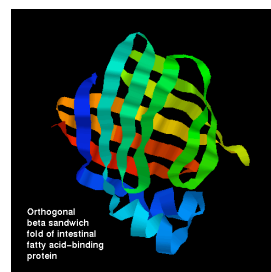
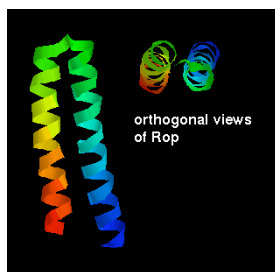


Protein Structure Assessment & Protein Interactions



David Wishart
University of Alberta, Edmonton, Canada
david.wishart@ualberta.ca

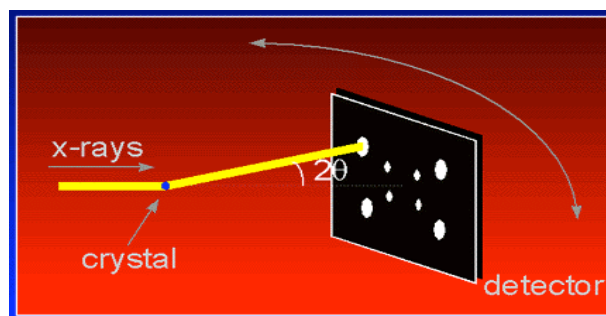
Much Ado About Structure

- Structure ↔ Function
- Structure ↔ Mechanism
- Structure ↔ Origins/Evolution
- Structure-based Drug Design
- Solving the Protein Folding Problem

Routes to 3D Structure

- X-ray Crystallography (the best)
- NMR Spectroscopy (close second)
- Cryoelectron microscopy (distant 3rd)
- Homology Modelling (sometimes VG)
- Threading (sometimes VG)

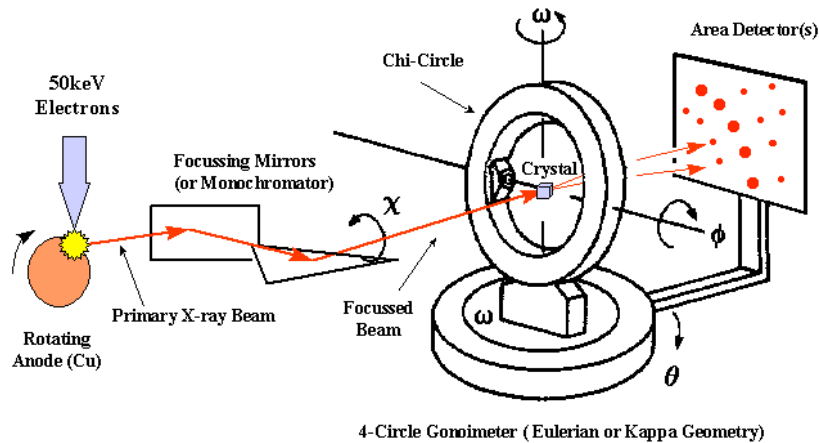
X-ray Crystallography



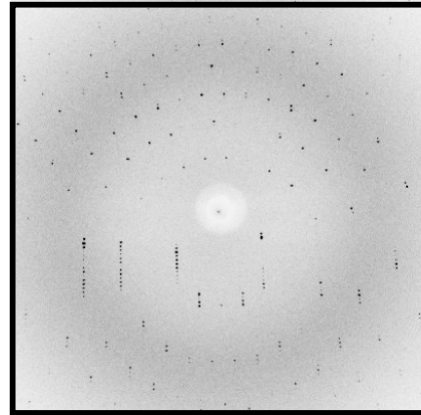
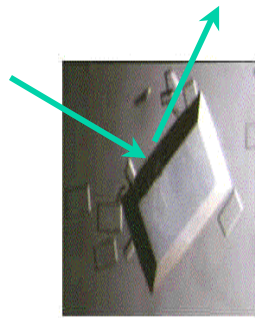
X-ray Crystallography

- Crystallization
- Diffraction Apparatus
- Diffraction Principles
- Conversion of Diffraction Data to Electron Density
- Resolution
- Chain Tracing

Diffraction Apparatus

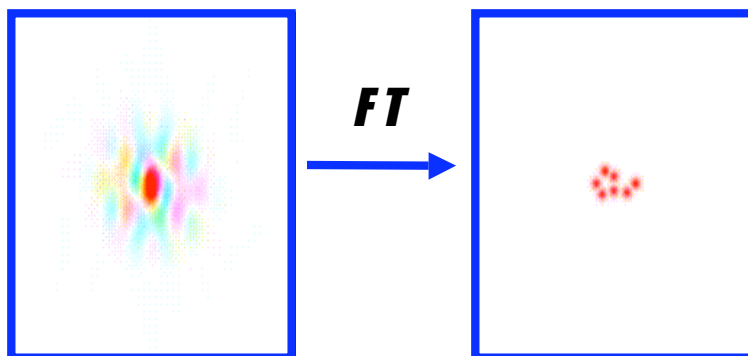


Protein Crystal Diffraction

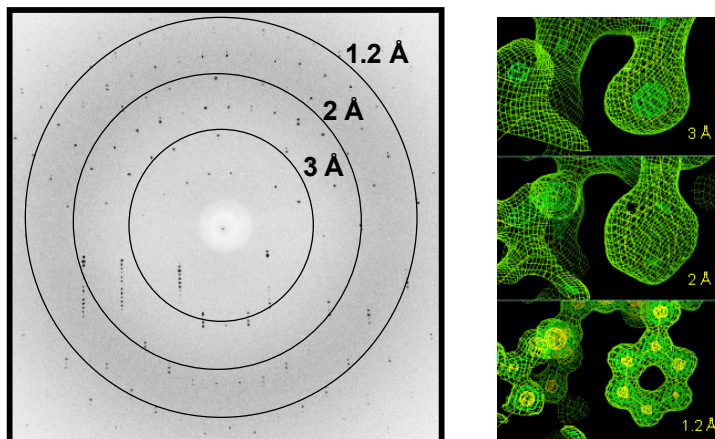


Diffraction Pattern

Converting Diffraction Data to Electron Density



Resolution

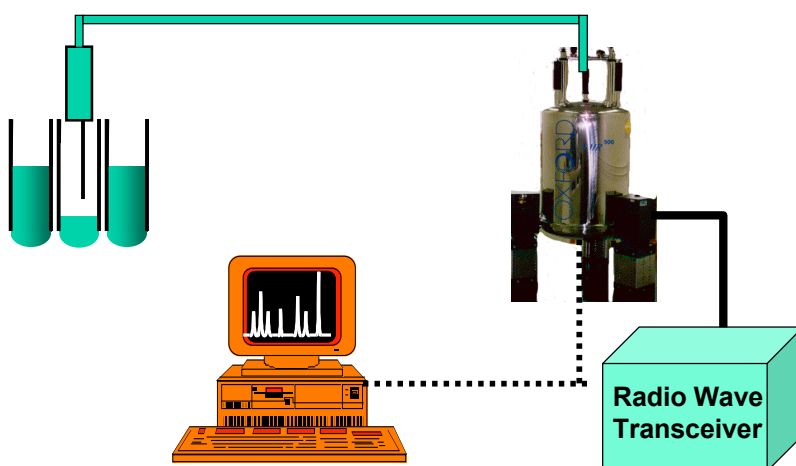


The Final Result

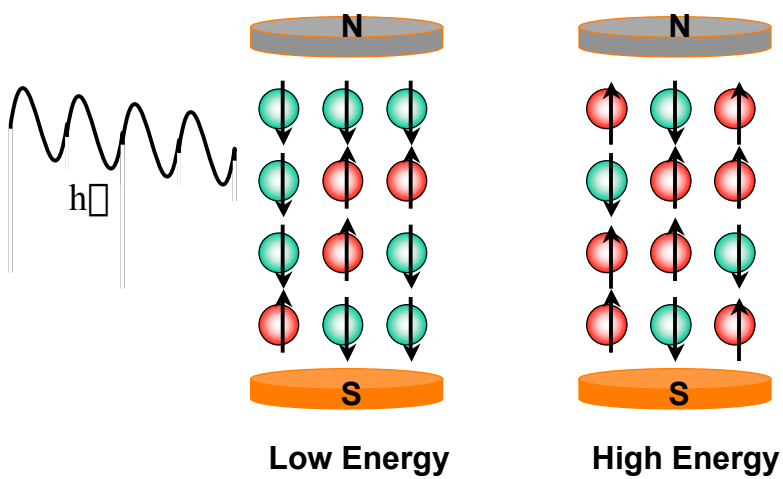
```
ORIGX2      0.000000  1.000000  0.000000      0.000000      2TRX 147
ORIGX3      0.000000  0.000000  1.000000      0.000000      2TRX 148
SCALE1      0.011173  0.000000  0.004858      0.000000      2TRX 149
SCALE2      0.000000  0.019585  0.000000      0.000000      2TRX 150
SCALE3      0.000000  0.000000  0.018039      0.000000      2TRX 151
ATOM       1  N  SER A  1      21.389  25.406  -4.628  1.00  23.22  2TRX 152
ATOM       2  CA SER A  1      21.628  26.691  -3.983  1.00  24.42  2TRX 153
ATOM       3  C  SER A  1      20.937  26.944  -2.679  1.00  24.21  2TRX 154
ATOM       4  O  SER A  1      21.072  28.079  -2.093  1.00  24.97  2TRX 155
ATOM       5  CB SER A  1      21.117  27.770  -5.002  1.00  28.27  2TRX 156
ATOM       6  OG SER A  1      22.276  27.925  -5.861  1.00  32.61  2TRX 157
ATOM       7  N  ASP A  2      20.173  26.028  -2.163  1.00  21.39  2TRX 158
ATOM       8  CA ASP A  2      19.395  26.125  -0.949  1.00  21.57  2TRX 159
ATOM       9  C  ASP A  2      20.264  26.214   0.297  1.00  20.89  2TRX 160
ATOM      10  O  ASP A  2      19.760  26.575   1.371  1.00  21.49  2TRX 161
ATOM      11  CB ASP A  2      18.439  24.914  -0.856  1.00  22.14  2TRX 162
```

