



UNIVERSITY OF UTAH  
HEALTH SCIENCES

# Family Studies and Complex Disease

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Department of Human Genetics

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# Family studies and complex disease: Applications

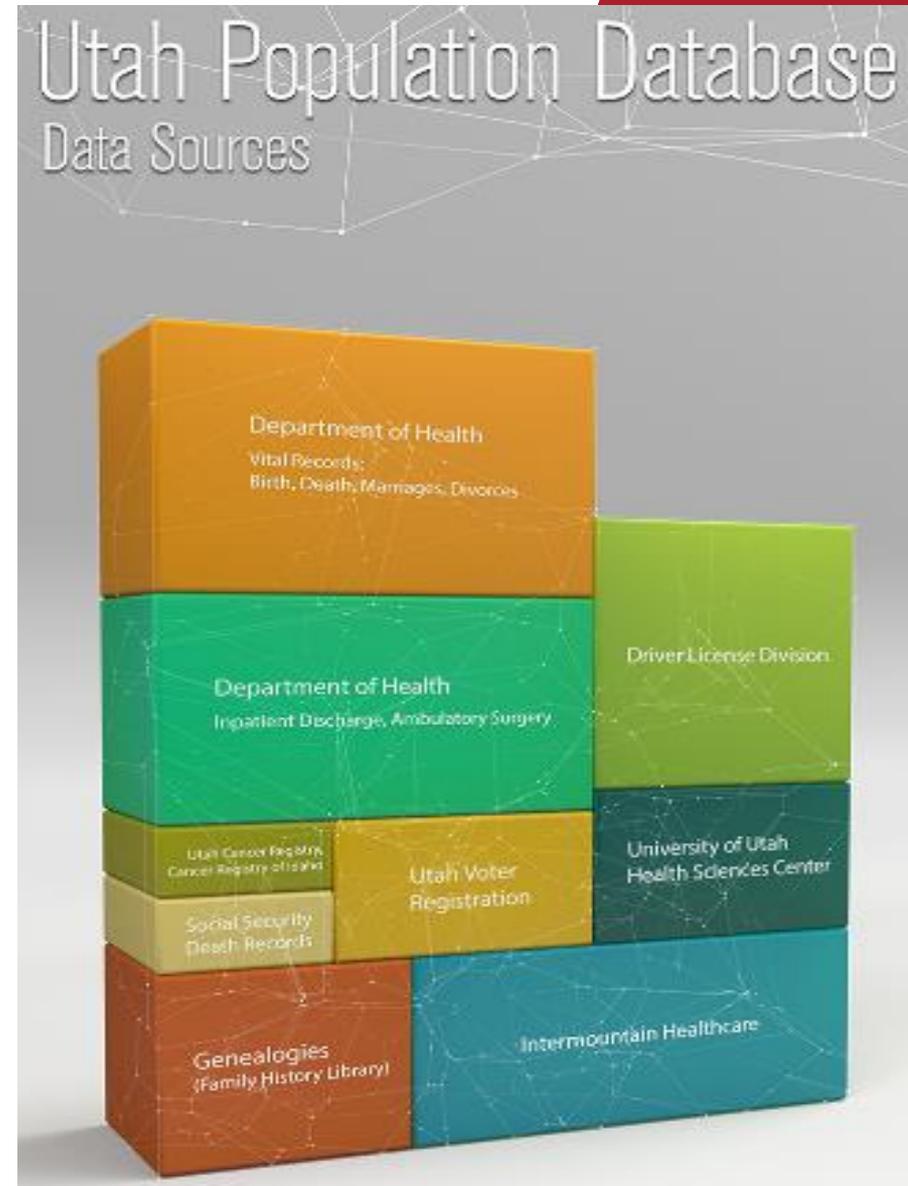
- Detection of causal *de novo* mutations
  - CNVs: e.g., autism, schizophrenia
  - SNVs: e.g., intellectual disability (Gilssen *et al.*, 2014, *Nature*), congenital heart disease (Jin *et al.*, 2017, *Nat. Genet.*)
- Parent-of-origin effects
  - e.g., paternal CNV bias for autism (Brandler *et al.*, 2018, *Science*)
- Germline and somatic mosaicism
- Detection of Mendelian subsets with complex disease phenotypes (*BRCA1*, *BRCA2*, *APC*, *etc.*)
- Multigenerational pedigrees allow detection of shared genomic segments containing rare disease-causing variants
- Families provide opportunity for long-term, longitudinal studies and return of clinically significant results

# A Utah nuclear family



# Utah Population Database (UPDB)

- University of Utah resource to support medical research, initiated in the 1970s
- 10 million people in large, multigenerational pedigrees are linked to more than 25 million medically relevant records
- Geocode information available
- Now expanded to 100 million people



# Utah Genome Project (UGP): >7,000 cases sequenced; >50 diseases

WGS/WES + large pedigrees from 10-million-member Utah Population Database

- Amyotrophic lateral sclerosis
- Genetics of extreme thinness to develop obesity interventions
- Spontaneous preterm birth
- Familial childhood cancers (Ewing sarcoma, Wilms tumor, germ cell tumors)
- DNA repair genes and cancer
- Hematologic cancers: CLL, CML, multiple myeloma
- Common cancers: breast, prostate, colorectal
- Psoriasis and psoriatic arthritis
- Juvenile idiopathic arthritis
- Crohn disease
- Osteoarthritis
- Idiopathic pulmonary fibrosis
- Familial cardiac arrhythmia
- Autism
- Suicide
- Primary ovarian insufficiency
- WGS of Utah CEPH pedigrees (n= ~600)

