Current Topics in Genome Analysis **Spring 2010**

Week 3: Biological Sequence Analysis II Andy Baxevanis, Ph.D.

Overview

- Week 2
 - Similarity vs. Homology
 - Global vs. Local Alignments

- Scoring Matrices
- BLAST
- BLAT
- Week 3
 - Profiles, Patterns, Motifs, and Domains
 - Structures: VAST, Cn3D, and de novo Prediction
 - Multiple Sequence Alignment

Sequence Comparisons

- Homology searches
 - Usually "one-against-one"

BLAST, FASTA

- Allows for comparison of individual sequences against databases comprised of individual sequences
- Profile searches
 - Uses collective characteristics of a family of proteins
 - Search can be "one-against-many" Pfam, InterPro,

CDD

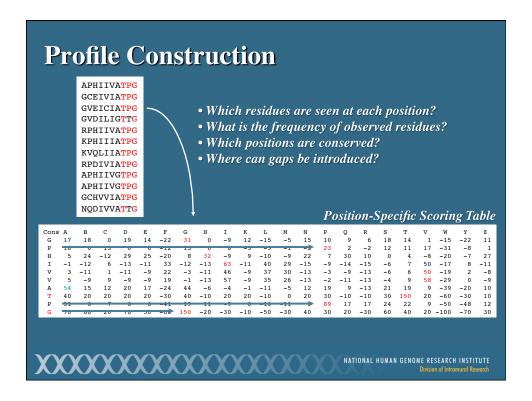
or "many-against-one"

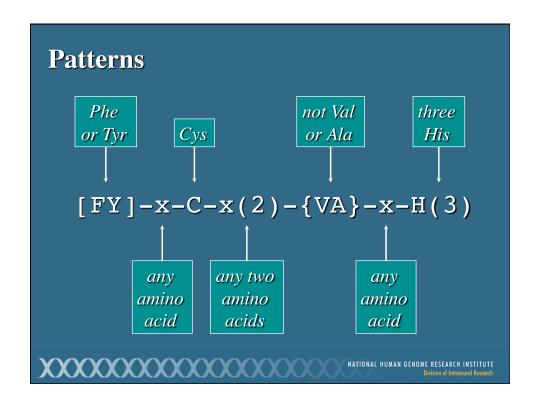
PSI-BLAST

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Profiles

- Numerical representations of multiple sequence alignments
- Depend upon patterns or motifs containing conserved residues
- Represent the common characteristics of a protein family
- Can find similarities between sequences with little or no sequence identity
- Allow for the analysis of distantly-related proteins





Pfam

- Collection of multiple alignments of protein domains and conserved protein regions (regions which probably have structural or functional importance)
- Each Pfam entry contains:
 - Multiple sequence alignment of family members
 - Protein domain architectures

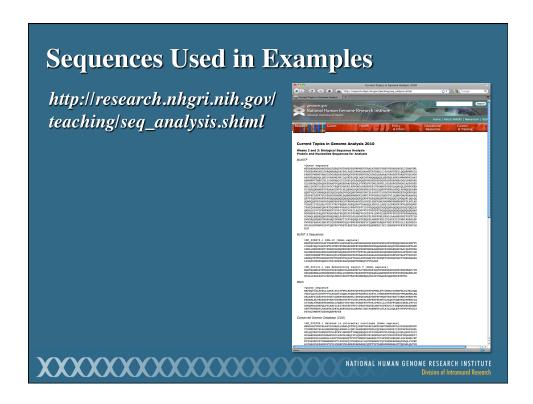
- Species distribution of family members
- Information on known protein structures
- Links to other protein family databases