<http://www-structure.llnl.gov/Xray/101index.html>

NMR Spectroscopy

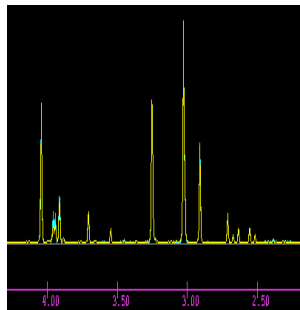


Principles of NMR



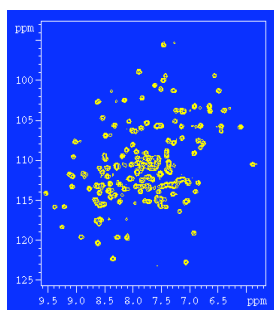
Multidimensional NMR

1D



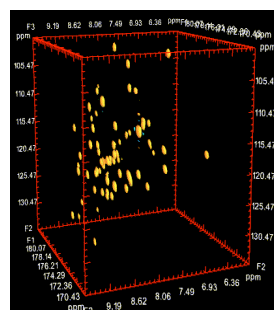
MW ~ 500

2D



MW ~ 10,000

3D

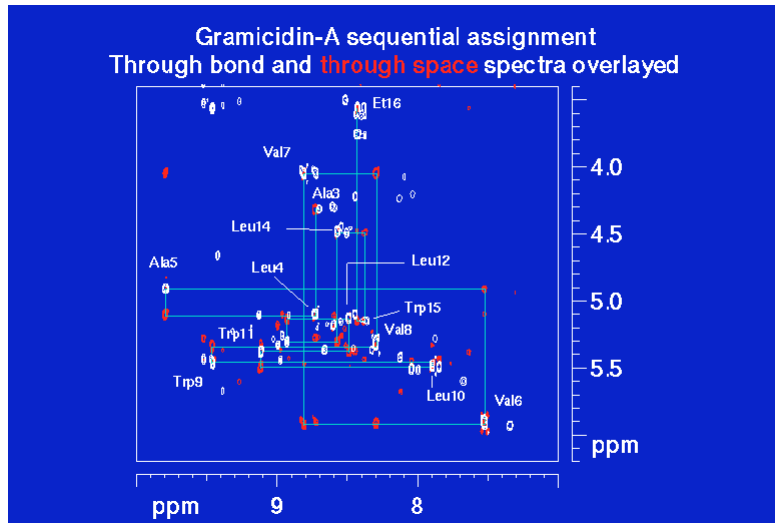


MW ~ 30,000

The NMR Process

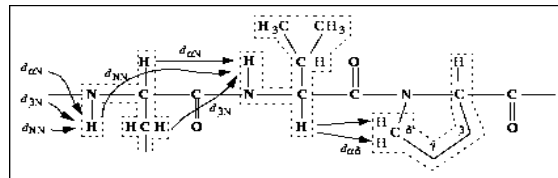
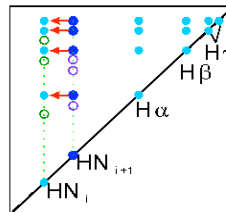
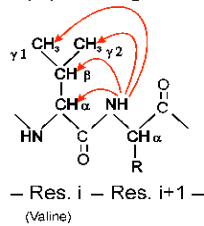
- Obtain protein sequence
- Collect TOCSY & NOESY data
- Use chemical shift tables and known sequence to assign TOCSY spectrum
- Use TOCSY to assign NOESY spectrum
- Obtain inter and intra-residue distance information from NOESY data
- Feed data to computer to solve structure

Assigning Chemical Shifts

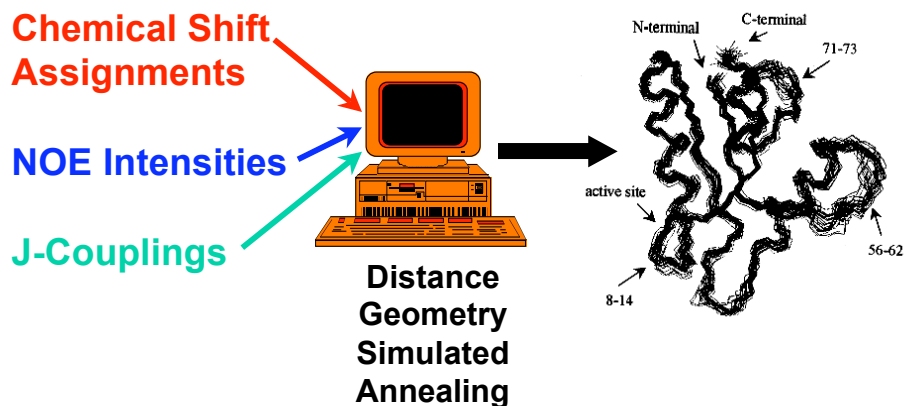


Measuring NOEs

Dipeptide Fragment



NMR Spectroscopy



The Final Result

ORIGX2	0.000000	1.000000	0.000000	0.000000	2TRX	147						
ORIGX3	0.000000	0.000000	1.000000	0.000000	2TRX	148						
SCALE1	0.011173	0.000000	0.004858	0.000000	2TRX	149						
SCALE2	0.000000	0.019585	0.000000	0.000000	2TRX	150						
SCALE3	0.000000	0.000000	0.018039	0.000000	2TRX	151						
ATOM	1	N	SER	A	1	21.389	25.406	-4.628	1.00	23.22	2TRX	152
ATOM	2	CA	SER	A	1	21.628	26.691	-3.983	1.00	24.42	2TRX	153
ATOM	3	C	SER	A	1	20.937	26.944	-2.679	1.00	24.21	2TRX	154
ATOM	4	O	SER	A	1	21.072	28.079	-2.093	1.00	24.97	2TRX	155
ATOM	5	CB	SER	A	1	21.117	27.770	-5.002	1.00	28.27	2TRX	156
ATOM	6	OG	SER	A	1	22.276	27.925	-5.861	1.00	32.61	2TRX	157
ATOM	7	N	ASP	A	2	20.173	26.028	-2.163	1.00	21.39	2TRX	158
ATOM	8	CA	ASP	A	2	19.395	26.125	-0.949	1.00	21.57	2TRX	159
ATOM	9	C	ASP	A	2	20.264	26.214	0.297	1.00	20.89	2TRX	160
ATOM	10	O	ASP	A	2	19.760	26.575	1.371	1.00	21.49	2TRX	161
ATOM	11	CB	ASP	A	2	18.439	24.914	-0.856	1.00	22.14	2TRX	162

X-ray Versus NMR

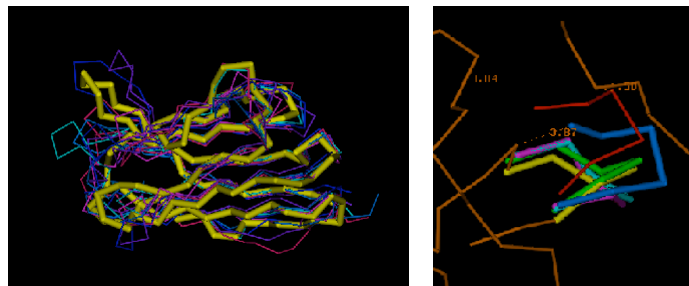
X-ray

- Producing enough protein for trials
- Crystallization time and effort
- Crystal quality, stability and size control
- Finding isomorphous derivatives
- Chain tracing & checking

NMR

- Producing enough labeled protein for collection
- Sample “conditioning”
- Size of protein
- Assignment process is slow and error prone
- Measuring NOE’s is slow and error prone

Comparative (Homology) Modelling



```
ACDEFGHIKLMNPQRST--FGHQWERT-----TYREWYEGHADS  
ASDEYAHLRILDPQRSTVAYAYE--KSFAPPGSFKWEYEAHADS  
MCDEYAHIRLMNPERSTVAGGHQWERT-----GSFKEWYAAHADD
```

Homology Modelling

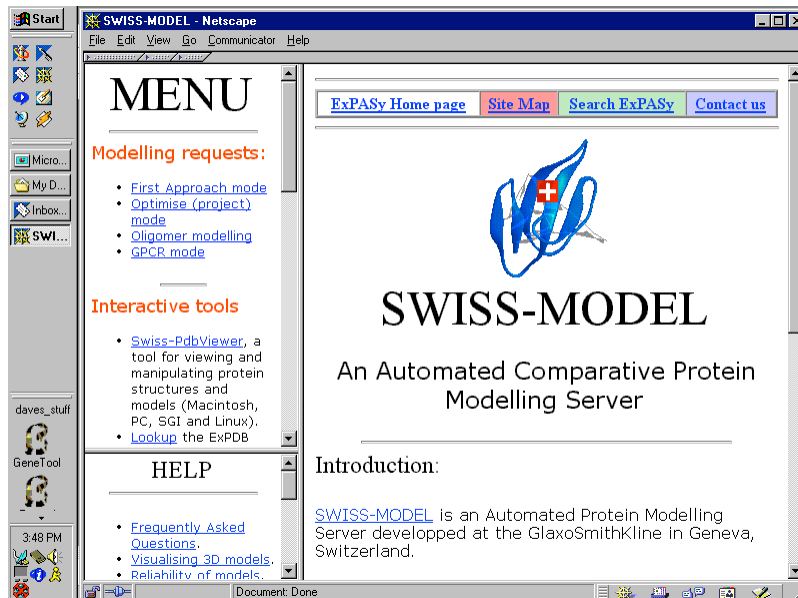
- Offers a method to “Predict” the 3D structure of proteins for which it is not possible to obtain X-ray or NMR data
- Can be used in understanding function, activity, specificity, etc.
- Of interest to drug companies wishing to do structure-aided drug design
- A keystone of Structural Proteomics

Homology Modelling

- Identify homologous sequences in PDB
- Align query sequence with homologues
- Find Structurally Conserved Regions (SCRs)
- Identify Structurally Variable Regions (SVRs)
- Generate coordinates for core region
- Generate coordinates for loops
- Add side chains (Check rotamer library)
- Refine structure using energy minimization
- Validate structure

Modelling on the Web

- **Prior to 1998 homology modelling could only be done with commercial software or command-line freeware**
- **The process was time-consuming and labor-intensive**
- **The past few years has seen an explosion in automated web-based homology modelling servers**
- **Now anyone can homology model!**



The screenshot shows a Netscape browser window displaying the SWISS-MODEL website. The browser title is "SWISS-MODEL - Netscape". The page content includes a "MENU" section with links for "Modelling requests" (First Approach mode, Optimise (project) mode, Oligomer modelling, GPCR mode) and "Interactive tools" (Swiss-PdbViewer, Lookup the ExpDB). There is also a "HELP" section with links for "Frequently Asked Questions", "Visualising 3D models", and "Reliability of models". The main content area features a blue ribbon logo with a red cross, the text "SWISS-MODEL", and the subtitle "An Automated Comparative Protein Modelling Server". Below this is an "Introduction:" section stating that SWISS-MODEL is an Automated Protein Modelling Server developed at the GlaxoSmithKline in Geneva, Switzerland. The browser's address bar shows the URL "http://www.expasy.ch/swissmod/SWISS-MODEL.html".

<http://www.expasy.ch/swissmod/SWISS-MODEL.html>

Server classes

- [Help](#)
- [Administration](#)
- [Build/check/repair model](#)
- [Structure validation](#)
- [Analyse a residue](#)
- [Protein analysis](#)
- [2-D graphics](#)
- [3-D graphics](#)
- [Hydrogen \(bonds\)](#)
- [Accessibility](#)
- [Atomic contacts](#)
- [Coordinate manipulations](#)
- [Rotamer related](#)
- [Cysteine related](#)
- [Water](#)
- [Ions](#)
- [Docking](#)
- [Crystal symmetry](#)
- [mutation prediction](#)
- [Other options](#)

Homology Modelling

Introduction
 Build a model on a template structure.

Methods
 WHAT IF will not align your sequences. You have to align the sequences before you give them to this server. Read the [notes](#) of the file formats and the [notes](#) on how this server works.
Please use the browse function rather than typing file names.