# Evaluating *de novo* mutations (DNMs) using Utah CEPH pedigrees



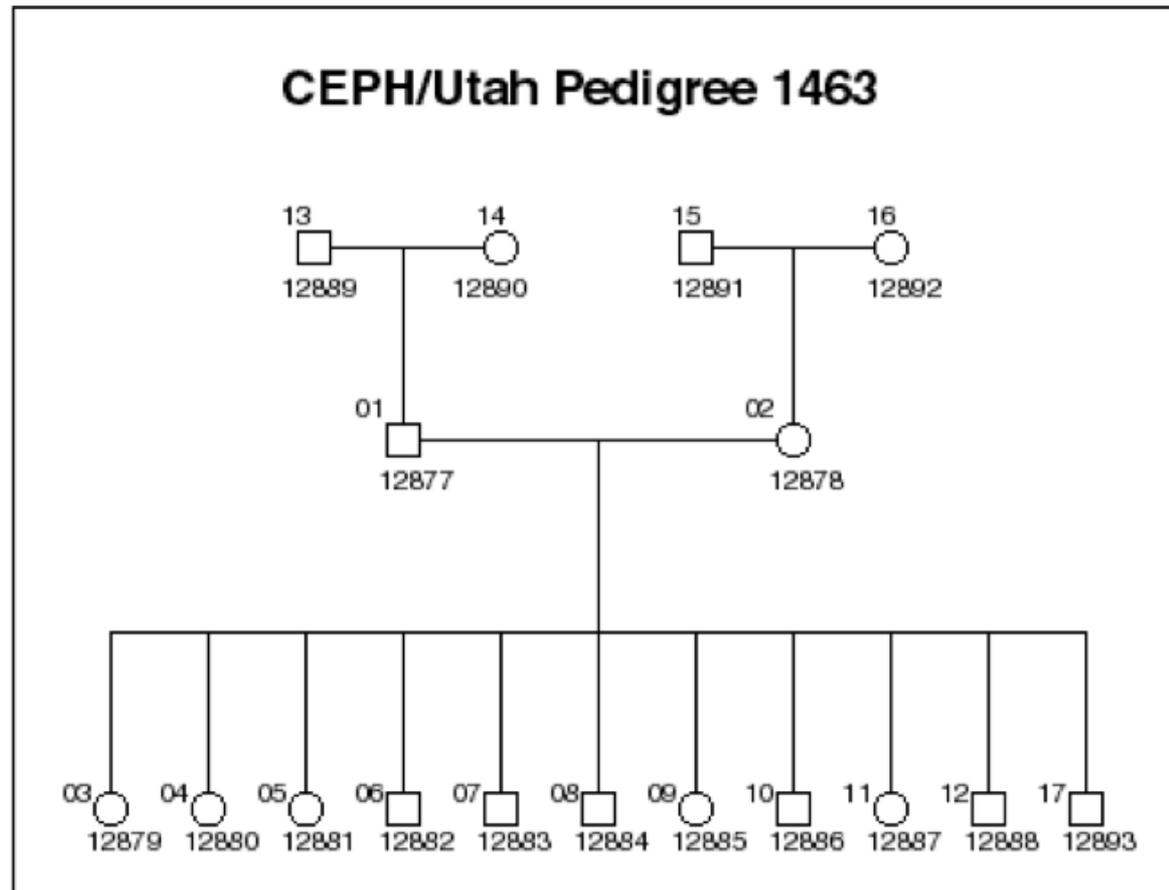
Tom Sasani,  
PhD student

**Centre d'Etude du  
Polymorphisme Humain (CEPH):  
Collaborative Genetic Mapping  
of the Human Genome**

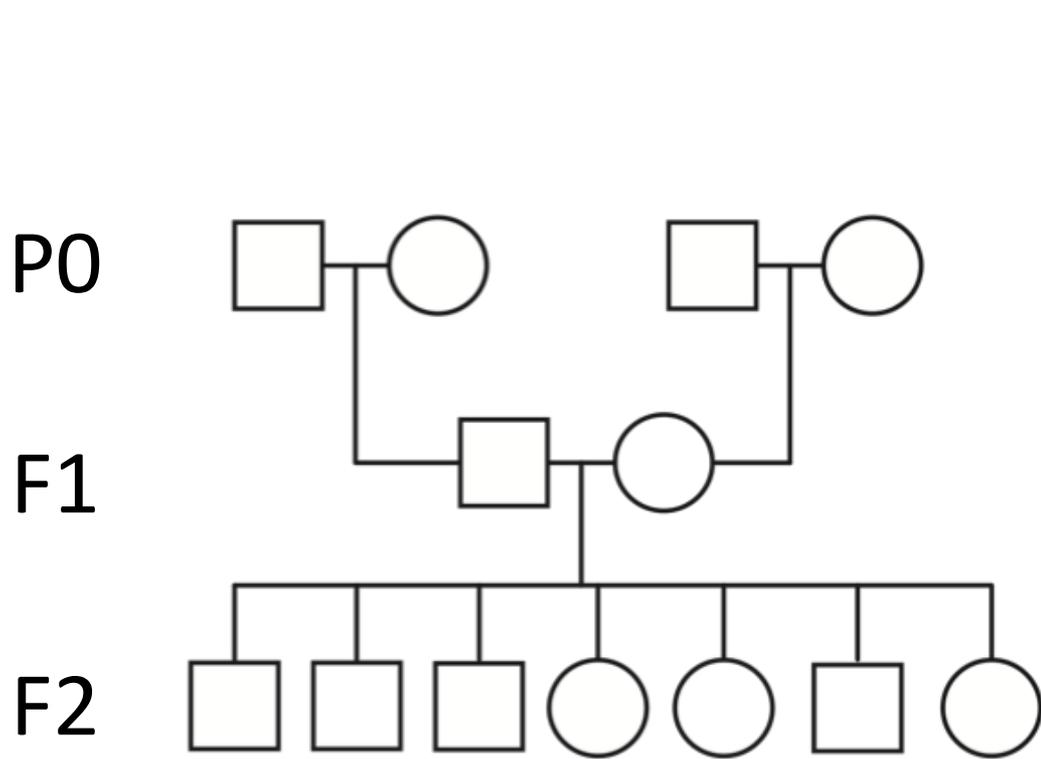
JEAN DAUSSET,\* HOWARD CANN,\*  
DANIEL COHEN,\* MARK LATHROP,\*  
JEAN-MARC LALOUEL,† AND RAY WHITE†<sup>1</sup>

\*Centre d'Etude du Polymorphisme Humain (CEPH), 27 rue  
Juliette Dodu, 75010 Paris, France; and †Howard Hughes  
Medical Institute and Department of Human Genetics,  
University of Utah Health Sciences Center, Salt Lake City,  
Utah 84132