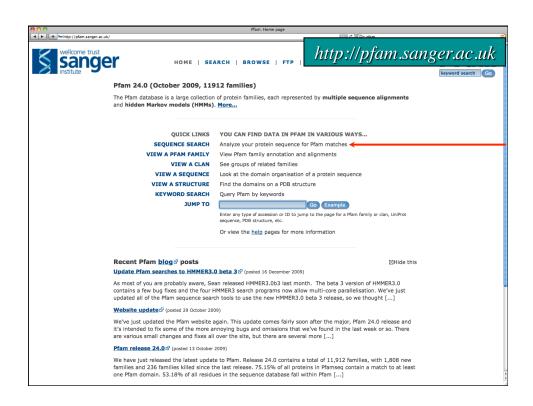
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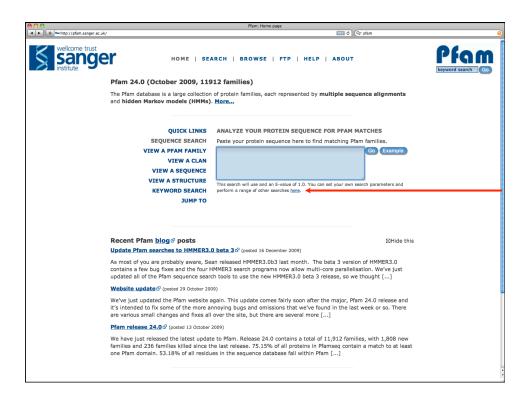
Pfam

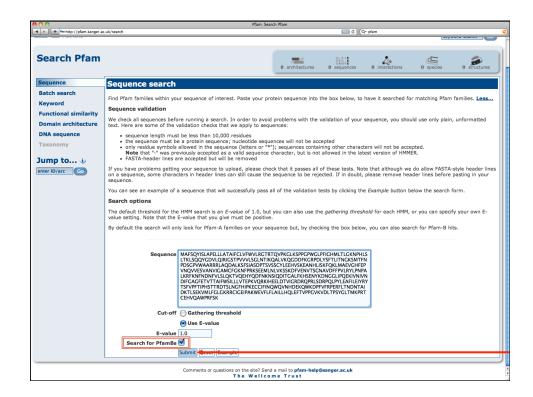
- Pfam A
 - Based on *curated* multiple alignments ("seed alignment")

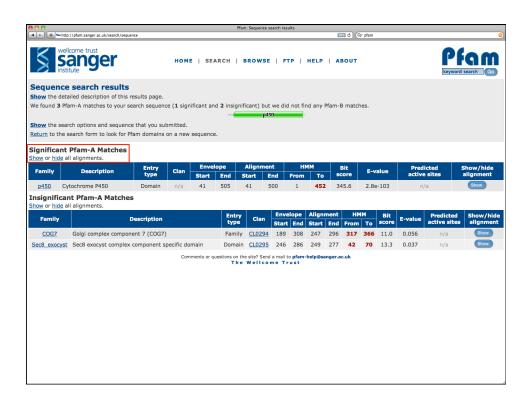
- Hidden Markov models (HMMs) used to find all detectable protein sequences belonging to the family
- Given the method used to construct the alignments, hits are highly likely to be true positives
- Pfam B
 - Automatically generated from database searches
 - Deemed "lower quality", but can be useful when no Pfam A family is identified