Give the template pdb-file

Give the template pir-file

Give the aligned sequence pir-file

The sequence files have to be well aligned in order to make build a sensible

<http://www.cmbi.kun.nl:1100/WIWWWI/>

The Final Result

```

ORIGX2      0.000000  1.000000  0.000000      0.000000      2TRX 147
ORIGX3      0.000000  0.000000  1.000000      0.000000      2TRX 148
SCALE1      0.011173  0.000000  0.004858      0.000000      2TRX 149
SCALE2      0.000000  0.019585  0.000000      0.000000      2TRX 150
SCALE3      0.000000  0.000000  0.018039      0.000000      2TRX 151
ATOM        1  N  SER A  1      21.389  25.406  -4.628  1.00  23.22      2TRX 152
ATOM        2  CA SER A  1      21.628  26.691  -3.983  1.00  24.42      2TRX 153
ATOM        3  C  SER A  1      20.937  26.944  -2.679  1.00  24.21      2TRX 154
ATOM        4  O  SER A  1      21.072  28.079  -2.093  1.00  24.97      2TRX 155
ATOM        5  CB SER A  1      21.117  27.770  -5.002  1.00  28.27      2TRX 156
ATOM        6  OG SER A  1      22.276  27.925  -5.861  1.00  32.61      2TRX 157
ATOM        7  N  ASP A  2      20.173  26.028  -2.163  1.00  21.39      2TRX 158
ATOM        8  CA ASP A  2      19.395  26.125  -0.949  1.00  21.57      2TRX 159
ATOM        9  C  ASP A  2      20.264  26.214   0.297  1.00  20.89      2TRX 160
ATOM       10  O  ASP A  2      19.760  26.575   1.371  1.00  21.49      2TRX 161
ATOM       11  CB ASP A  2      18.439  24.914  -0.856  1.00  22.14      2TRX 162
    
```

The PDB

- **PDB - Protein Data Bank**
- **Established in 1971 at Brookhaven National Lab (7 structures)**
- **Primary archive for macromolecular structures (proteins, nucleic acids, carbohydrates)**
- **Moved from BNL to RCSB (Research Collaboratory for Structural Bioinformatics) in 1998**

The screenshot shows the Protein Data Bank website in a Netscape browser window. The address bar displays <http://www.rcsb.org/pdb/>. The page features a navigation menu with links for [DEPOSIT data](#), [DOWNLOAD files](#), [browse LINKS](#), [BETA TEST new features](#), and [BETA XML files](#). A section titled "Current Holdings" reports 22611 structures, last updated on 23-Sep-2003. A "Molecule of the Month" section highlights the Estrogen Receptor. The main content area includes a search bar for "Search the Archive" and a list of "PDB Mirrors" from various international institutions. The footer contains the URL <http://www.rcsb.org/pdb/>.

<http://www.rcsb.org/pdb/>

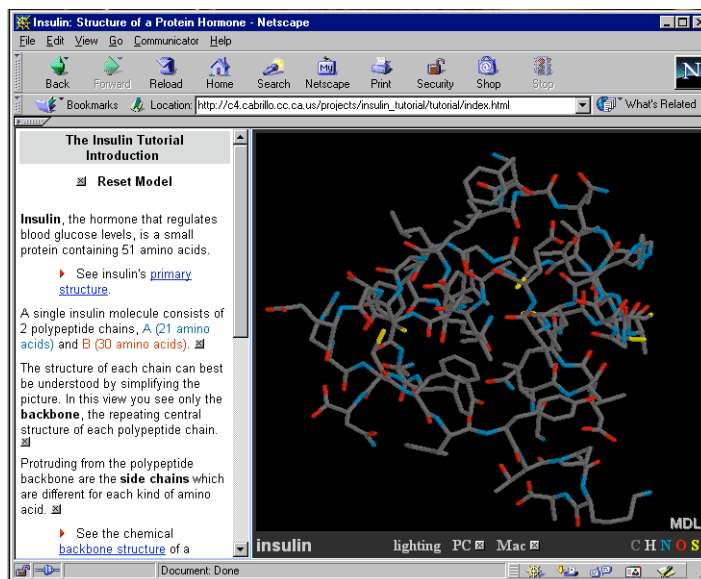
The PDB

- **Contains coordinate data (primarily) from X-ray, NMR and modelling**
- **Contains files in 2 formats**
 - PDB format
 - mmCIF (macromolecular Crystallographic Information File Format)
- **Contains 22,000+ entries**
- **Currently growing exponentially**

Viewing 3D Structures

The screenshot shows a Netscape browser window titled "Structure Explorer - 1NSW". The address bar contains the URL "http://www.rcsb.org/pdb/cgi/explorer.cgi?pdb=graphics&pdbid=1NSW". The page content is organized into a sidebar on the left and a main content area. The sidebar includes links for "View Structure", "Download/Display File", "Structural Neighbors", "Geometry", "Other Sources", "Sequence Details", and "Structure Factors (compressed)". The main content area is titled "Interactive 3D Display:" and contains a list of display options for the asymmetric unit, such as "VRML (default options)", "VRML (custom options, full screen display)", "Rasmol", "Swiss-PdbViewer", "MICE - Molecular Interactive Collaborative Environment", "FirstGlance", "Protein Explorer", "Shimura Millergram", and "Java". A "QuickPDB" button is also present. Below the list, there are sections for "Still Images:" with sub-sections for "Asymmetric Unit" and "Assumed Biological Molecule", each containing a small 3D molecular model. A "Download Help" box is visible on the right side of the main content area.

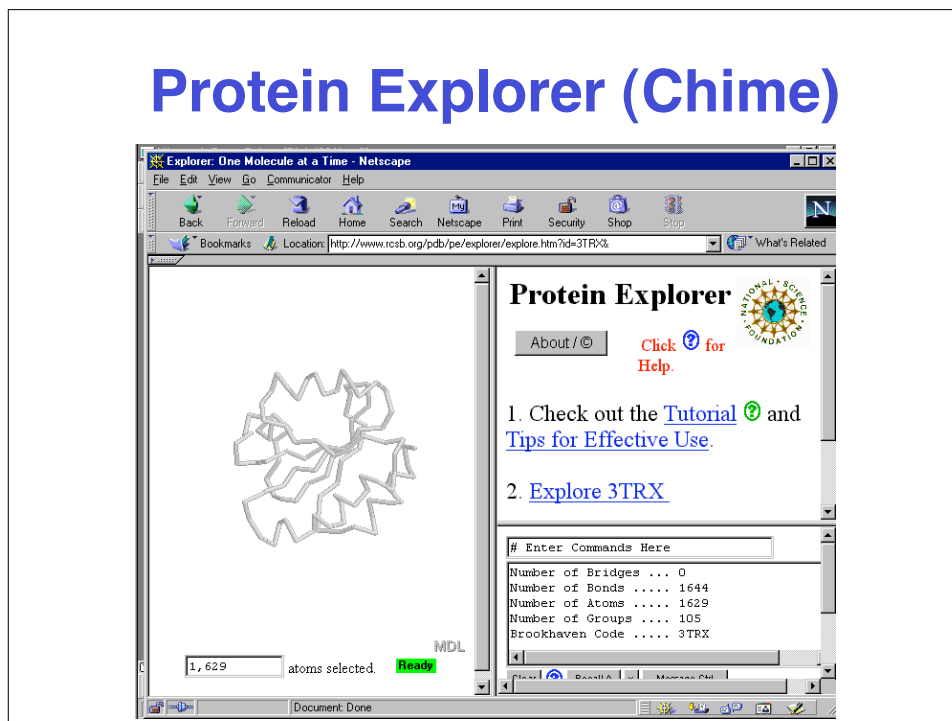
Chime



Chime

- <http://www.mdlchime.com/chime/>
- **Very simple viewing program with limited manipulation capacity**
- **Uses Rasmol for its back end source**
- **View both large and small molecules**
- **Browser Plug-in (Like PDF reader)**
- **Compatible with Netscape 4.7X and higher as well as IE 5.5 and higher**

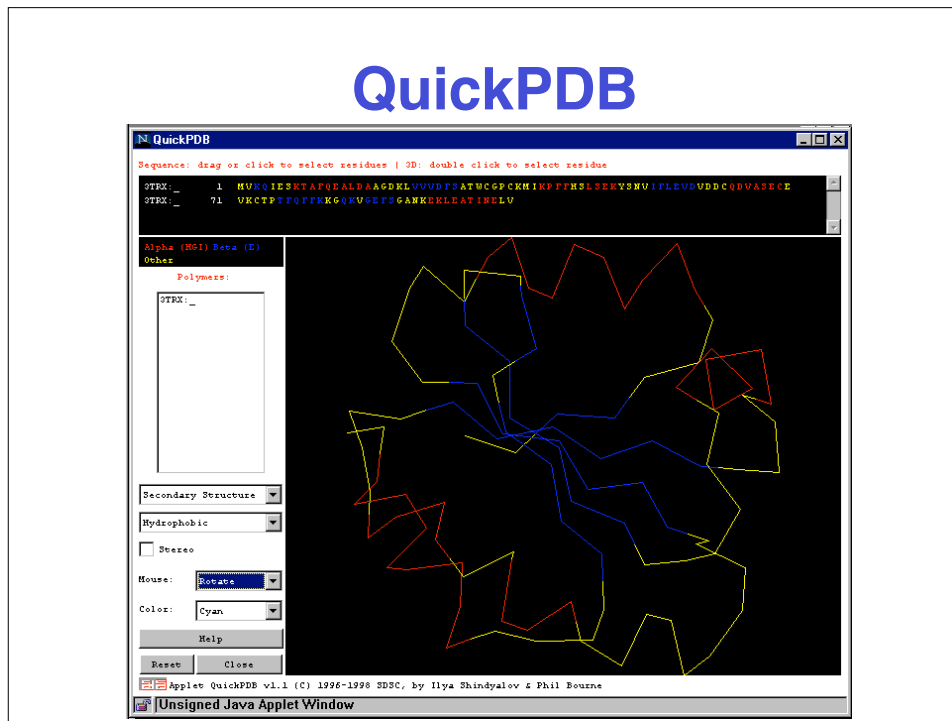
Protein Explorer (Chime)



Protein Explorer

- <http://www.umass.edu/microbio/chime/explorer/>
- **Uses Chime & Rasmol for its back-end**
- **Very flexible, user friendly, well documented, offers morphing, sequence structure interface, comparisons, context-dependent help, smart zooming, off-line**
- **Browser Plug-in (Like PDF reader)**
- **Compatible with Netscape (Mac & Win)**

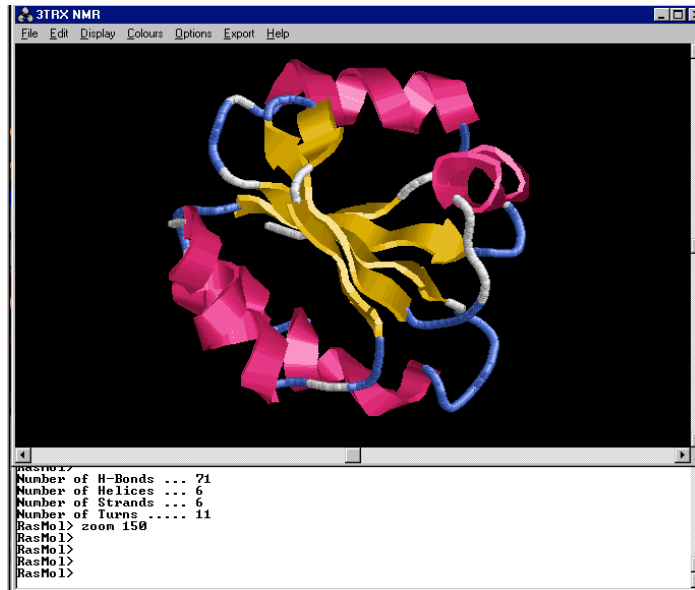
QuickPDB



Quick PDB

- <http://www.sdsc.edu/pb/Software.html>
- Very simple viewing program with limited manipulation and very limited rendering capacity -- Very fast
- Java Applet (Source code available)
- Compatible with most browsers and computer platforms