*Genomics*, 1990



# Overview of CEPH/Utah dataset



603 individuals

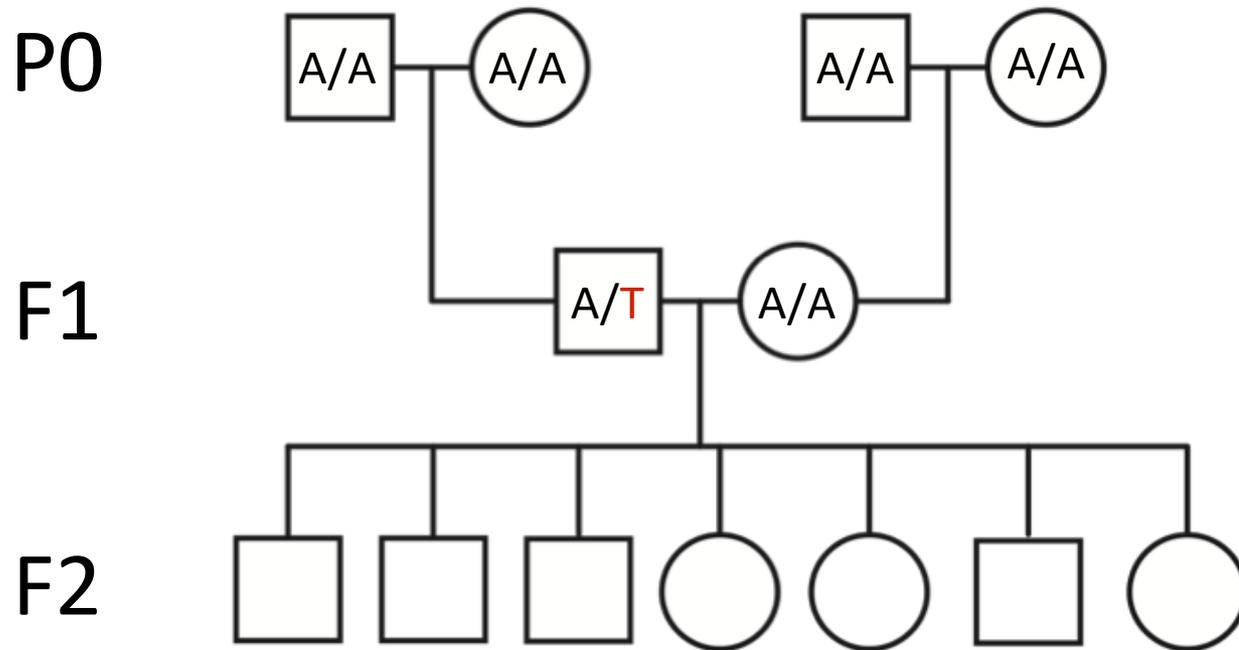
34 families

4-16 F2 per family

***DNA from original  
blood draws***

Illumina sequencing @  
~32X coverage for all  
individuals

# CEPH/Utah pedigrees offer excellent power for detecting and validating DNMs by transmission

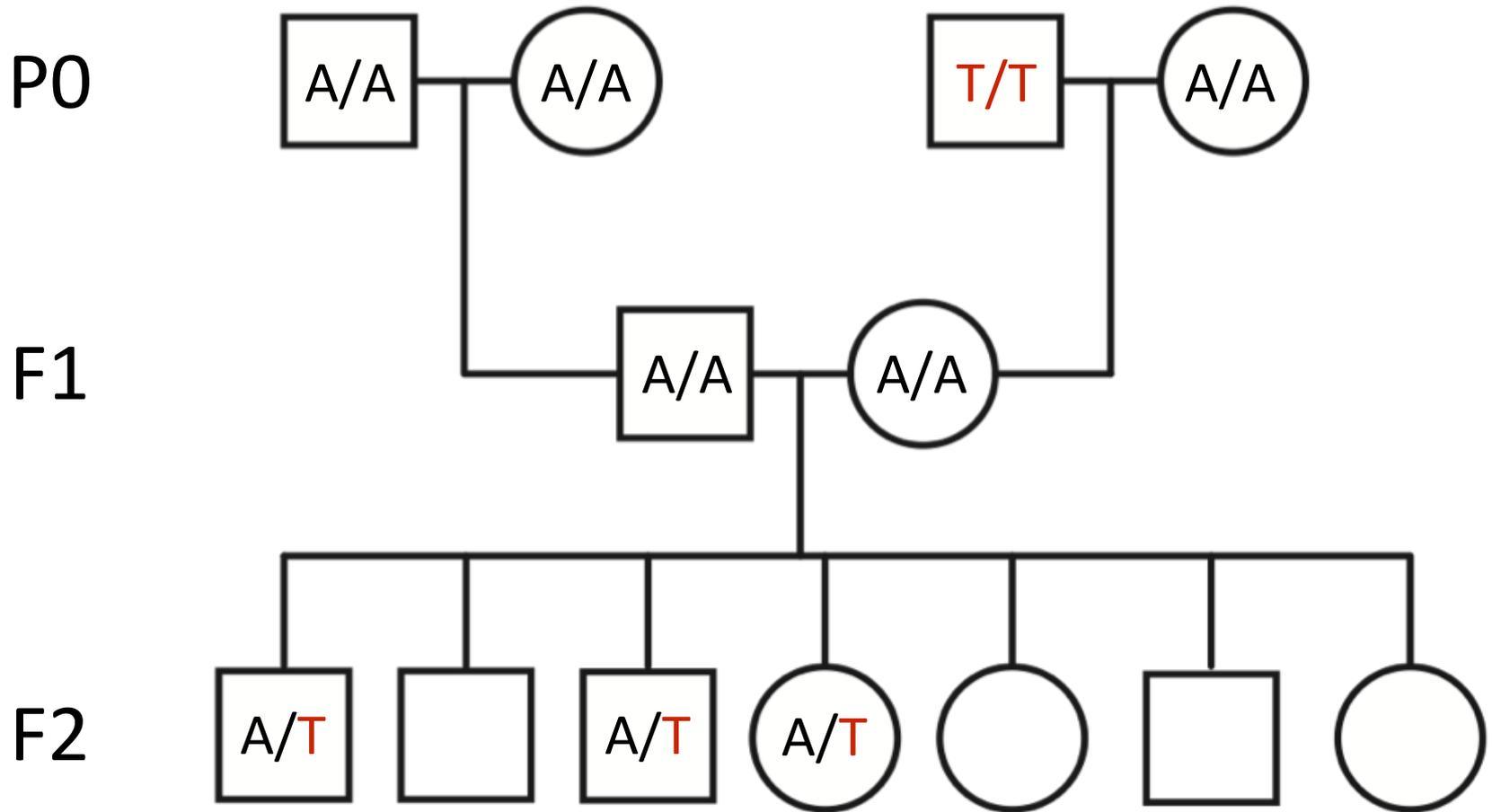


$P$  (not inherited)

$$0.5 \times 0.5 \times 0.5 \times 0.5 \times 0.5 \times 0.5 \times 0.5$$

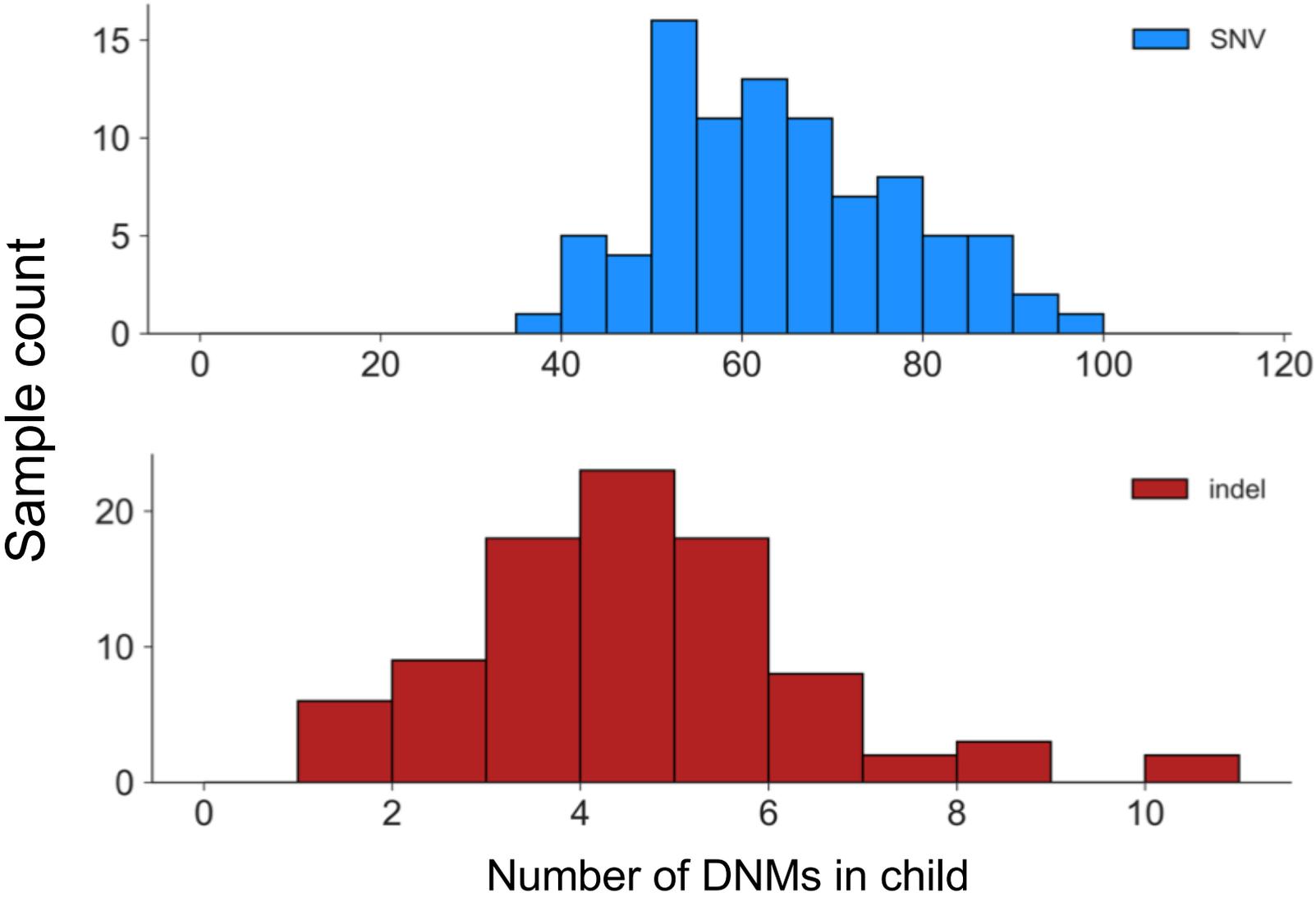
$$= 0.5^7 = 0.0078 - \text{a likely false positive}$$