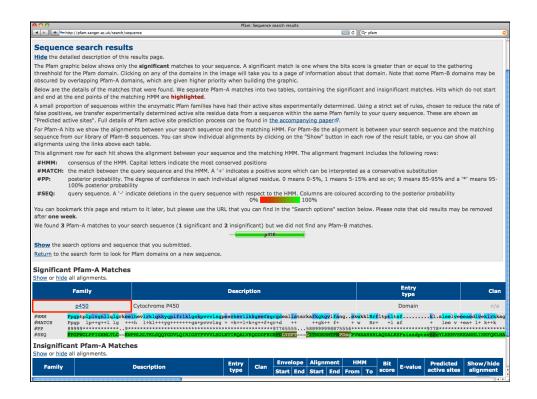




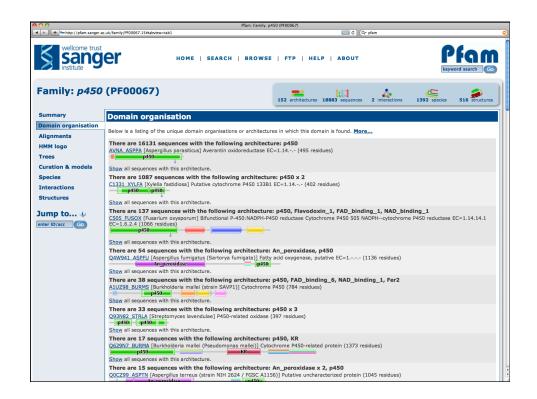


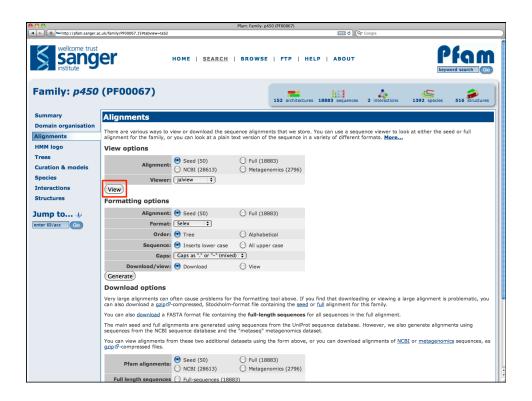


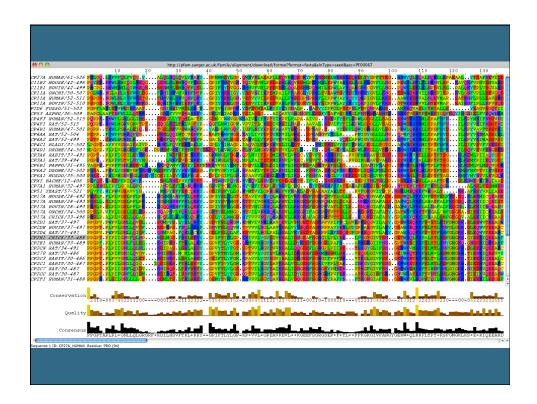


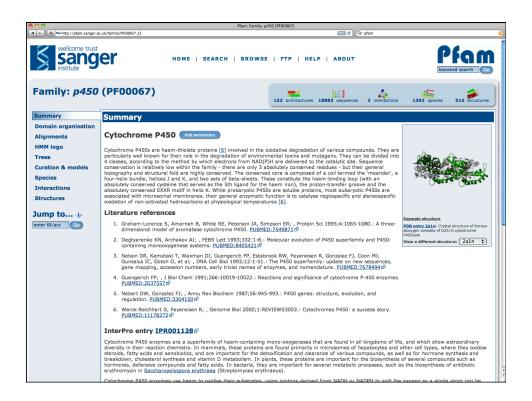


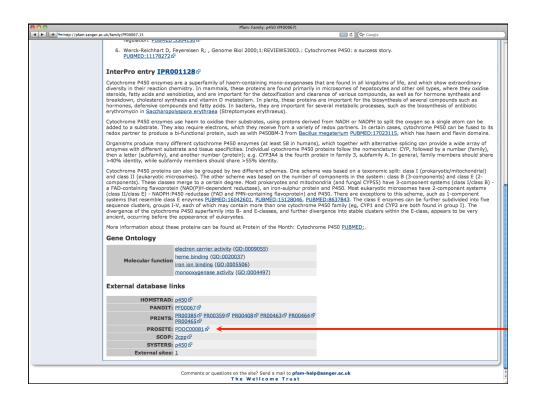




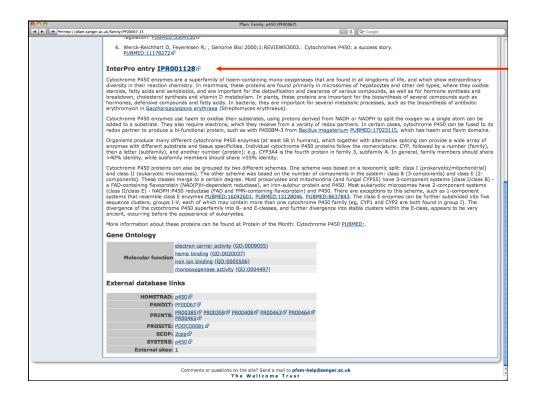


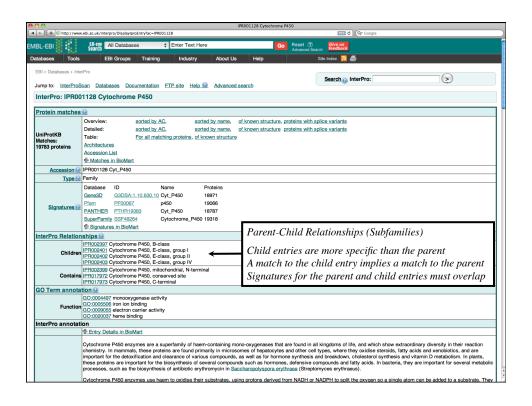