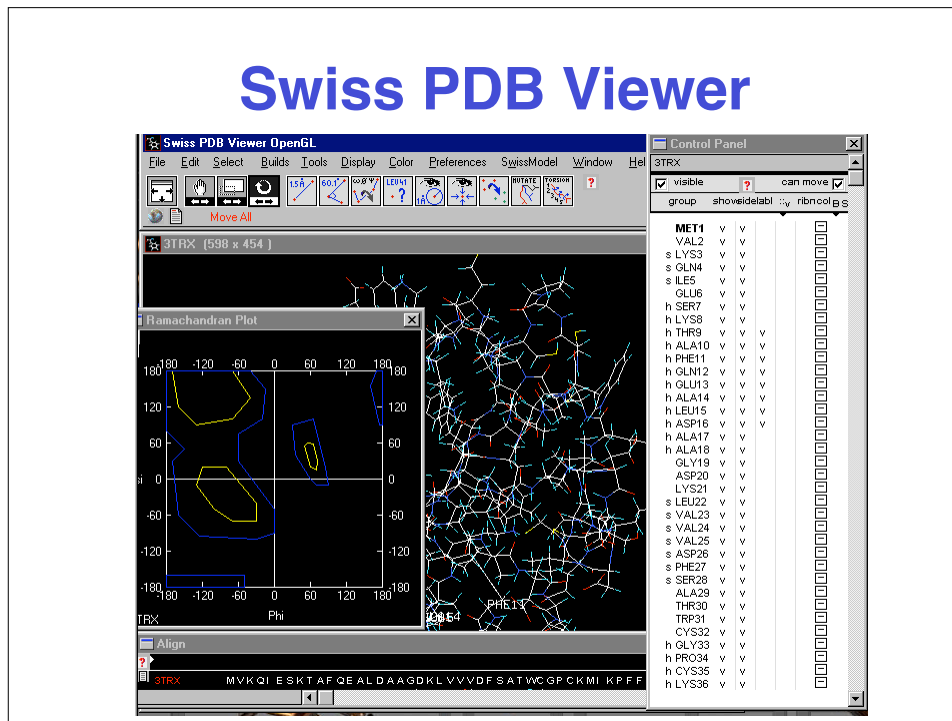
Rasmol



Rasmol

- <http://www.umass.edu/microbio/rasmol/>
- Very simple viewing program with limited manipulation capacity, easy to use!
- “Grand-daddy” of all visual freeware
- Runs as installed “stand-alone” program
- Source code available
- Runs on Mac, Windows, Linux, SGI and most other UNIX platforms

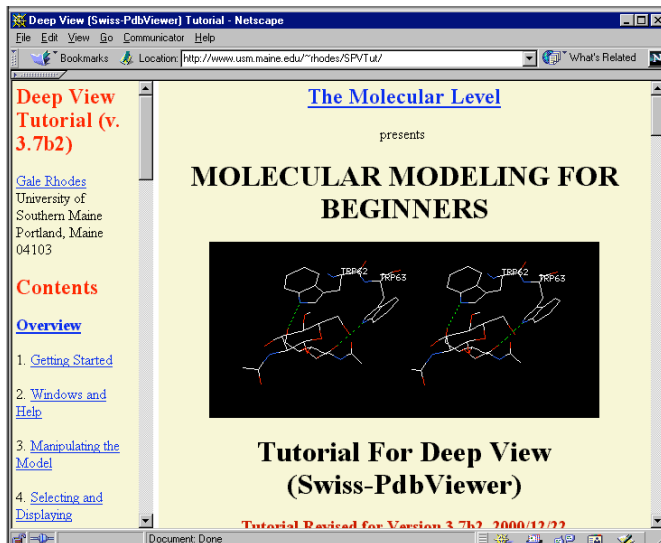
Swiss PDB Viewer



Swiss PDB Viewer

- <http://www.expasy.ch/spdbv/>
- Among most sophisticated molecular rendering, manipulation and modelling packages (commercial or freeware)
- Supports threading, hom. Modelling, energy minimization, seq/struc interface
- Stand-alone version only
- Compatible on Mac, Win, Linux, SGI

Swiss PDB Tutorial



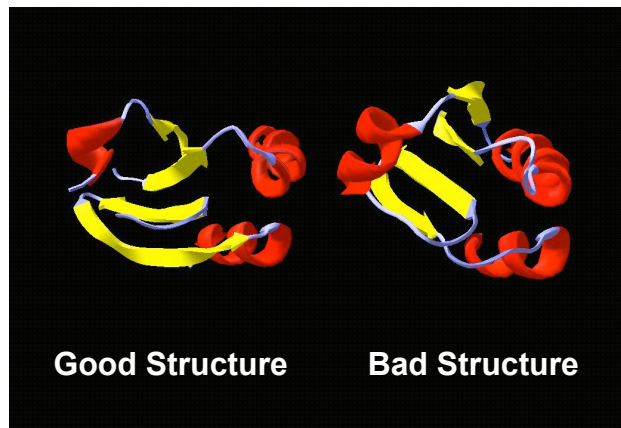
<http://www.usm.maine.edu/~rhodes/SPVTut/>

Summary

Mac Win Unix Rendr SeqView Super E Min Modeling

Rasmol	+	+	+	++	-	-	-	-
Chime	+	+	-	+	-	-	-	-
Prot. Expl.	+	+	-	++	+	+	-	-
Quick PDB	+	+	+	+	+	-	-	-
Biomer	+	+	+	++	-	+	+	+
SwP Viewer	+	+	+	+++	+	+	+	+
MolMol	-	+	+	+++	-	+	-	+

Assessing 3D Structures



Why Assess Structure?

- **A structure can (and often does) have mistakes**
- **A poor structure will lead to poor models of mechanism or relationship**
- **Unusual parts of a structure may indicate something important (or an error)**

Famous “bad” structures

- **Azobacter ferredoxin (wrong space group)**
- **Zn-metallothionein (mistraced chain)**
- **Alpha bungarotoxin (poor stereochemistry)**
- **Yeast enolase (mistraced chain)**
- **Ras P21 oncogene (mistraced chain)**
- **Gene V protein (poor stereochemistry)**

How to Assess Structure?

- **Assess experimental fit (look at R factor {X-ray} or rmsd {NMR})**
- **Assess correctness of overall fold (look at disposition of hydrophobes, location of charged residues)**
- **Assess structure quality (packing, stereochemistry, bad contacts, etc.)**

A Good Protein Structure..

X-ray structure

- R = 0.59 random chain
- R = 0.45 initial structure
- R = 0.35 getting there
- R = 0.25 typical protein
- R = 0.15 best case
- R = 0.05 small molecule

NMR structure

- rmsd = 4 Å random
- rmsd = 2 Å initial fit
- rmsd = 1.5 Å OK
- rmsd = 0.8 Å typical
- rmsd = 0.4 Å best case
- rmsd = 0.2 Å dream on

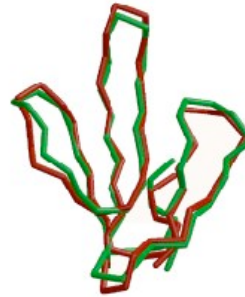
Cautions...

- A low R factor or a good RMSD value does not guarantee that the structure is “right”
- Differences due to crystallization conditions, crystal packing, solvent conditions, concentration effects, etc. can perturb structures substantially
- Long recognized need to find other ways to ID good structures from bad (not just assessing experimental fit)

Structure Variability



X-ray to X-ray
Interleukin 1□
(41bi vs 2mlb)



NMR to X-ray
Erabutoxin
(3ebx vs 1era)

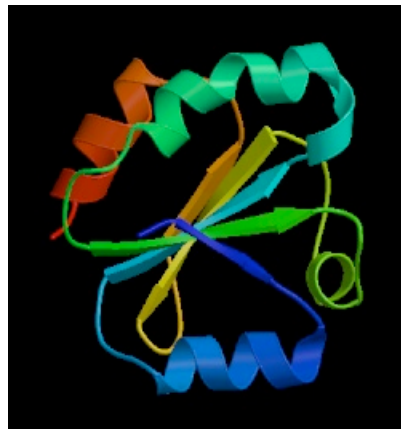
A Good Protein Structure..

- **Minimizes disallowed torsion angles**
- **Maximizes number of hydrogen bonds**
- **Maximizes buried hydrophobic ASA**
- **Maximizes exposed hydrophilic ASA**
- **Minimizes interstitial cavities or spaces**



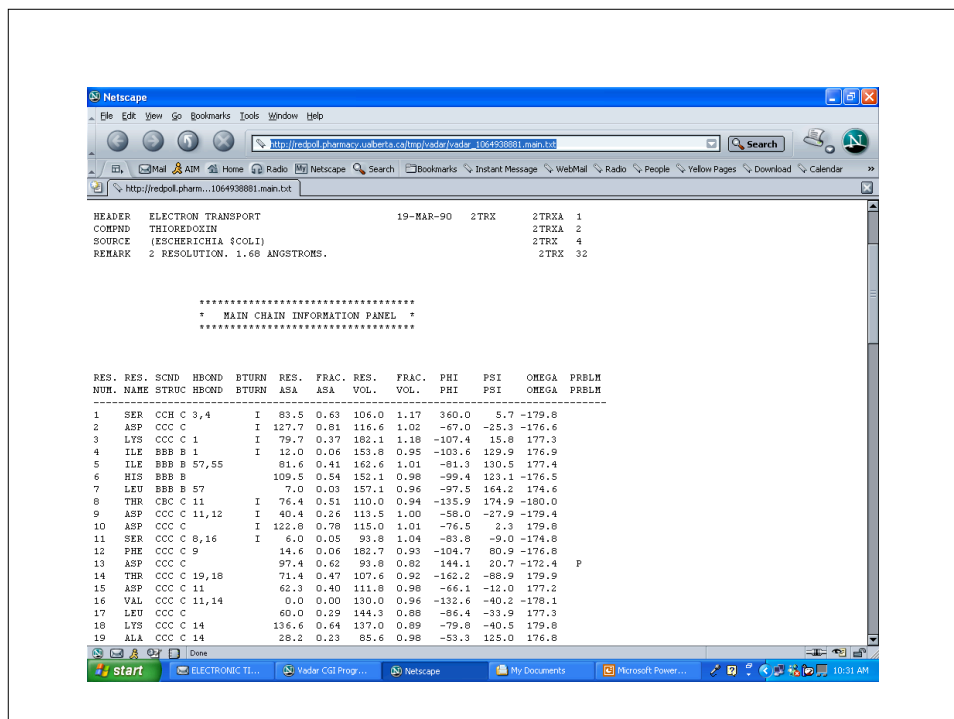
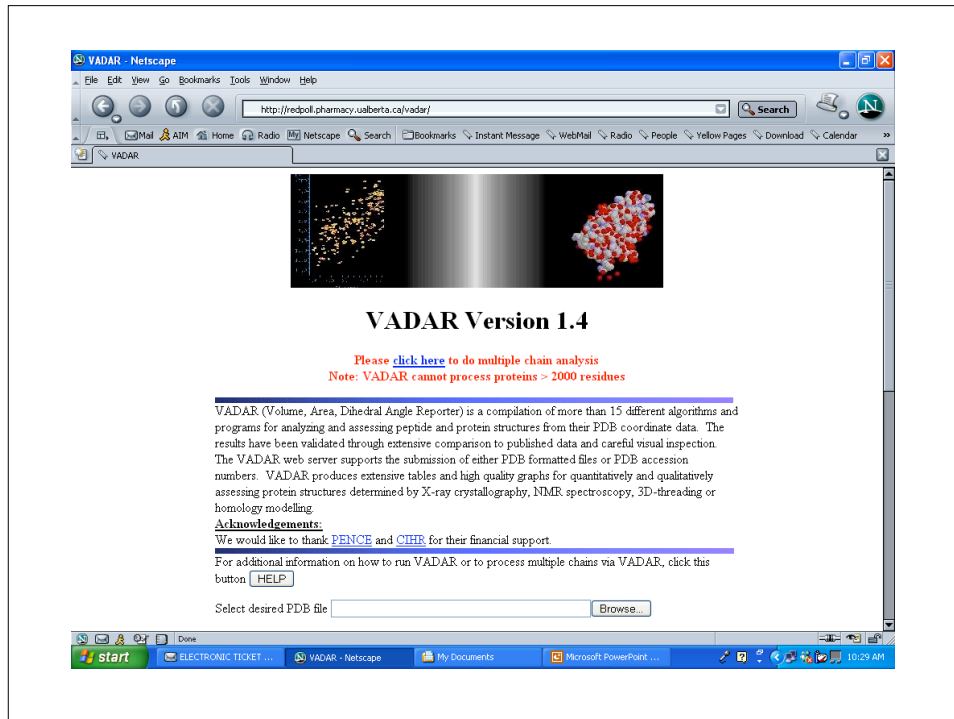
A Good Protein Structure..