# Estimating a false negative rate of DNM detection

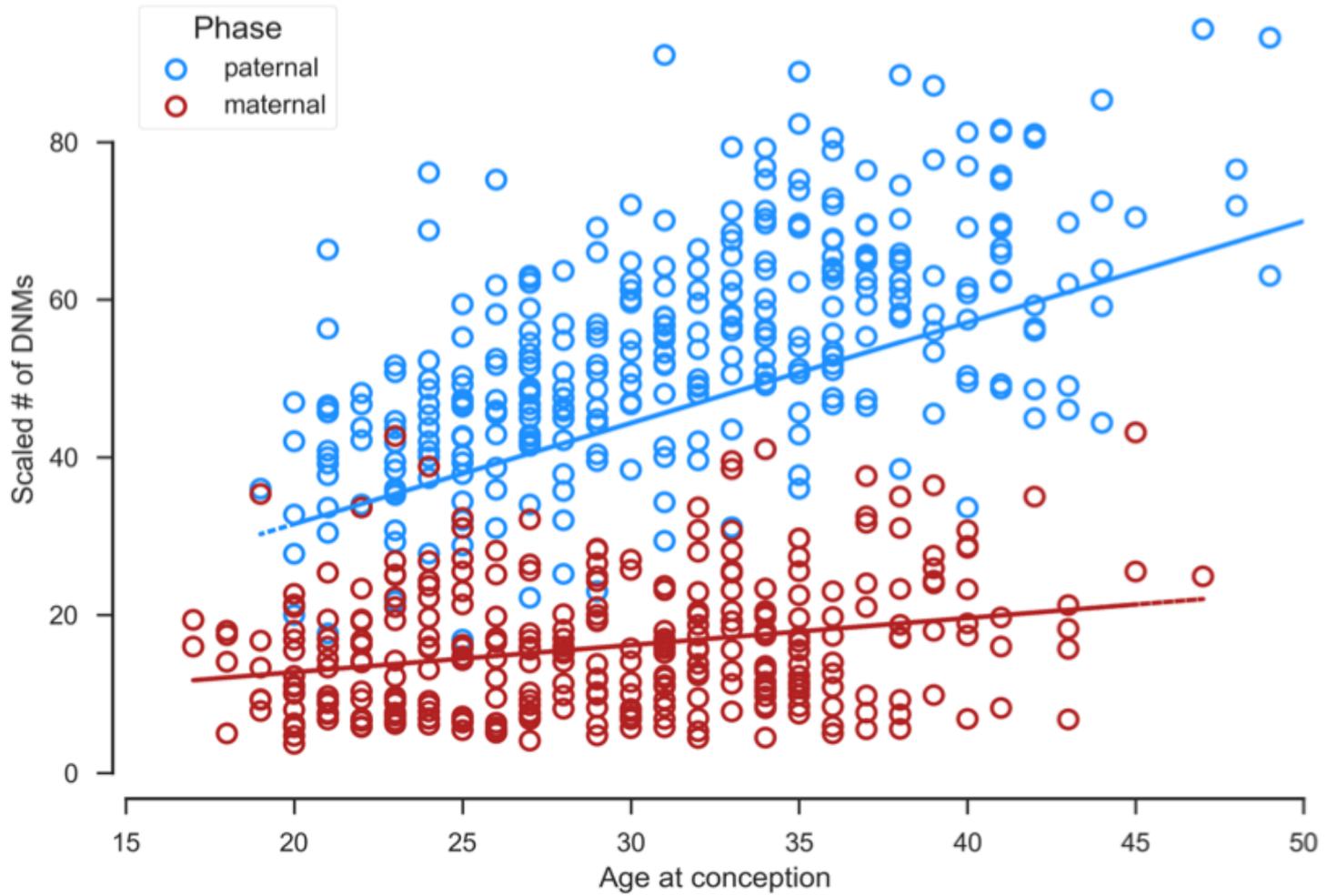


False negative rate is  $< 4\%$

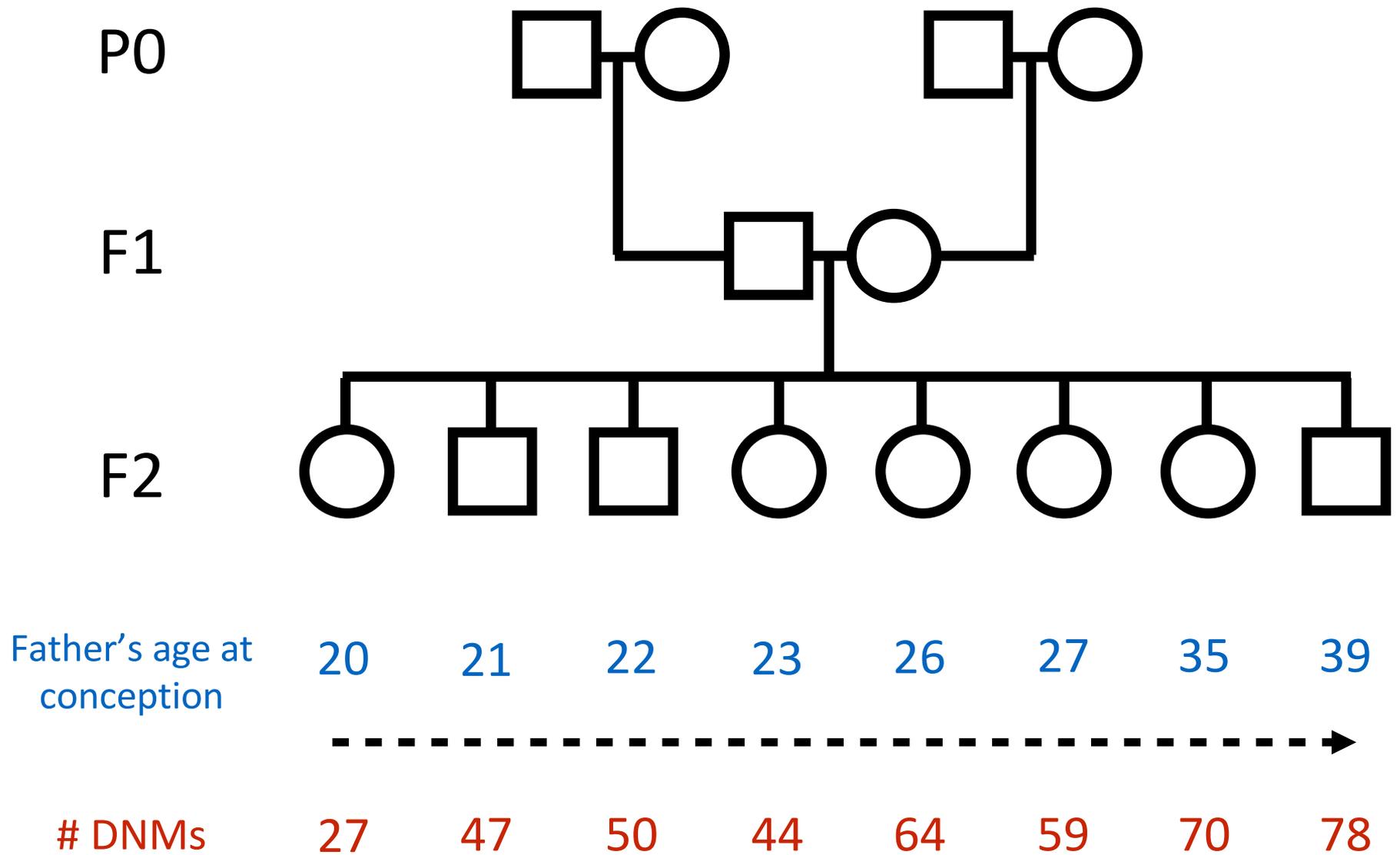
By validating DNM in the F1 generation by transmission, we find an average of 71 DNMs per individual ( $\mu = 1.3 \times 10^{-8}$ )



We observe ~1.3 additional DNM per year of paternal age;  
0.3 DNM per year of maternal age

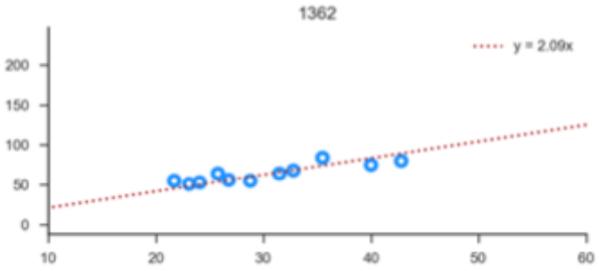
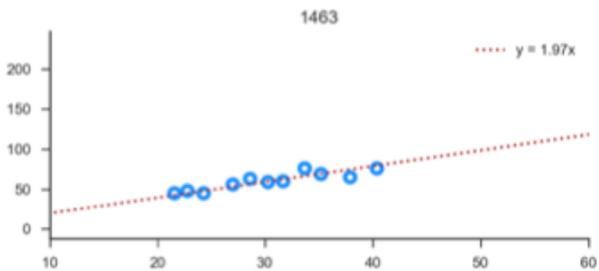
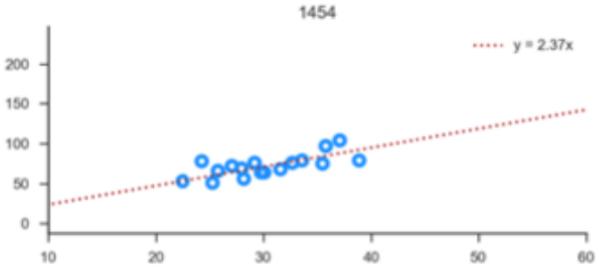
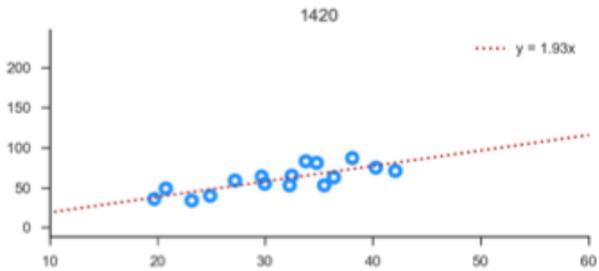
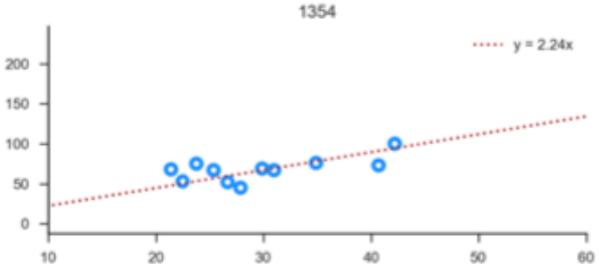
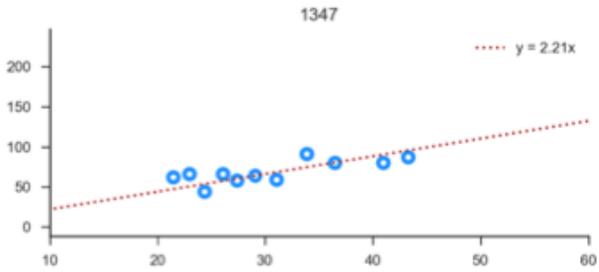
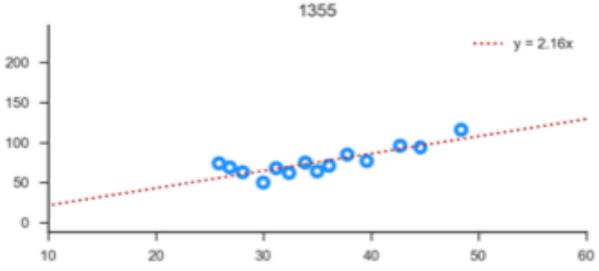
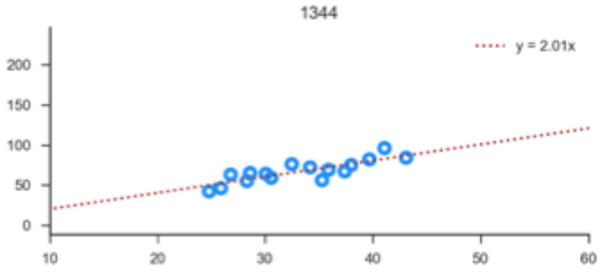


