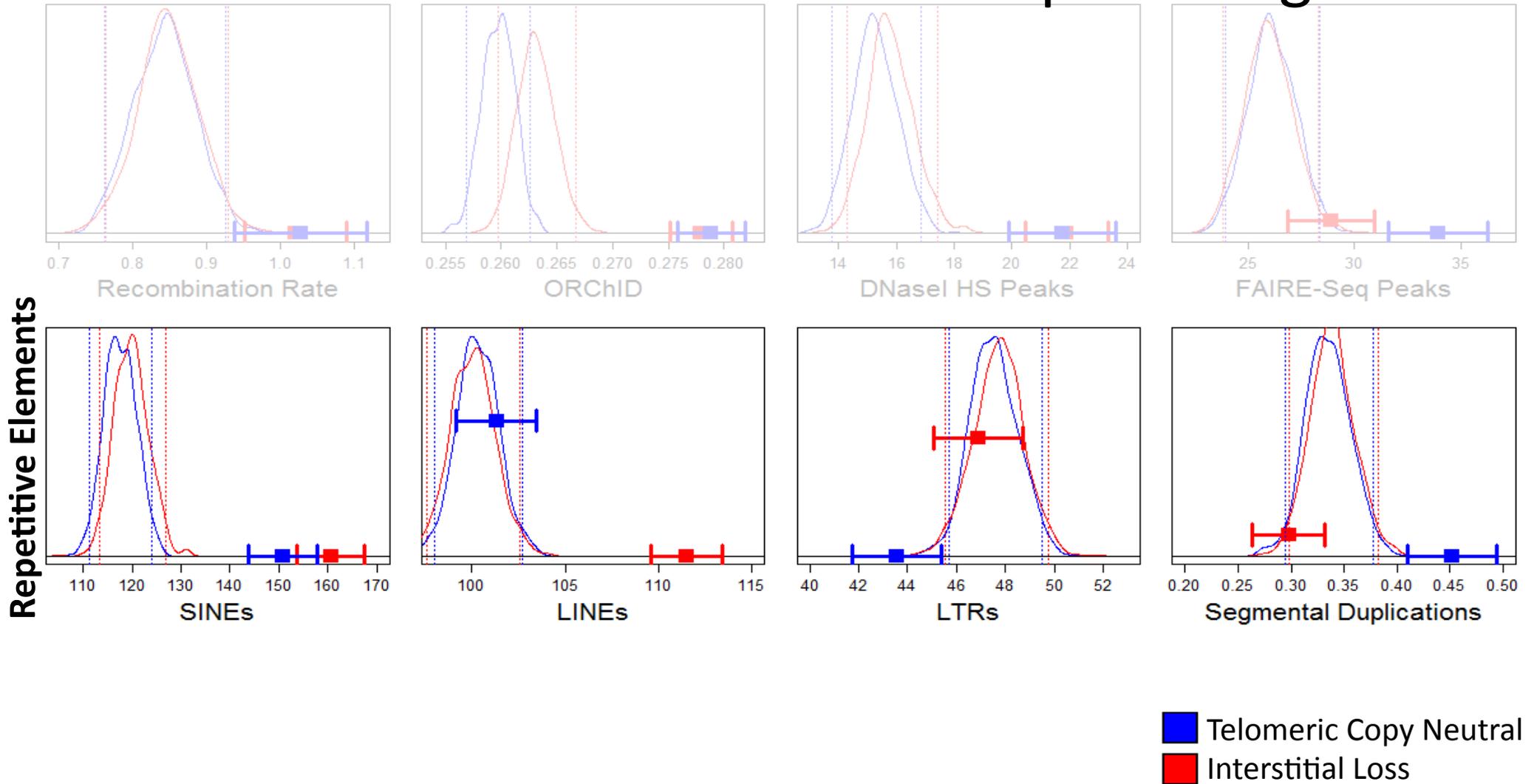
# ENCODE Features around Breakpoint Regions