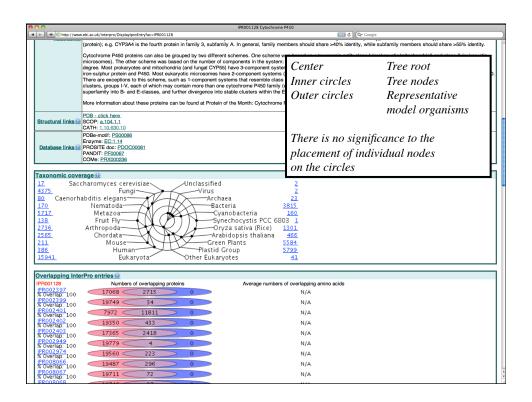


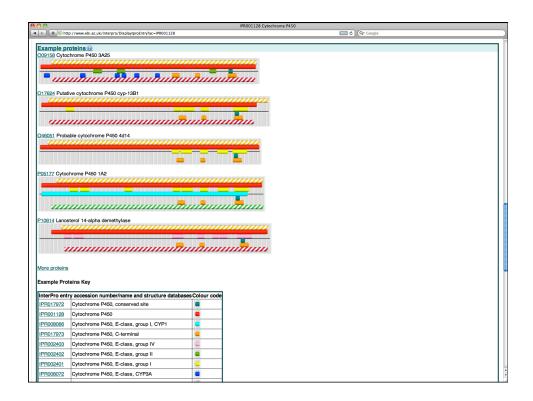


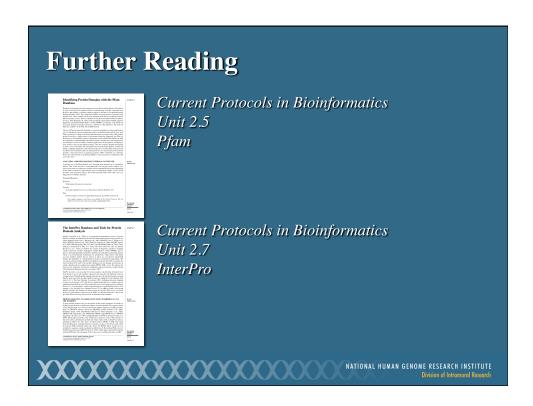












Conserved Domain Database (CDD)

- Identify conserved domains in a protein sequence
- "Secondary database"
 - Pfam A and B
 - Simple Modular Architecture Research Tool (SMART)
 - Clusters of Orthologous Groups