# Large F2 allows longitudinal evaluation of DNM rate



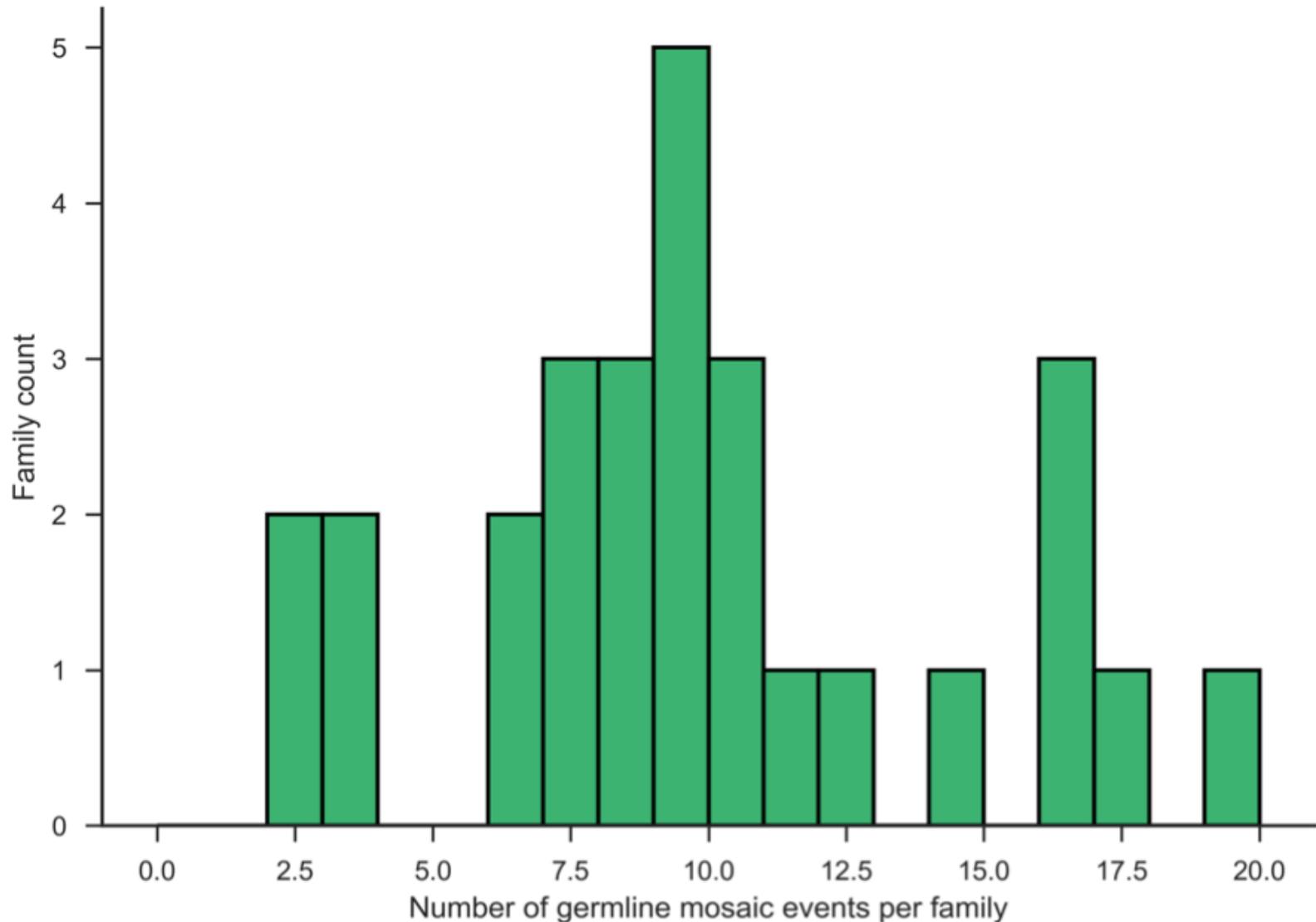
# Paternal age effect within individual CEPH pedigrees

# DNMs in proband



Paternal age at birth

# 339 germline mosaic events (de novo variants in multiple F2s) identified across CEPH families



# “Return to CEPH”

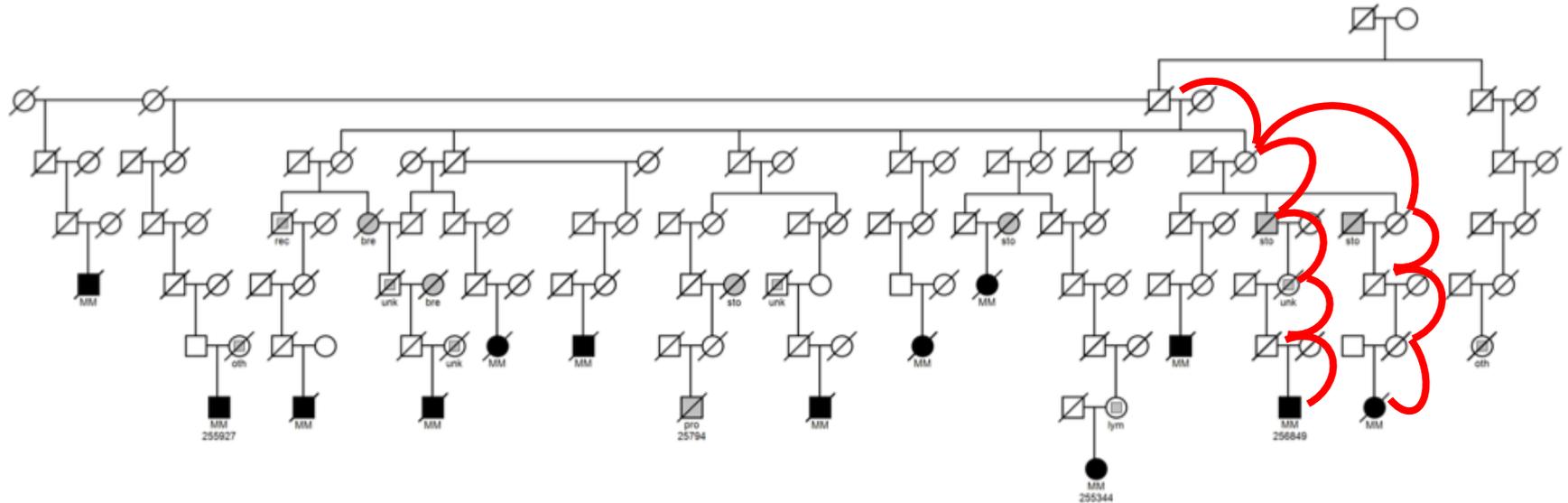
- Families were recontacted in 1990s
- 180 phenotype variables collected
  - Lipid profiles
  - Blood chemistry panel
  - Blood pressure, ECG, echocardiogram
  - Anthropometrics
  - Behavioral assessments
  - Plasma, urine samples
- Fourth-generation data collection to be initiated with recontact of families and follow-up