■ Telomeric Copy Neutral  
■ Interstitial Loss

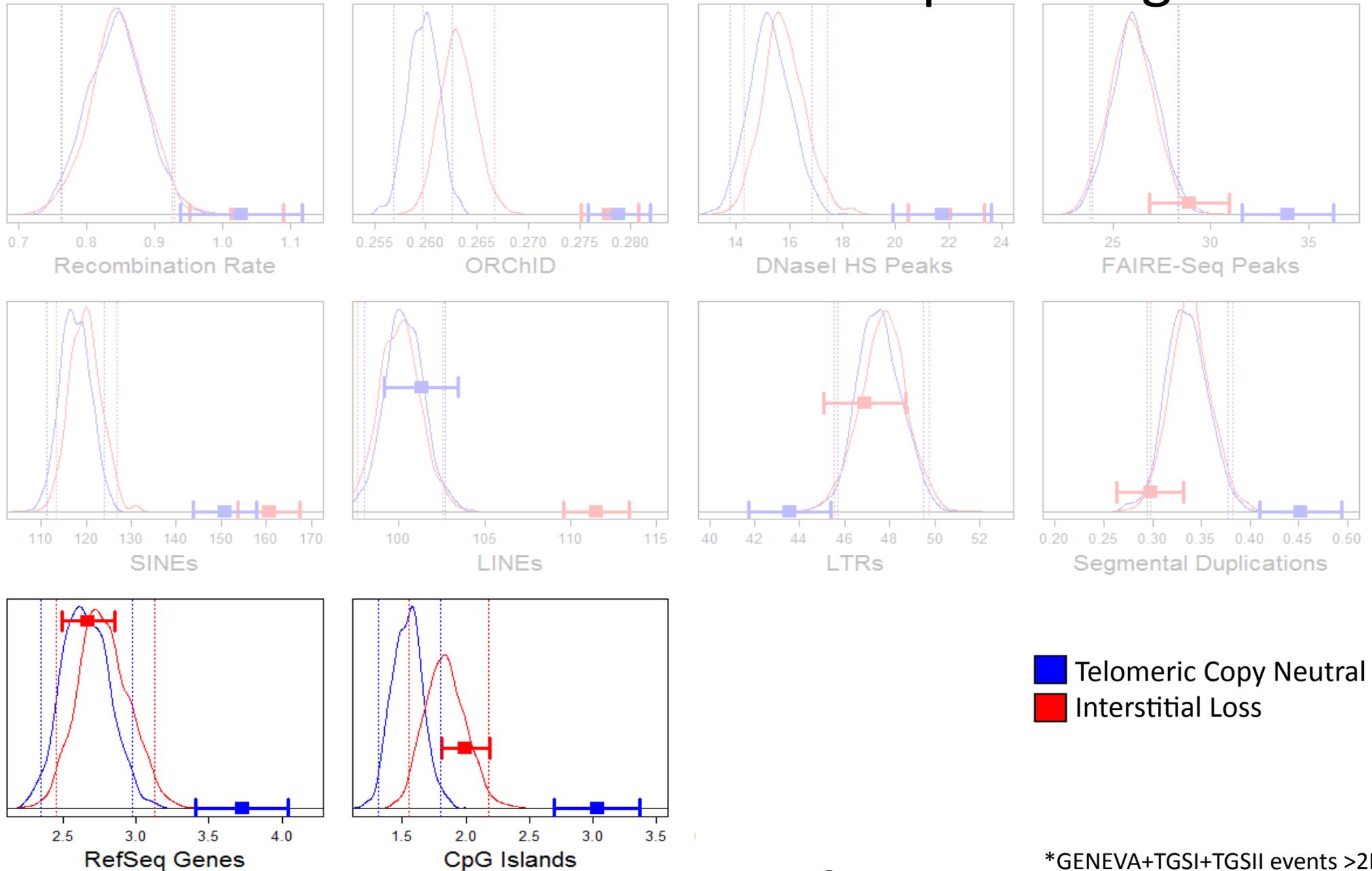
\*GENEVA+TGS1+TGSII events >2Mb

# ENCODE Features around Breakpoint Regions

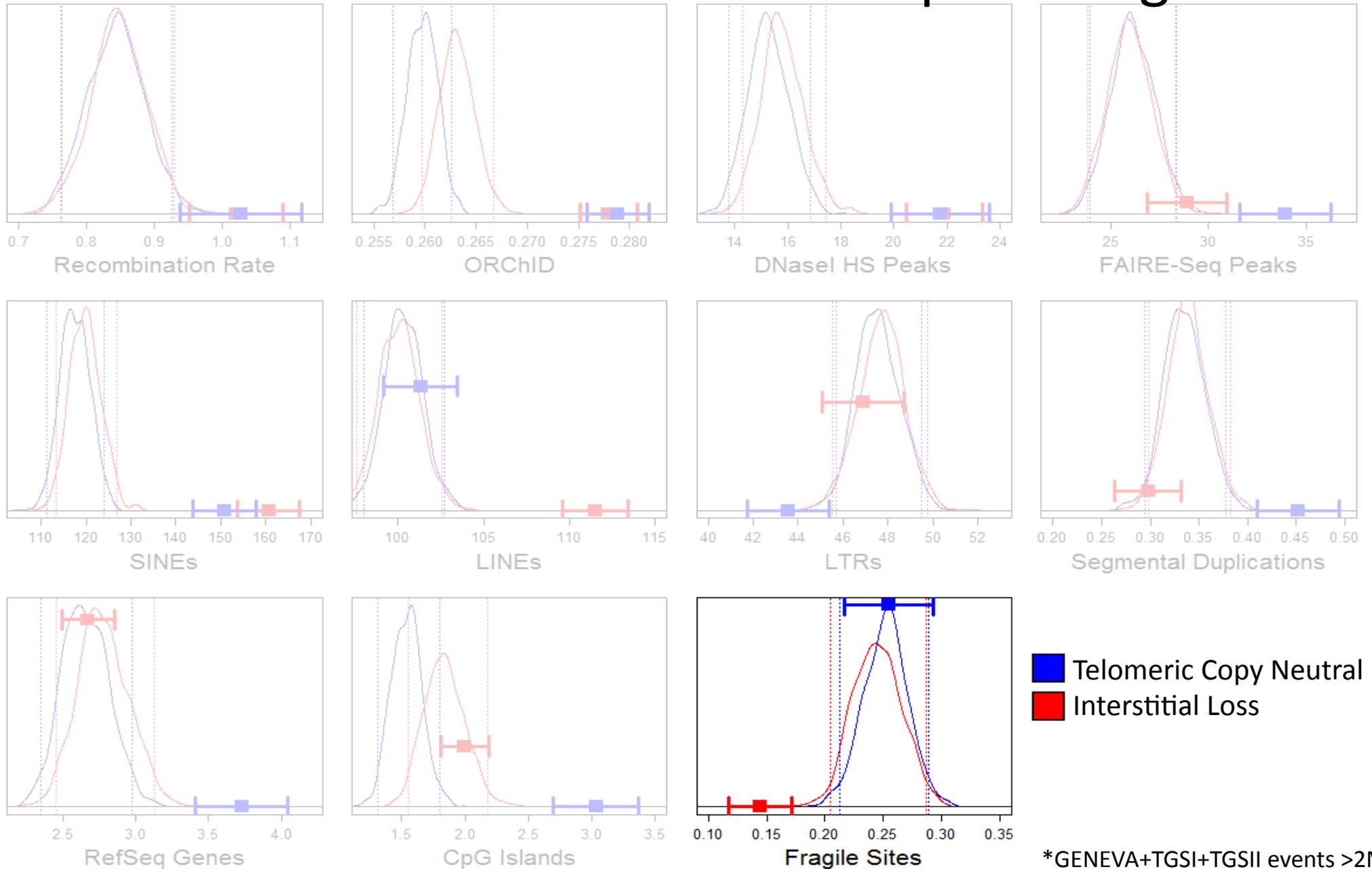


\*GENEVA+TGS1+TGSII events >2Mb

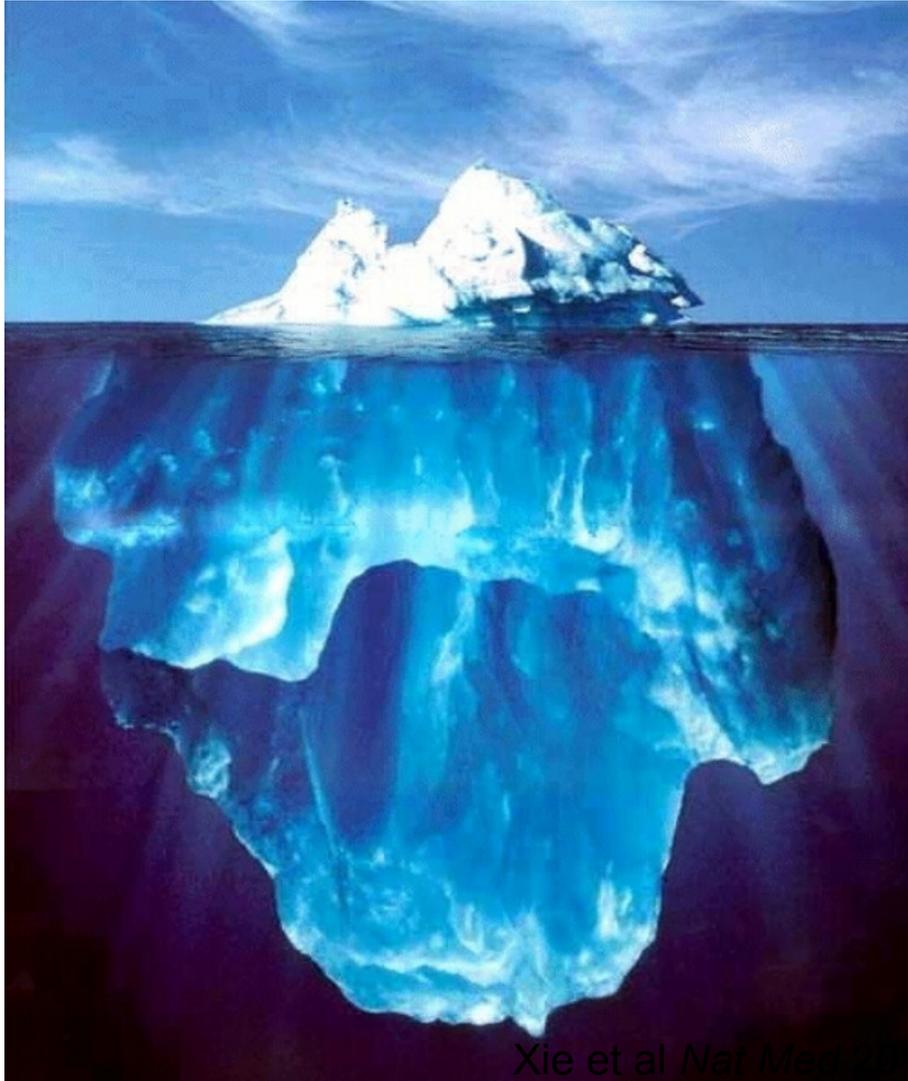
# ENCODE Features around Breakpoint Regions



# ENCODE Features around Breakpoint Regions



# Detectable Mosaicism: Tip of the Iceberg?



Large events detected  
by SNP arrays/aCGH

Detect smaller events  
with new algorithms  
for NGS (NEJM 2014)

“U” shape curve  
Seen in very young &  
aging population

Significance for aging  
diseases