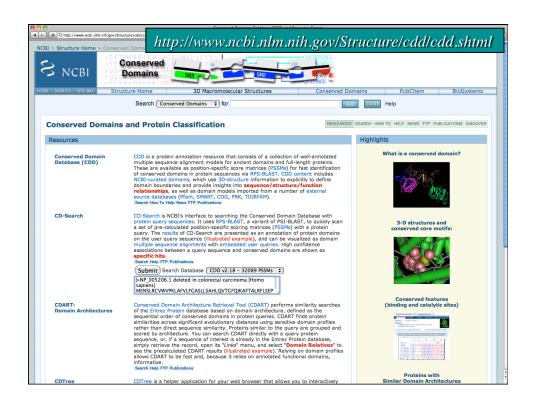
- PRK
- TIGRFAM

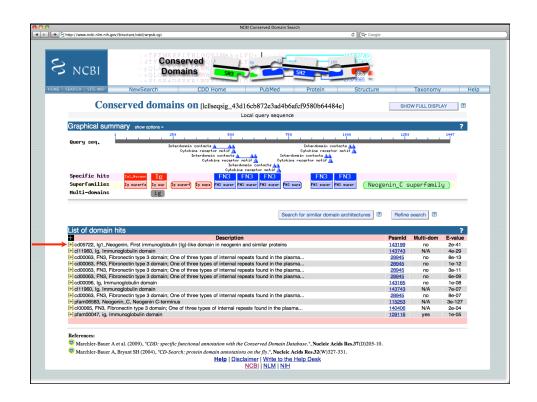
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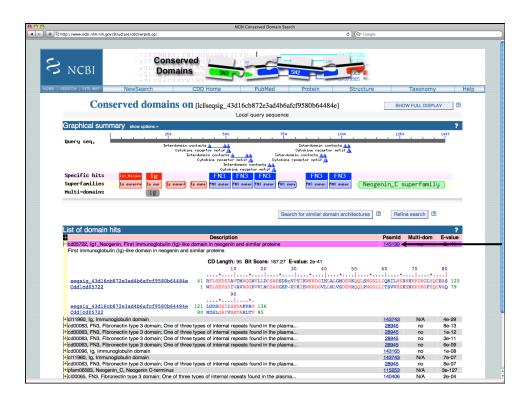
Conserved Domain Database (CDD)

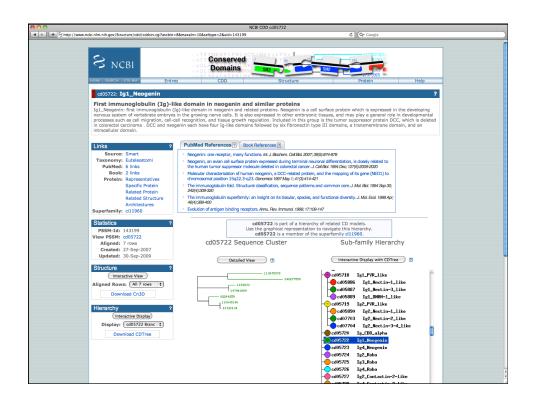
• Search performed using RPS-BLAST

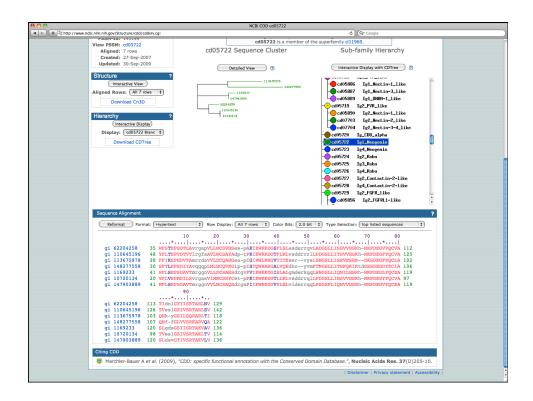
- Query sequence is used to search a database of precalculated position-specific scoring tables
- *Not* the same method used by Pfam or InterPro