# Finding new genes in large pedigrees: Shared Genomic Segments (SGS)



Nicola Camp, PhD

- Ideal for large pedigrees -- inherited genomic sharing unlikely in very distant relatives
  - Inherited sharing across 15 meioses is unlikely across a genome
  - Shared environment unlikely for distant relatives
  - Small sample sizes have good power for gene detection



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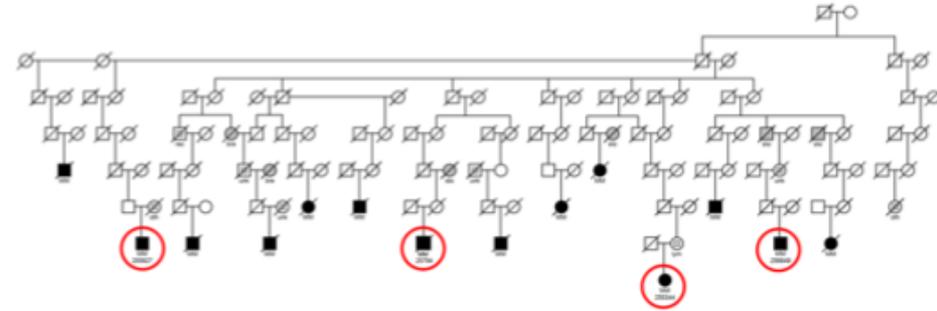
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- High-density SNPs provide good resolution of genomic regions
  - SNP microarrays
  - SNPs extracted from WGS

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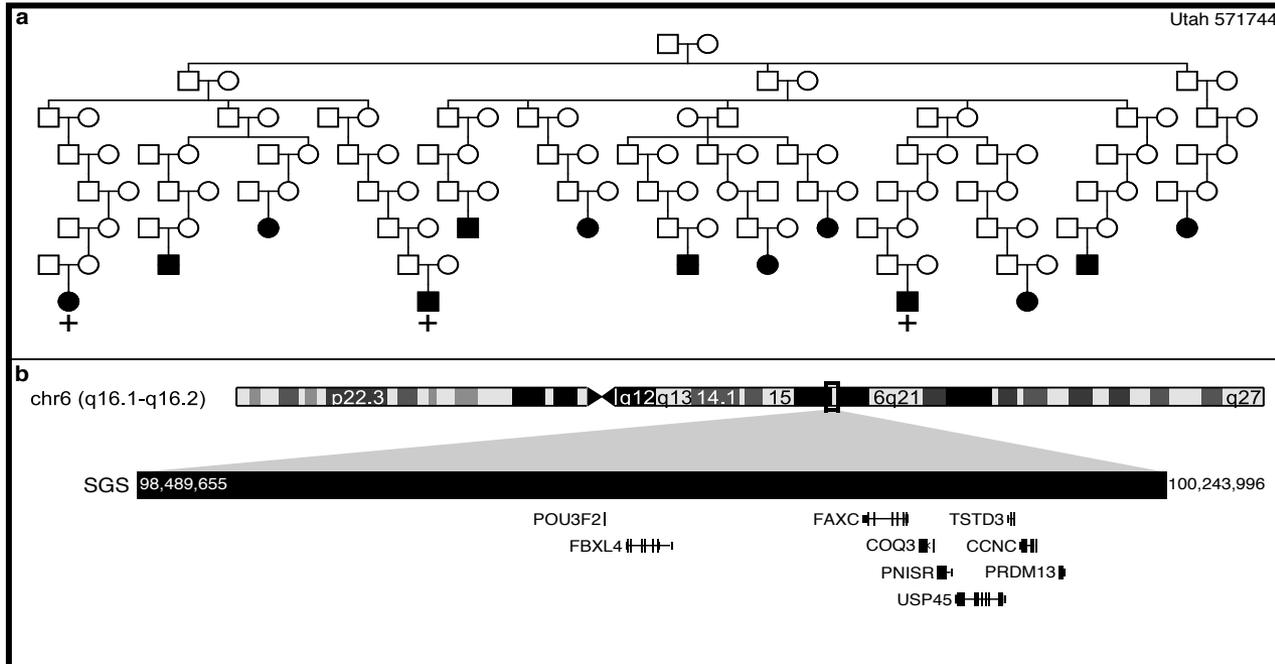
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- High-density SNPs provide good resolution of genomic regions
  - SNP microarrays
  - SNPs extracted from WGS
- Simple concept -- identity-by-state (IBS) used to infer identity by descent (IBD)
  - Length shared IBS is significantly longer than expected by chance, then likely IBD
  - Empirical estimate of genome-wide significance
    - Pedigree structure
    - Marker and LD map
  - Intra-familial heterogeneity assessed with all-subsets approach

# SGS applied to multiple myeloma

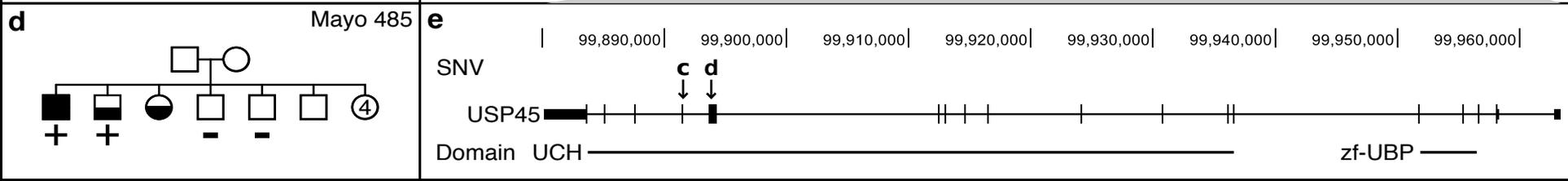
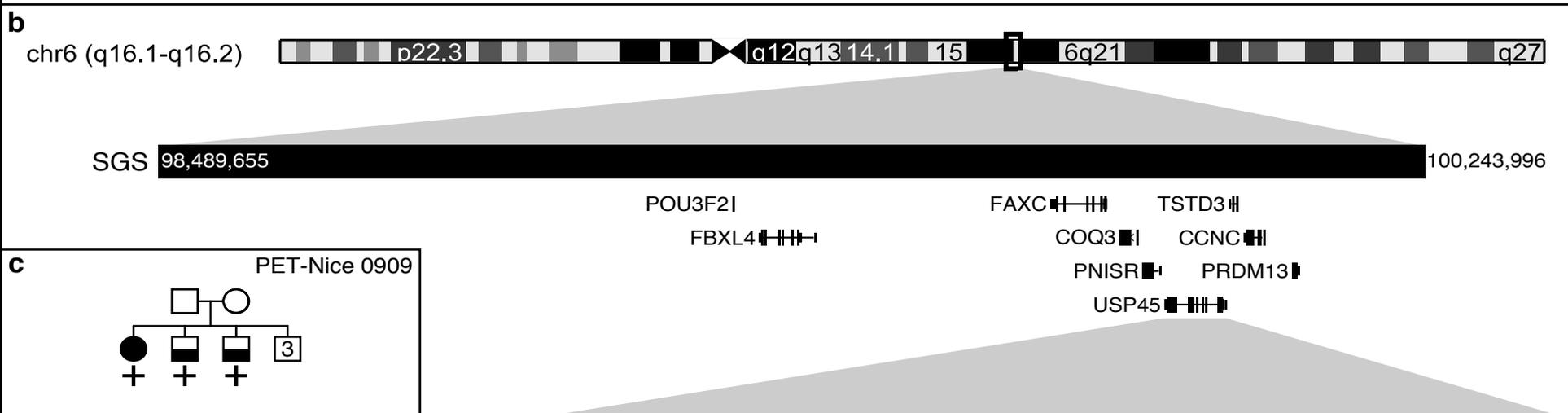
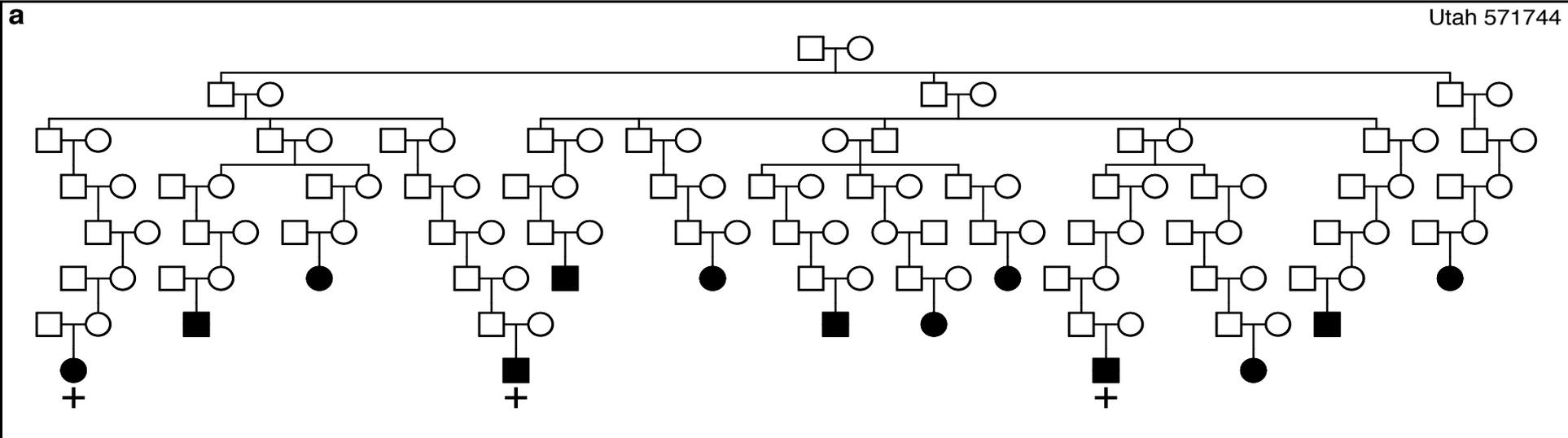
- Late-age onset hematologic cancer
- <50% five-year survival
- GWAS accounts for ~20% of heritability (Mitchell *et al.*, 2016, *Nat. Comm.*)
- 11 high-risk MM pedigrees identified from UPDB
  - Genotype SNP array
  - 3-4 sampled cases per pedigree
  - 8-23 meioses between cases
- Shared Genomic Segment analysis
- Exome sequencing
  - Utah pedigrees
  - Mayo pedigrees
  - INSERM, France pedigrees
  - Cornell pedigrees
  - 200 early onset MM
  - Publicly available data
    - coMMpass MM sporadic cases
    - controls