# Current Challenges of Explaining Susceptibility

- Tissue Specificity
  - Tissue of origin
  - Adjacent cells
  - Immunological Modulation
    - Example: Selective Success of Immune Blockade (PD-1)
- Timing of Effect
- Interaction with environmental stimuli

# Immense Value of ENCODE

## Scientific

- Spectacular Resource for Understanding the Functional Basis of Susceptibility
  - Prioritization of variants
- Opportunity to Explore Novel Elements
  - Individual
  - Interactions

## Cultural

- **Team Science**
  - Short Term
  - Long Term
- **Establish Thresholds & Standards**
  - Driven by Questions at hand

## **NCI-DCEG**

Joseph Fraumeni

Robert Hoover

Peggy Tucker

Meredith Yeager

Mitch Machiela

Nilanjan Chatterjee

Juhyn Park

Zhaoming Wang

Weiyin Zhou

Nathaniel Rothman

Mila Prokunina-Olsson

Joshua Sampson

Victoria Fisher

# **Acknowledgements**

## **NCI-Special Experts**

David Hunter-HSPH

Peter Kraft-HSPH

Montse Garcia-Closas-ICR

***More than 400 Collaborators***

acknowledgements

# The OncoArray Consortium



The **GAME-ON** Network



**HARVARD** | SCHOOL OF PUBLIC HEALTH



Dartmouth-Hitchcock  
**NORRIS COTTON  
CANCER CENTER**

Keck Medicine of **USC**



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Disease Research  
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