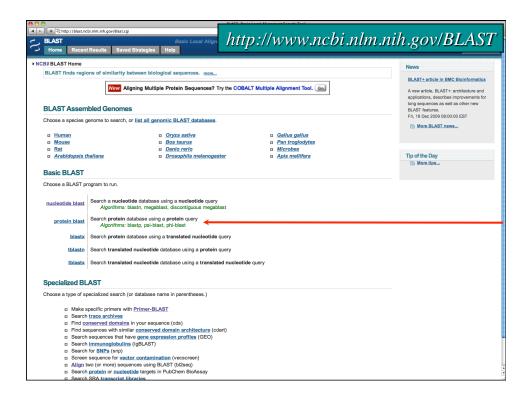


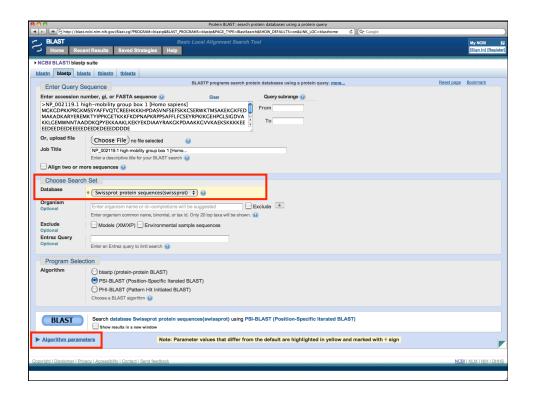


PSI-BLAST

• Position-Specific Iterated BLAST search

- Easy-to-use version of a profile-based search
 - Perform BLAST search against protein database
 - Use results to calculate a position-specific scoring matrix
 - PSSM replaces query for next round of searches
 - May be iterated until no new significant alignments are found
 - Convergence all related sequences deemed found
 - Divergence query is too broad, make cutoffs more stringent

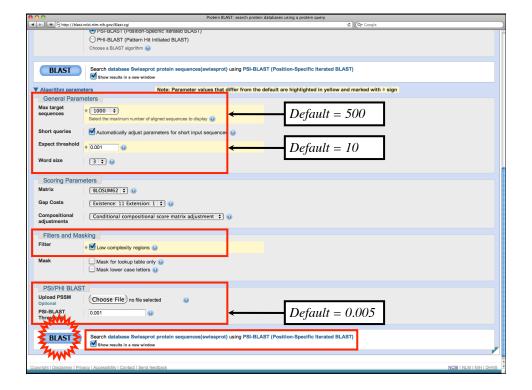


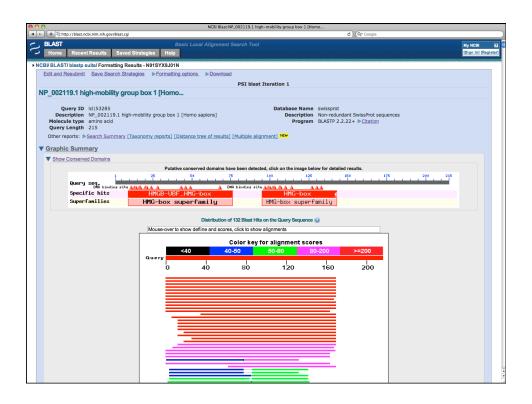


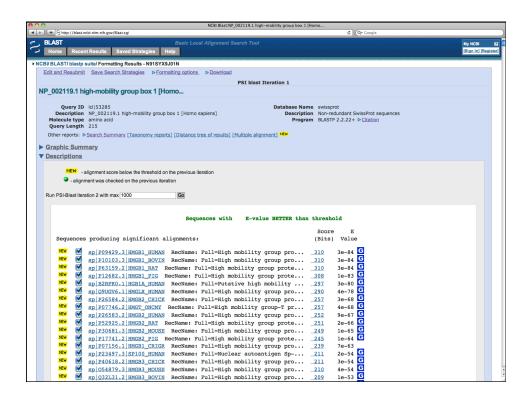
Swiss-Prot

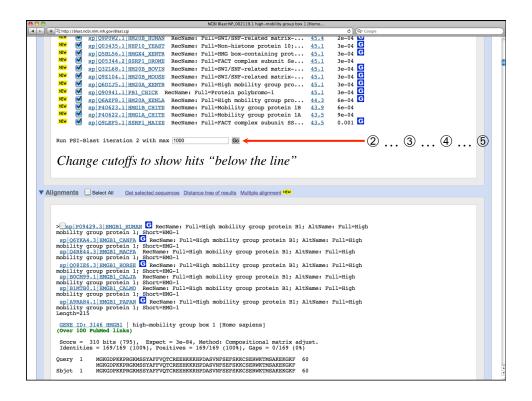
- *Goal:* Provide a single reference sequence for each protein sequence
- Distinguishing Features
 - Non-redundancy
 - Integration with other databases (db_xref)
 - Ongoing curation by EBI staff and external experts
 - Expert annotation includes editing/updates of
 - **CC** Comment lines