# One genome-wide significant SGS region (1.8 Mb) containing nine genes



# Exome sequencing in the region



# One gene, *USP45*, contains pathogenic variants

Table 1. Significant

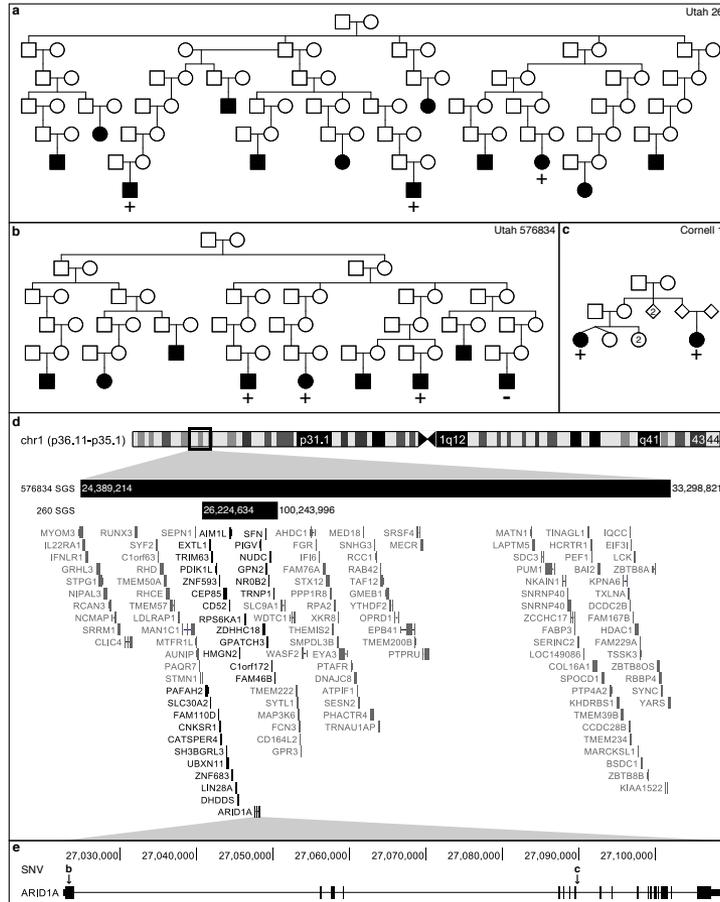
Family	MM	MGUS	Meioses	Type	Position <sup>~</sup>	Length <sup>†</sup>	P-Value	Gene	Consequence	Impact	AAF <sup>^</sup>
Utah 571744	3		20	SGS	6:98,489,655—100,243,996	1.8	3.3x10 <sup>-6*</sup>				
PET-Nice 0909	1	2	3	SNV	6:99,891,443			<i>USP45</i>	p.Gln691*	Stop Gain	None
Mayo 458	1	1	2	SNV	6:99,893,787			<i>USP45</i>	p.Gln621Glu	Missense	None

<sup>~</sup>Build HG19. <sup>†</sup>In mega-bases. <sup>\*</sup>Genome-wide significant. <sup>^</sup>Alternate allele frequency based on the non-TCGA, non-Finnish, European ExAC individuals.

## *USP45* is a DNA repair gene

- Deubiquitylates ERCC1, a catalytic subunit of the ERCC1-ERCC4 DNA repair complex
- Mouse USP45 knockout models
  - higher levels of ubiquitylated ERCC1
  - cells are hypersensitive to UV radiation and DNA inter-strand cross-links
  - DNA damage repair is impaired
- Well-established role in tumor resistance to chemotherapy

# Regions identified by multiple pedigrees



*ARID1A* involved in chromatin remodeling

Intolerant to LOF and missense variation

Prominent role in cancer somatic mutations  
 --tumor suppressor

# ARID1A

**Table 1.** overlapping SGSs and segregating SNVs.

Family	MM	MGUS	Meioses	Type	Position <sup>~</sup>	Length <sup>l</sup>	P-Value	Gene	Consequence	Impact	AAF <sup>^</sup>
Utah 576834	3		12	SGS	1:24,389,214—33,298,821	8.9	3.0x10 <sup>-4</sup>				
Utah 260	3		16	SGS	1:26,224,634—27,384,988	1.2	2.1x10 <sup>-4</sup>				
Utah 576834	3		12	SNV	1:27,023,162			ARID1A	p.Ser090Gly	Missense	0.0000
Cornell MM12	2	0	4	SNV	1:27,089,712			ARID1A	p.Met890Val	Missense	0.0001

<sup>~</sup>Build HG19. <sup>l</sup>In mega-bases. <sup>\*</sup>Genome-wide significant. <sup>^</sup>Alternate allele frequency based on the non-TCGA, non-Finnish, European ExAC individuals.

Ding (2017) *Cancer Research*. Pediatric ALL mutation profiles. ARID1A potential tumor suppressor; Rao (2017) *Carcinogenesis*. 5 most frequently somatically deleted genes in liver cancer; (2017) *Cancer Discovery*. NEWS HIGHLIGHT. ARID1A deficiency promotes colon cancer via enhancer dysregulation; Mathur (Dec 2016) *Nature Genetics*. ARID1A loss impairs enhancer mediated regulation and drives colon cancer in mice. Coatham (Dec 2016) *Modern Pathology*. Inactivating mutations and loss of ARID1A expression in aggressive ovarian and endometrial cancer; Agrihoti (Nov 2016) *Nature Genetics*. Recurrent mutations in ARID1A in nerve sheath tumors; Berns (Nov 2016) *Clinical Cancer Research*. Reduced ARID1A expression controls resistance in breast cancer (HER2/P13K/mTOR targeting agents);...