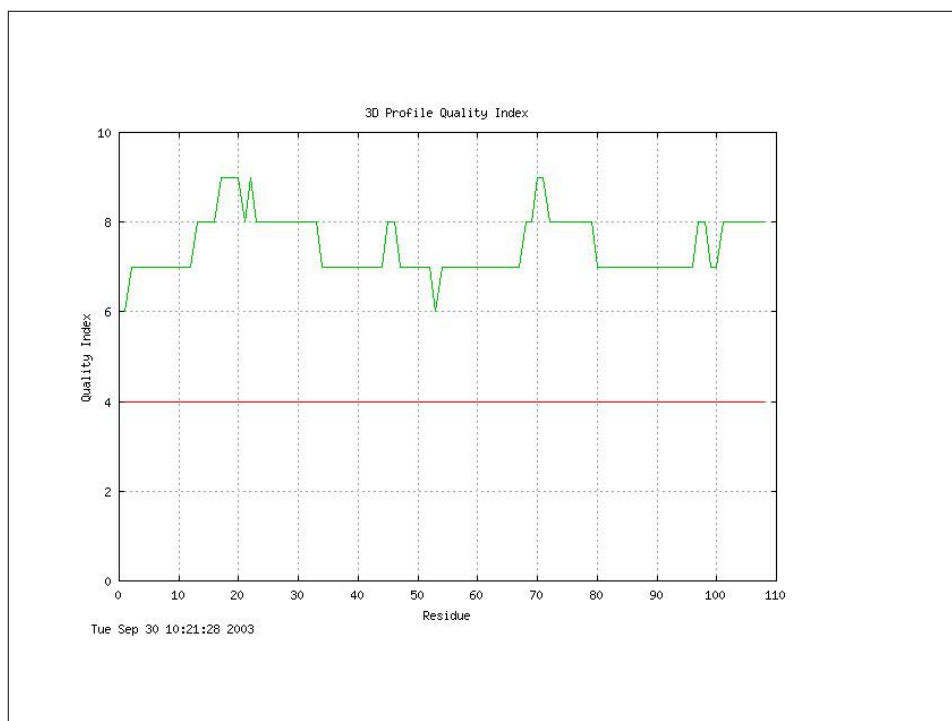
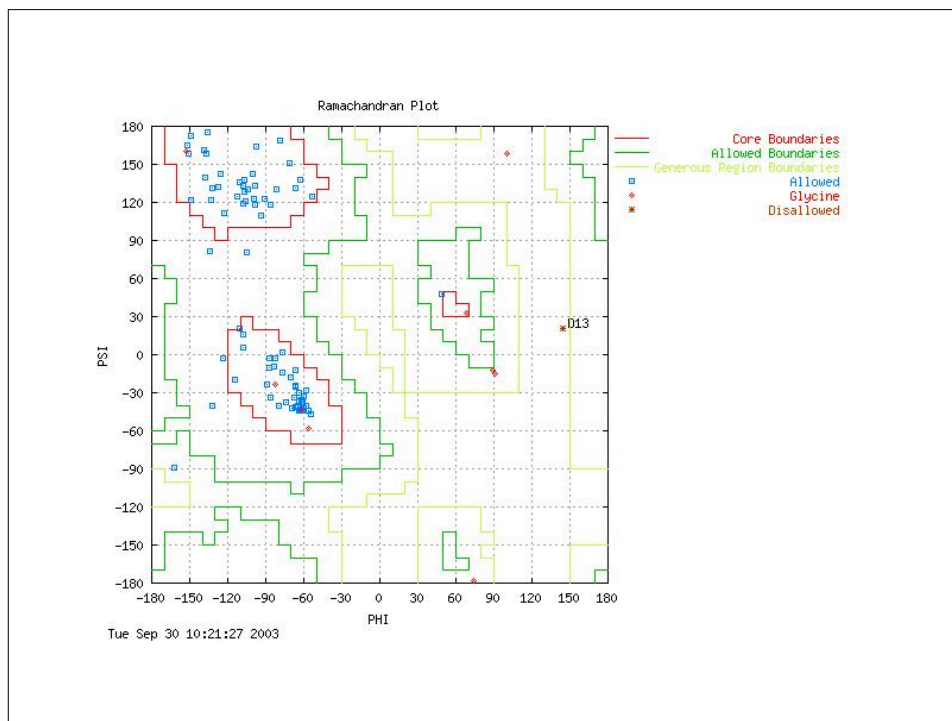
- Minimizes number of “bad” contacts
- Minimizes number of buried charges
- Minimizes radius of gyration
- Minimizes covalent and noncovalent (van der Waals and coulombic) energies



Structure Validation Servers

- **WhatIf Web Server** -
<http://www.cmbi.kun.nl:1100/WIWWWI/>
- **Biotech Validation Suite** -
<http://biotech.ebi.ac.uk:8400/cgi-bin/sendquery>
- **Verify3D** -
http://www.doe-mpi.ucla.edu/Services/Verify_3D/
- **VADAR** - <http://redpoll.pharmacy.ualberta.ca>

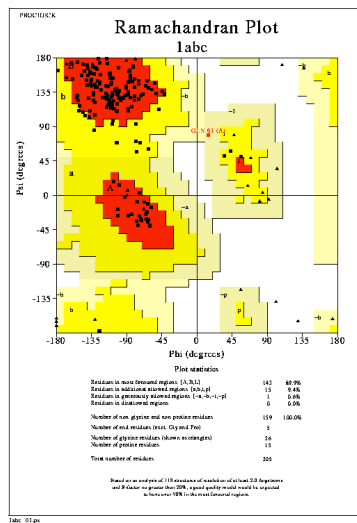
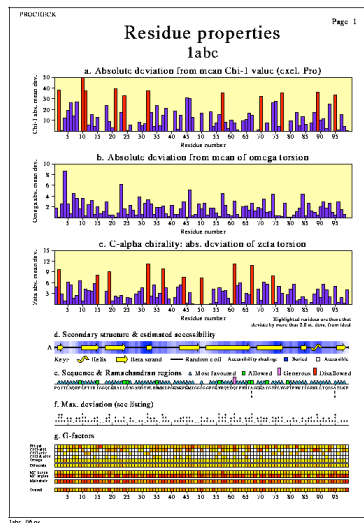




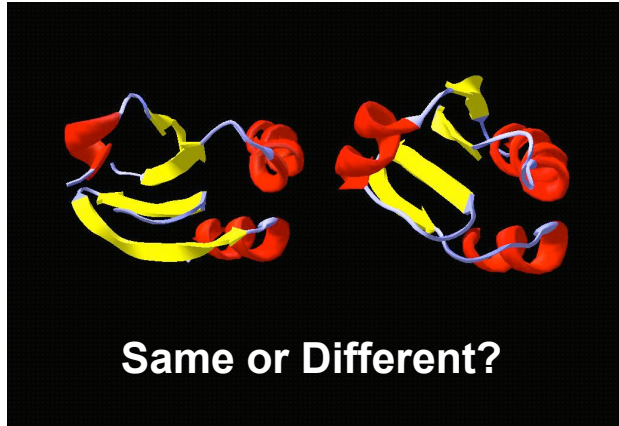
Structure Validation Programs

- **PROCHECK** -
<http://www.biochem.ucl.ac.uk/~roman/procheck/procheck.html>
- **PROSA II** -
<http://lore.came.sbg.ac.at/People/mo/Prosa/prosa.html>
- **VADAR** -
<http://www.pence.ualberta.ca/ftp/vadar/>
- **DSSP** -
<http://www.embl-heidelberg.de/dssp/>

Procheck



Comparing 3D Structures



Same or Different?

Qualitative vs. Quantitative

Rigid Body Superposition



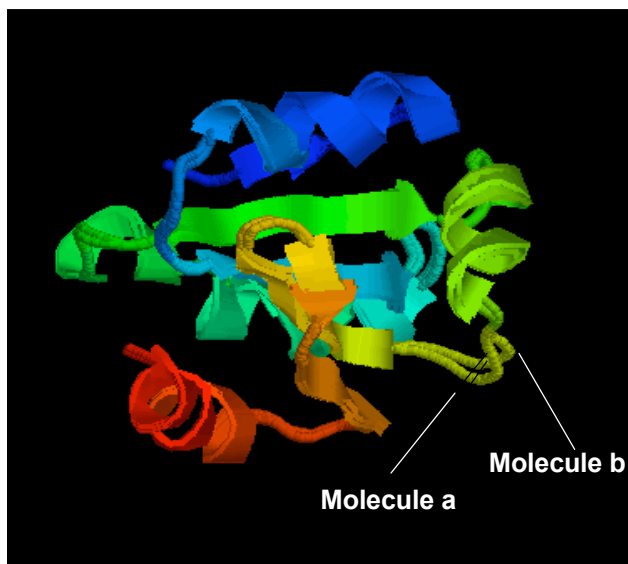
Superposition

- Objective is to match or overlay 2 or more similar objects
- Requires use of translation and rotation operators (matrices/vectors)
- Least squares or conjugate gradient minimization (McLachlan/Kabsch)
- Lagrangian multipliers
- Quaternion-based methods (*fastest*)

Superposition - Applications

- Ideal for comparing or overlaying two or more protein structures
- Allows identification of structural homologues (CATH and SCOP)
- Allows loops to be inserted or replaced from loop libraries (comparative modelling)
- Allows side chains to be replaced or inserted with relative ease

Measuring Superpositions



RMSD - Root Mean Square Deviation

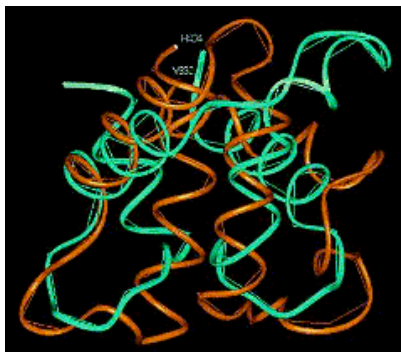
- Method to quantify structural similarity - same as standard deviation
- Requires 2 superimposed structures (designated here as “a” & “b”)
- N = number of atoms being compared

$$\text{RMSD} = \frac{\sqrt{\sum_i (x_{ai} - x_{bi})^2 + (y_{ai} - y_{bi})^2 + (z_{ai} - z_{bi})^2}}{\sqrt{N}}$$