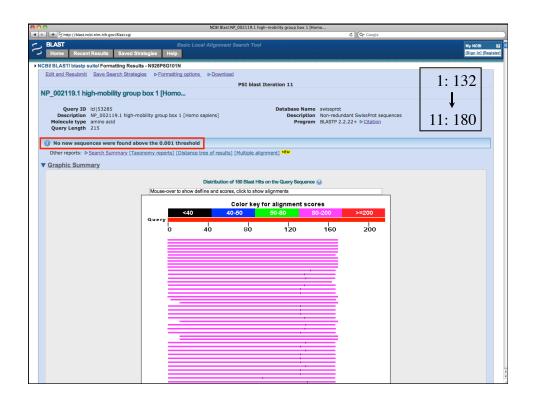
- **FT** Feature table
- Distinct accession series [OPQ] 12345











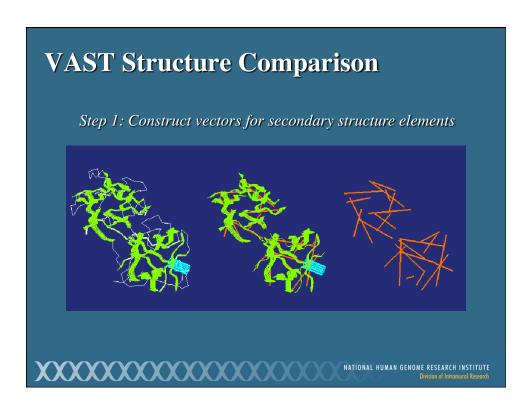
Overview

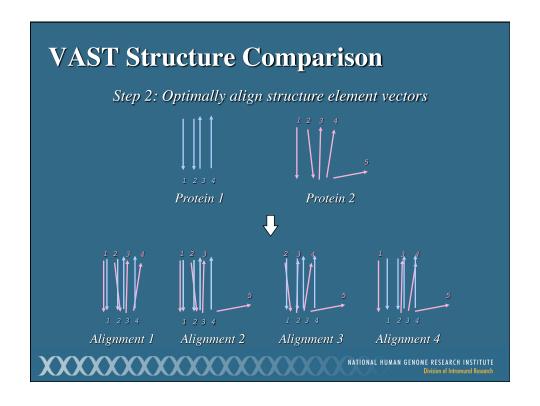
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Predicting Tertiary Structure

- Sequence specifies conformation, *but* conformation does *not* specify sequence
- Structure is conserved to a much greater extent than sequence
- Similarities between proteins may not necessarily be detected through "traditional" methods





VAST Shortcomings

XXXXXXXXXXX

- Not the best method for determining structural similarities
- Reducing a structure to a series of vectors necessarily results in a loss of information (less confidence in prediction)
- Regardless of the "simplicity" of the method, provides a simple and fast first answer to the question of structural similarity