# Other examples of SGS analyses in large Utah pedigrees

- Preterm birth
- Amyotrophic lateral sclerosis
- Small intestinal neuroendocrine tumors
- Autism
- Suicide
- Crohn disease

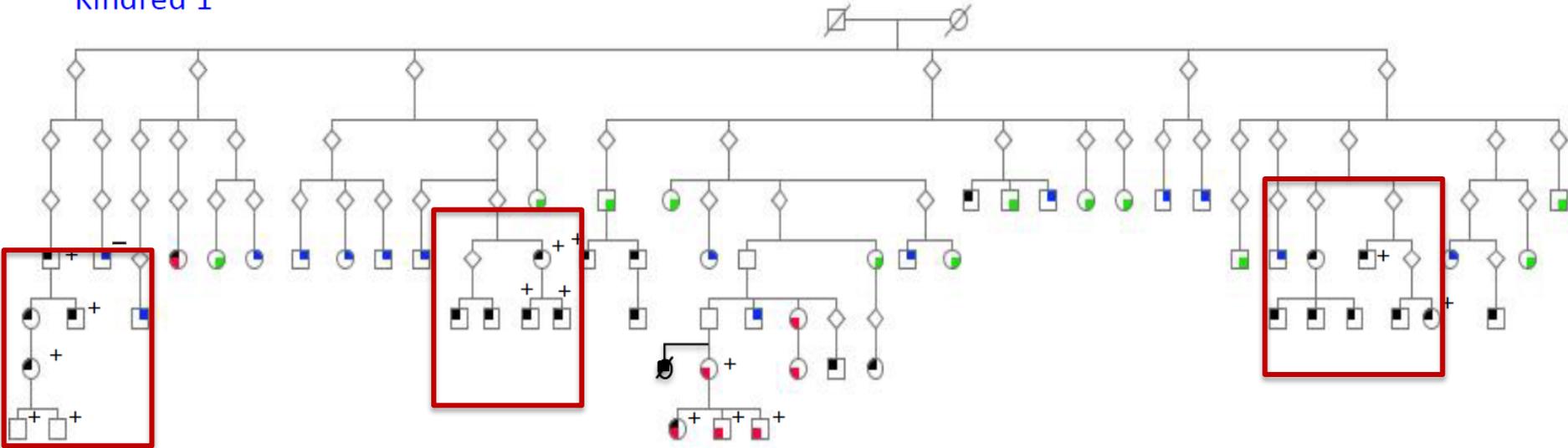
# Family data can identify at-risk pedigree members: Young-onset atrial fibrillation



Martin Tristani-Firouzi, MD

Kindred 1

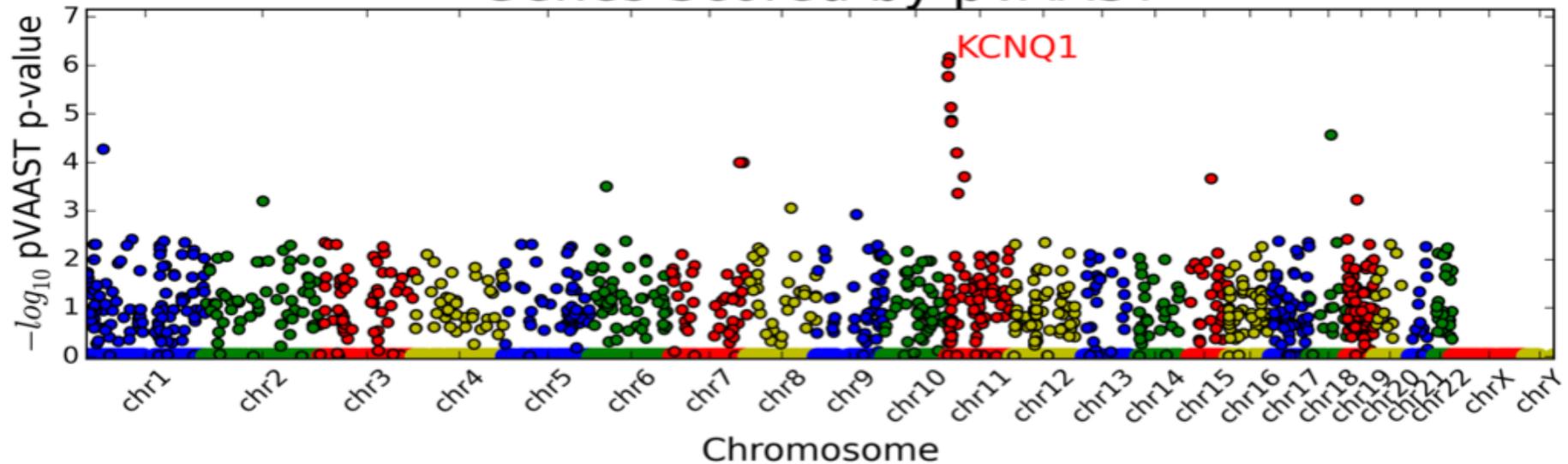
1812 1820



■ AF onset < 40    ■ AF onset > 60    ■ AF NOS    ■ LQTS

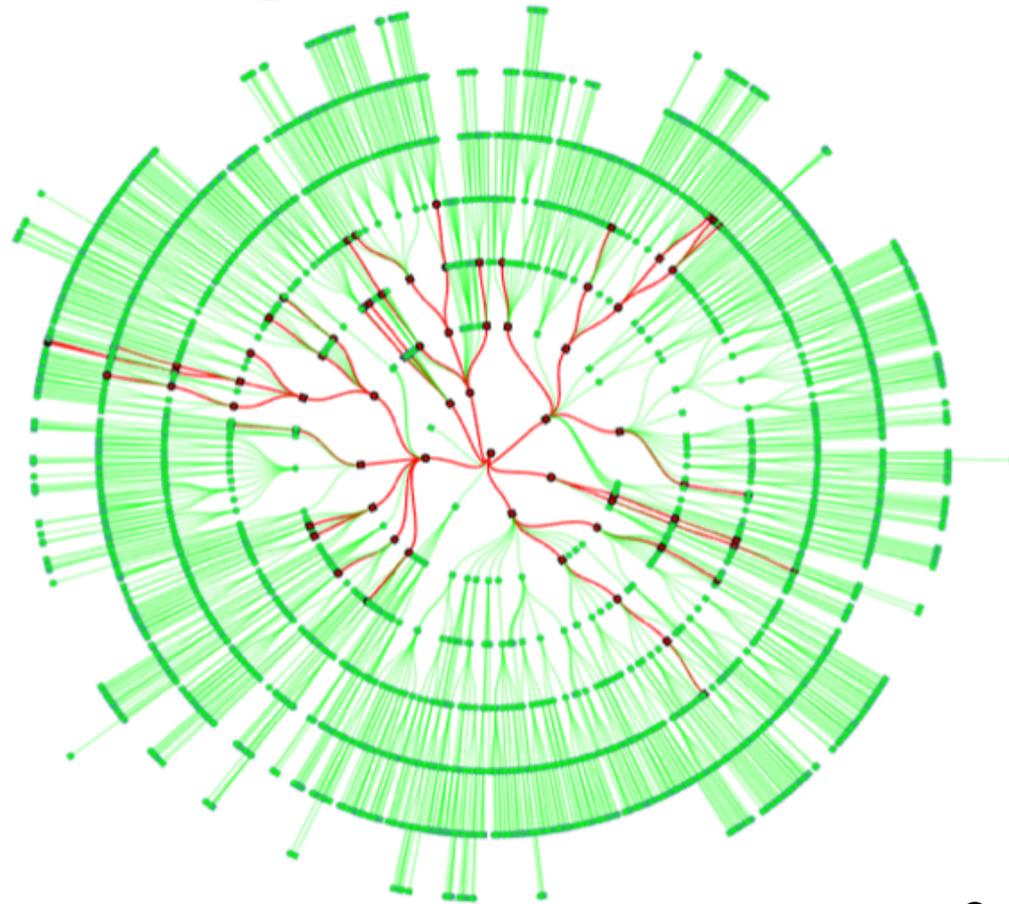
# KCNQ1 mutation causes atrial fibrillation in all family branches

Genes Scored by pVAAST



*KCNQ1* (*LQT1*) originally identified as a long QT syndrome gene in a large Utah family (Keating *et al.*, 1991, *Science*)

# Projecting medical records onto family histories



Collaboration underway  
with AncestryDNA

# SUMMARY: THE VALUE OF FAMILIES

- Evaluation of *de novo* events
- Parent-of-origin effects
- Mosaicism detection
- Discovery of rare variants difficult to identify in samples of unrelated subjects
- Follow-up with communication of testing results

# ACKNOWLEDGMENTS

## Jorde Lab

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Karin Chen, MD  
Kristi Russell  
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Scott Watkins, MS  
Jin Xing, PhD  
David Witherspoon, PhD

## Programmers:

Andrew Farrell, PhD  
Matt Velinder

## CEPH Project

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Benning Society  
Mark Leppert  
Ray White  
DNA samples: Lisa Baird  
UGP: Barry Moore  
Shawn Rynearson  
Deb Neklason  
Breanna Caruso

## DNA Core Facilities

Derek Warner  
HCI sequencing core  
ARUP sequencing core  
WashU Genome Center

## Quinlan Lab

Aaron Quinlan  
Thomas Sasani  
Ryan Layer, PhD