RMSD

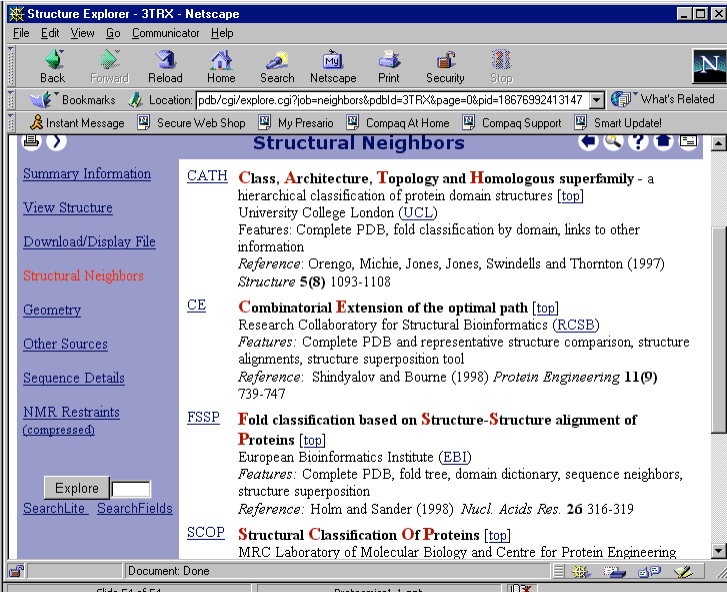
- 0.0-0.5 Å → Essentially identical
- <1.5 Å → Very good fit
- < 5.0 Å → Moderately good fit
- 5.0-7.0 Å → Structurally related
- > 7.0 Å → Dubious relationship
- > 12.0 Å → Completely unrelated

Detecting Unusual Relationships



Similarity between Calmodulin and Acetylcholinesterase

Classifying Protein Folds



Structural Neighbors

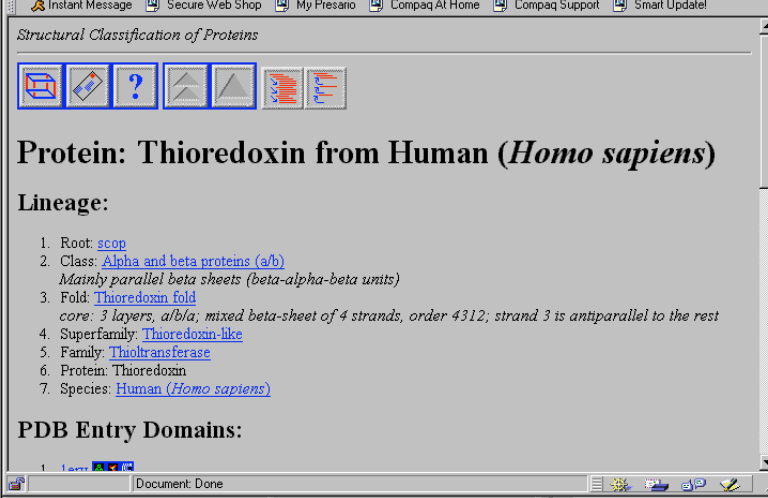
CATH **C**lass, **A**rchitecture, **T**opology and **H**omologous superfamily - a hierarchical classification of protein domain structures [top]
University College London (UCL)
Features: Complete PDB, fold classification by domain, links to other information
Reference: Orengo, Michie, Jones, Jones, Swindells and Thornton (1997) *Structure* **5**(8) 1093-1108

CE **C**ombinatorial **E**xtension of the optimal path [top]
Research Collaboratory for Structural Bioinformatics (RCSE)
Features: Complete PDB and representative structure comparison, structure alignments, structure superposition tool
Reference: Shindyalov and Bourne (1998) *Protein Engineering* **11**(9) 739-747

FSSP **F**old classification based on **S**tructure-**S**tructure alignment of **P**roteins [top]
European Bioinformatics Institute (EBI)
Features: Complete PDB, fold tree, domain dictionary, sequence neighbors, structure superposition
Reference: Holm and Sander (1998) *Nucl. Acids Res.* **26** 316-319

SCOP **S**tructural **C**lassification **O**f **P**roteins [top]
MRC Laboratory of Molecular Biology and Centre for Protein Engineering

SCOP Database



Structural Classification of Proteins

Protein: Thioredoxin from Human (*Homo sapiens*)

Lineage:

1. Root: [scop](#)
2. Class: [Alpha and beta proteins \(a/b\)](#)
Mainly parallel beta sheets (beta-alpha-beta units)
3. Fold: [Thioredoxin fold](#)
core: 3 layers, a/b/a; mixed beta-sheet of 4 strands, order 4312; strand 3 is antiparallel to the rest
4. Superfamily: [Thioredoxin-like](#)
5. Family: [Thiolytransferase](#)
6. Protein: Thioredoxin
7. Species: [Human \(*Homo sapiens*\)](#)

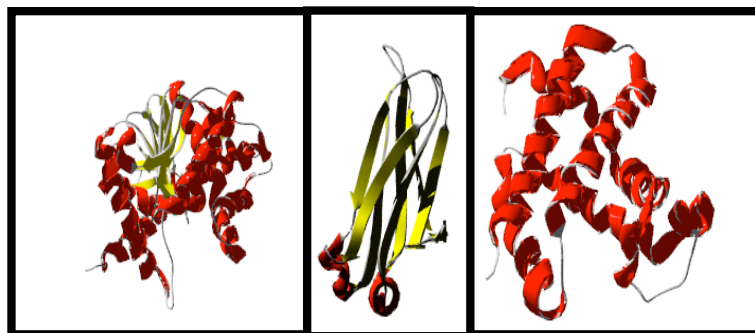
PDB Entry Domains:

<http://scop.mrc-lmb.cam.ac.uk/scop>

SCOP

- **Class** folding class derived from secondary structure content
- **Fold** derived from topological connection, orientation, arrangement and # 2° structures
- **Superfamily** clusters of low sequence ID but related structures & functions
- **Family** clusters of proteins with seq ID > 30% with v. similar struct. & function

Different Folding Classes

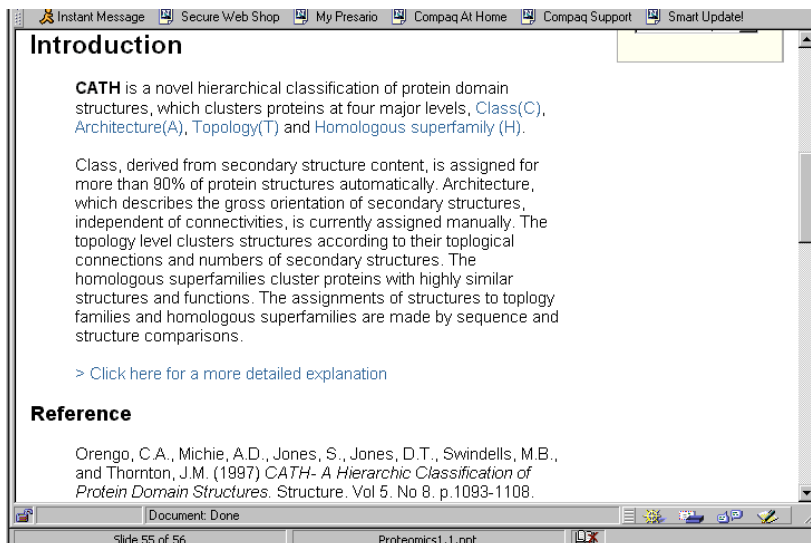


Lactate
Dehydrogenase:
Mixed /

Immunoglobulin
Fold:

Hemoglobin B
Chain:

CATH Database



The screenshot shows a web browser window with the title "Introduction" and the following text:

CATH is a novel hierarchical classification of protein domain structures, which clusters proteins at four major levels, **Class(C)**, **Architecture(A)**, **Topology(T)** and **Homologous superfamily (H)**.

Class, derived from secondary structure content, is assigned for more than 90% of protein structures automatically. Architecture, which describes the gross orientation of secondary structures, independent of connectivities, is currently assigned manually. The topology level clusters structures according to their topological connections and numbers of secondary structures. The homologous superfamilies cluster proteins with highly similar structures and functions. The assignments of structures to topology families and homologous superfamilies are made by sequence and structure comparisons.

> [Click here for a more detailed explanation](#)

Reference

Orengo, C.A., Michie, A.D., Jones, S., Jones, D.T., Swindells, M.B., and Thornton, J.M. (1997) *CATH- A Hierarchic Classification of Protein Domain Structures*. Structure. Vol 5. No 8. p.1093-1108.

The browser window also shows a taskbar with "Document: Done", "Slide 55 of 56", and "Proteomics1.1.ppt".

<http://www.biochem.ucl.ac.uk/bsm/cath/>

CATH

- **Class [C]** derived from secondary structure content (automatic)
- **Architecture (A)** derived from orientation of 2° structures (manual)
- **Topology (T)** derived from topological connection and # 2° structures
- **Homologous Superfamily (H)** clusters of similar structures & functions

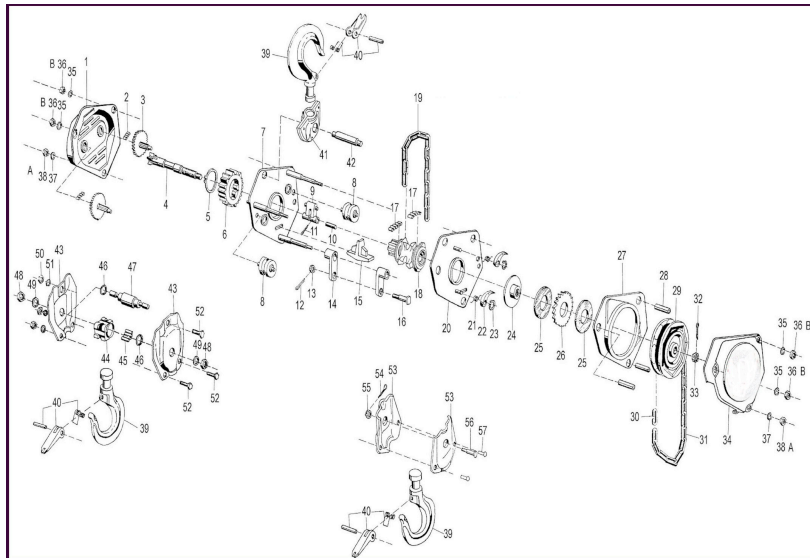
Other Servers/Databases

- **Dali** - <http://www.ebi.ac.uk/dali/>
- **VAST** - www.ncbi.nlm.nih.gov/Structure/VAST/vast.shtml
- **CE** - <http://cl.sdsc.edu/ce.html>
- **FSSP** - <http://www.ebi.ac.uk/dali/fssp/fssp.html>
- **PDBsum** - www.biochem.ucl.ac.uk/bsm/pdbsum/

Protein Interactions



The Protein Parts List



The Parts List

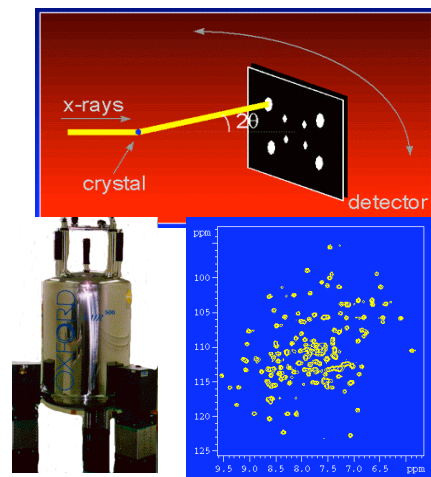
- Sequencing gives “serial number”
- Sequence alignment gives a name
- Microarrays give # of parts
- X-ray and NMR give a picture
- However, having a collection of parts and names doesn't tell you how to put something together or how things connect -- *this is biology*

Types of Interactions

- **Permanent (quaternary structure, formation of stable complexes)**
- **Transient (brief interactions, signaling events, pathways)**
- **About 1/4 to 1/3 of all proteins form complexes (dimers → multimers)**
- **Each protein may transiently interact with ~3 other proteins**

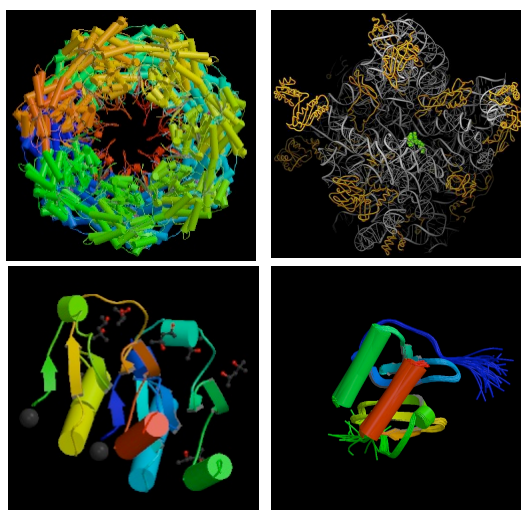
Protein Interaction Tools and Techniques - Experimental Methods

3D Structure Determination



- **X-ray crystallography**
 - grow crystal
 - collect diffract. data
 - calculate e- density
 - trace chain
- **NMR spectroscopy**
 - label protein
 - collect NMR spectra
 - assign spectra & NOEs
 - calculate structure using distance geom.

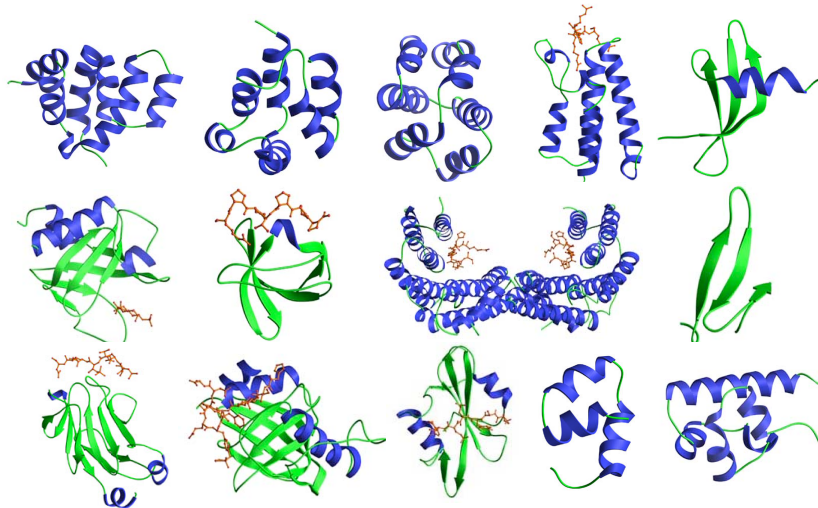
Quaternary Structure



Some interactions
are real

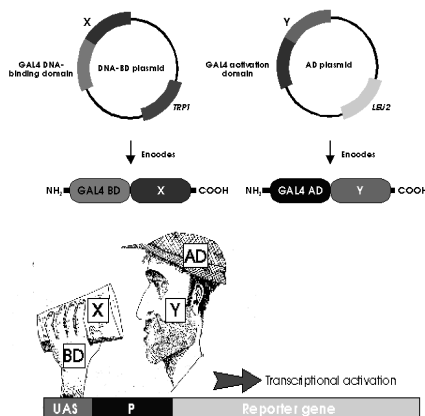
Others are not

Protein Interaction Domains



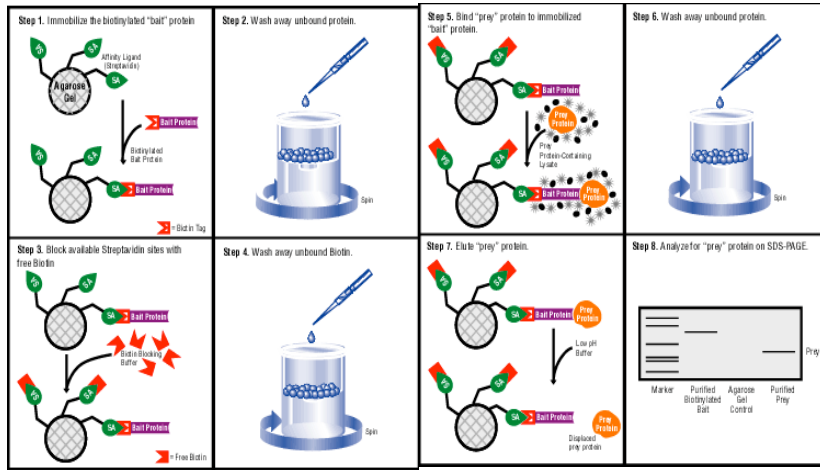
<http://www.mshri.on.ca/pawson/domains.html>

Yeast Two-Hybrid Analysis

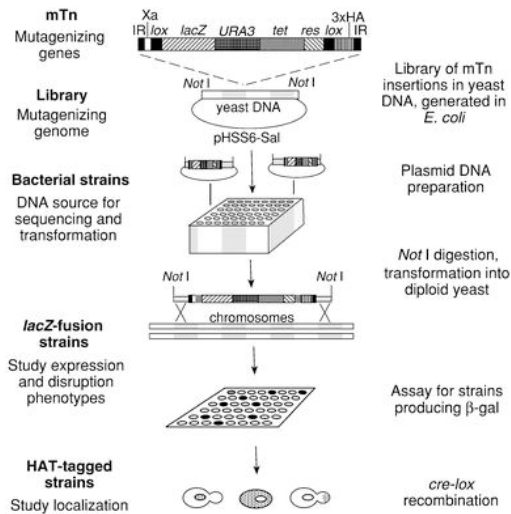


- Yeast two-hybrid experiments yield information on protein protein interactions
- GAL4 Binding Domain
- GAL4 Activation Domain
- X and Y are two proteins of interest
- If X & Y interact then reporter gene is expressed

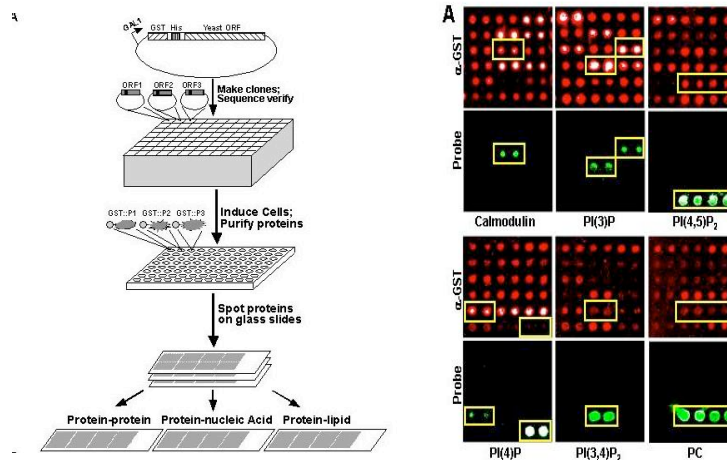
Affinity Pull-down



Transposon Tagging

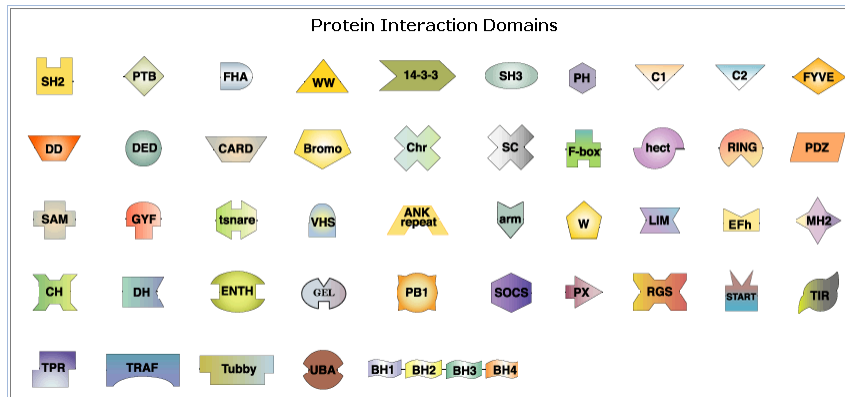


Protein Arrays



Protein Interaction Tools and Techniques - Computational Methods

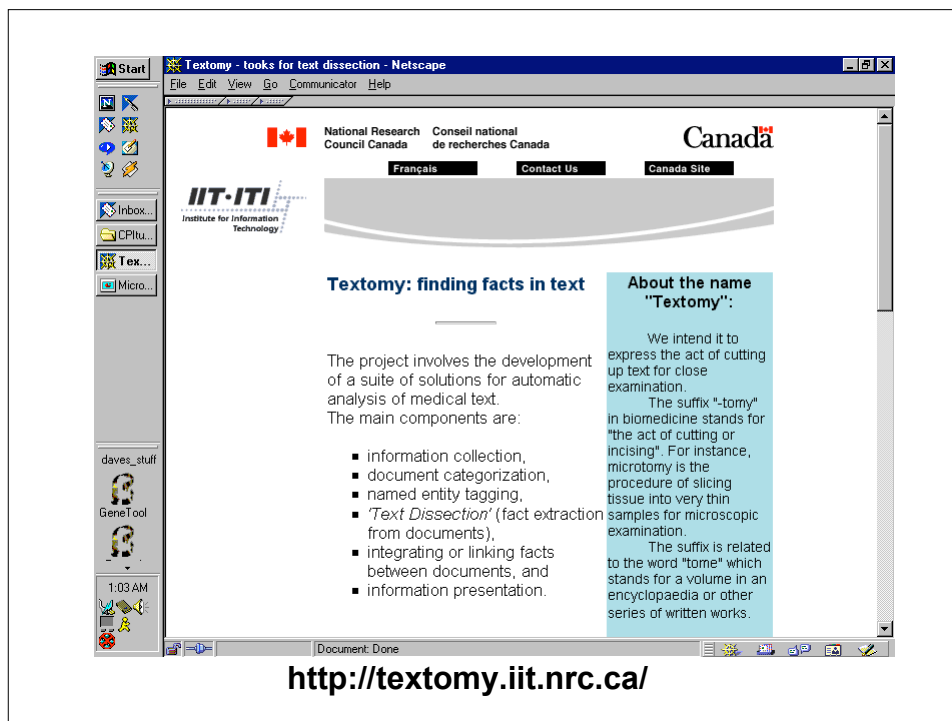
Sequence Searching Against Known Domains



<http://www.mshri.on.ca/pawson/domains.html>

Text Mining

- Searching Medline or Pubmed for words or word combinations
- “X binds to Y”; “X interacts with Y”; “X associates with Y” etc. etc.
- Requires a list of known gene names or protein names for a given organism
- Sometimes called “Textomy”



Pre-BIND

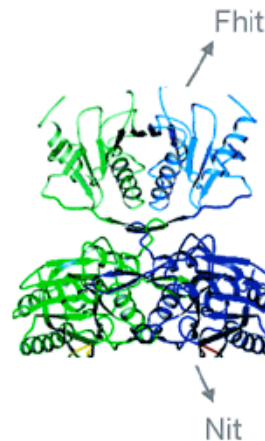
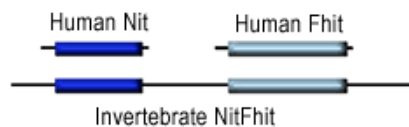
- *Donaldson et al. BMC Bioinformatics 2003 4:11*
- **Used Support Vector Machine (SVM) to scan literature for protein interactions**
- **Precision, accuracy and recall of 92% for correctly classifying PI abstracts**
- **Estimated to capture 60% of all abstracted protein interactions for a given organism**

Rosetta Stone Method

Monomeric proteins that are fused in other organisms tend to be functionally related and physically interacting.

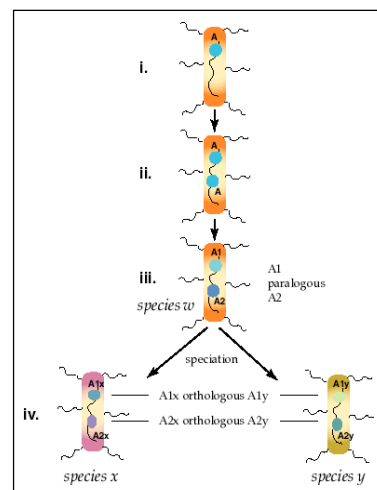
For example, using the Rosetta Stone™ method, it was found that human Nit and Fhit proteins are:

- fused in invertebrates
- form a heterocomplex in mammals



Interologs, Homologs, Paralogs...

- **Homolog**
 - Common Ancestors
 - Common 3D Structure
 - Common Active Sites
- **Ortholog**
 - Derived from Speciation
- **Paralog**
 - Derived from Duplication
- **Interolog**
 - Protein-Protein Interaction



A Flood of Data

- High throughput techniques are leading to more and more data on protein interactions
- This is where bioinformatics can play a key role
- Some suggest that this is the “future” for bioinformatics

Interaction Databases

- **BIND**
 - <http://www.bind.ca/>
- **DIP**
 - <http://dip.doe-mbi.ucla.edu/>
- **PIM**
 - <http://www.hybrigenics.fr/>
- **PathCalling**
 - <http://portal.curagen.com/expectpc/com.curagen.portal.servlet.Yeast>

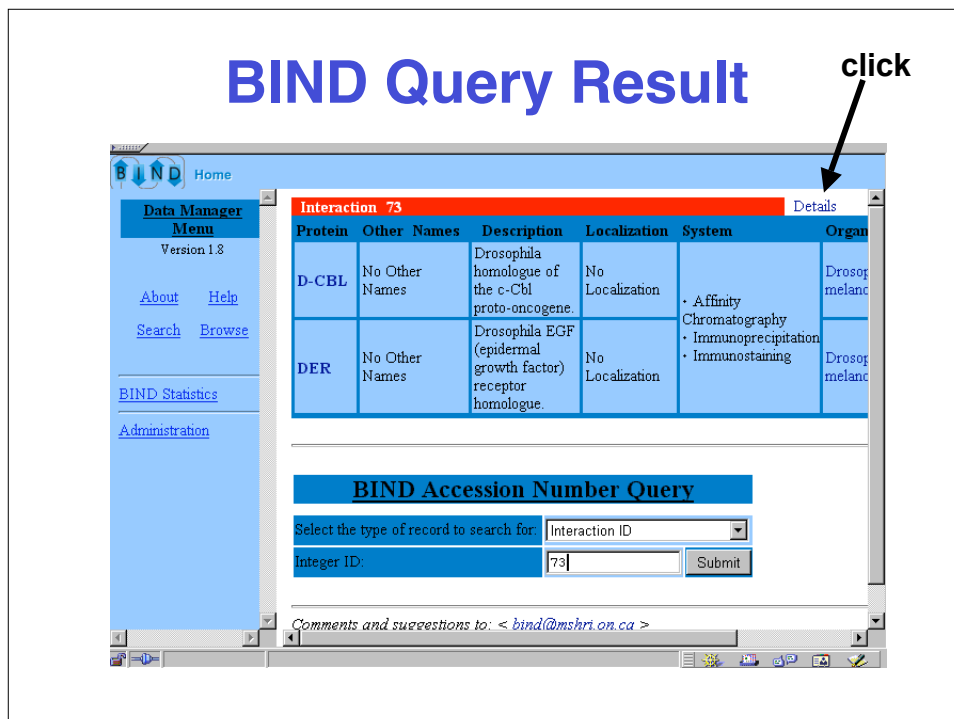


The BIND Database

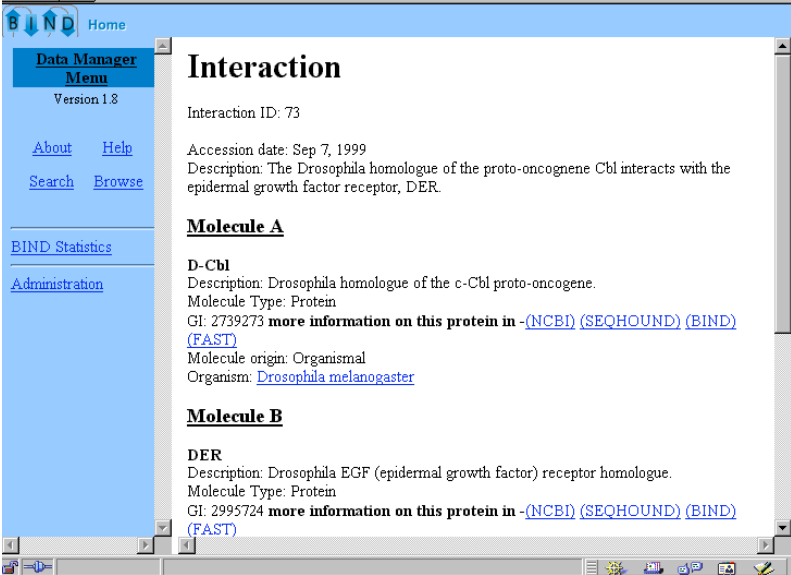
- **BIND - Biomolecular Interaction Network Database**
- **Conceived and Developed by Chris Hogue, Tony Pawson, Francis Ouellette**
- **Designed to capture almost all interactions between biomolecules (large and small)**
- **Largest database of its kind**

BIND Can Encode...

- **Simple binary interactions**
- **Enzymes, substrates and conformational changes**
- **Restriction enzymes**
- **Limited proteolysis**
- **Phosphorylation (reversible)**
- **Glycosylation**
- **Intron splicing**
- **Transcriptional factors**



BIND Details



The screenshot shows the BIND website interface. On the left is a navigation menu with links for 'Data Manager Menu', 'About', 'Help', 'Search', 'Browse', 'BIND Statistics', and 'Administration'. The main content area is titled 'Interaction' and displays the following information:

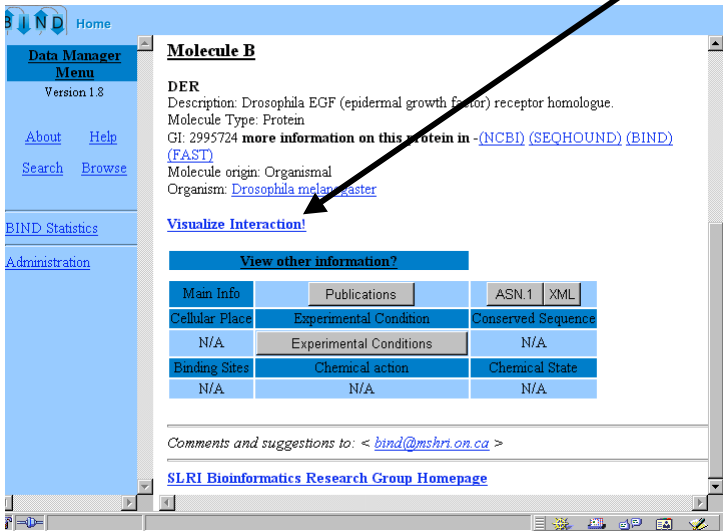
Interaction ID: 73
Accession date: Sep 7, 1999
Description: The Drosophila homologue of the proto-oncogene Cbl interacts with the epidermal growth factor receptor, DER.

Molecule A
D-Cbl
Description: Drosophila homologue of the c-Cbl proto-oncogene.
Molecule Type: Protein
GI: 2739273 [more information on this protein in - \(NCBI\) \(SEQHOUND\) \(BIND\) \(FAST\)](#)
Molecule origin: Organismal
Organism: [Drosophila melanogaster](#)

Molecule B
DER
Description: Drosophila EGF (epidermal growth factor) receptor homologue.
Molecule Type: Protein
GI: 2995724 [more information on this protein in - \(NCBI\) \(SEQHOUND\) \(BIND\) \(FAST\)](#)

BIND Details

click



The screenshot shows the BIND website interface for Molecule B. The left navigation menu is the same as in the previous image. The main content area is titled 'Molecule B' and displays the following information:

DER
Description: Drosophila EGF (epidermal growth factor) receptor homologue.
Molecule Type: Protein
GI: 2995724 [more information on this protein in - \(NCBI\) \(SEQHOUND\) \(BIND\) \(FAST\)](#)
Molecule origin: Organismal
Organism: [Drosophila melanogaster](#)

[Visualize Interaction!](#)

View other information?

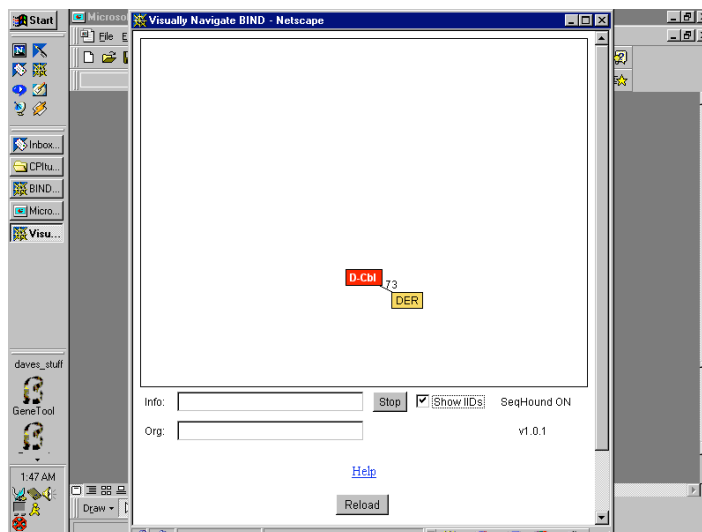
Man Info	Publications	ASN.1	XML
Cellular Place	Experimental Condition	Conserved Sequence	
N/A	Experimental Conditions	N/A	
Binding Sites	Chemical action	Chemical State	
N/A	N/A	N/A	

Comments and suggestions to: < bind@mshri.on.ca >

[SLRI Bioinformatics Research Group Homepage](#)

An arrow labeled 'click' points to the 'Visualize Interaction!' link.

BIND Details



Summary

- **First application of bioinformatics was probably in protein structure (the PDB)**
- **Structural biology continues to be a rich source for bioinformatics innovation and bioinformaticians**
- **Next “big” step in bioinformatics is to go from the “parts list” to figuring out how to put it all together**