Cn3D Viewer

Rendering: Tubes

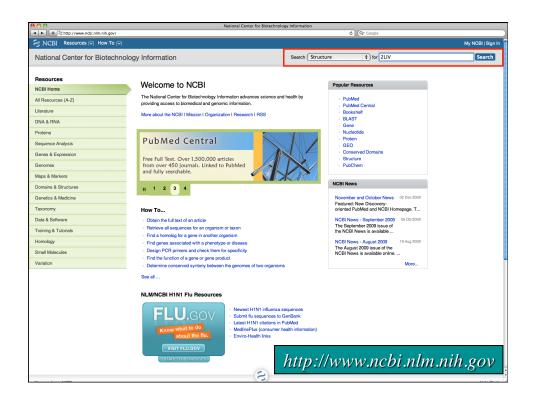
Coloring: Identity

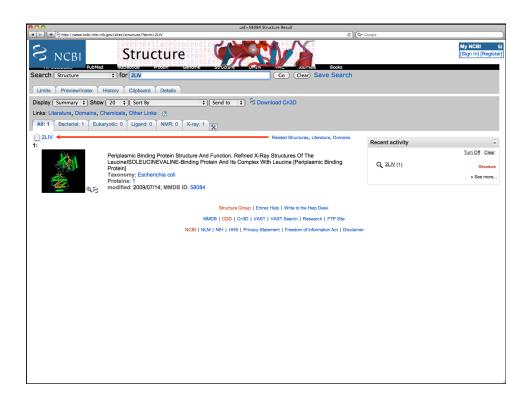
Red – matches

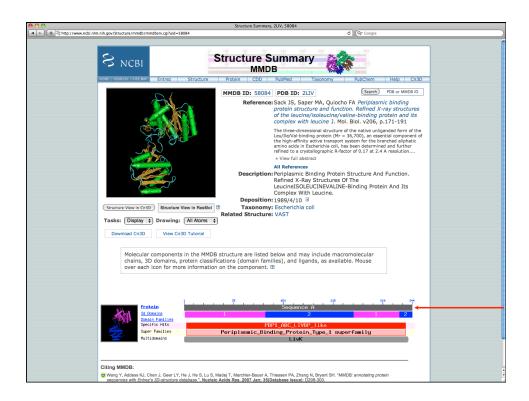
Blue – mismatches

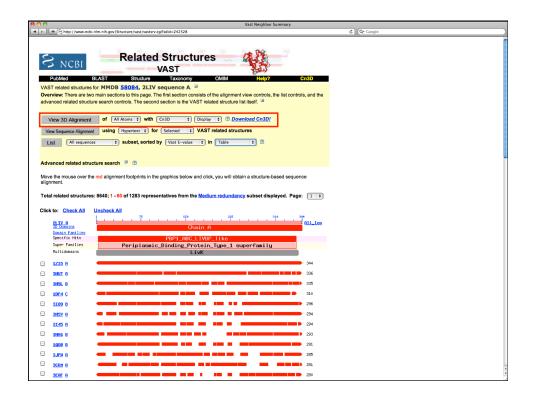
2.IV maghbors - Sequence Aliquement Viewer

2.IV maghbors - Sequence

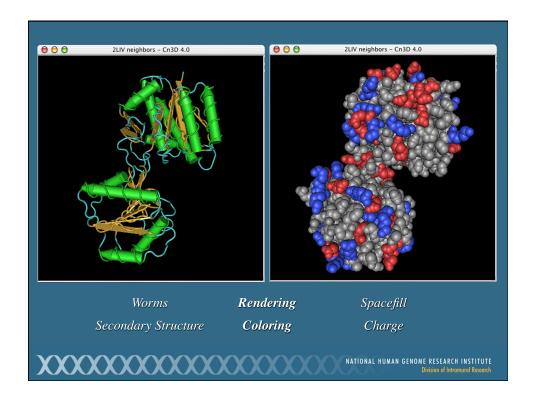


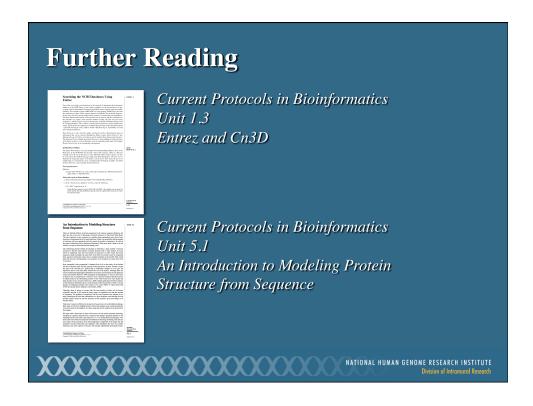










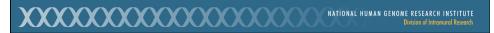


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Why do multiple sequence alignments?

- Identify conserved regions, patterns, and domains
 - Experimental design
 - Predicting structure and function
 - Identifying new members of protein families
- Perform phylogenetic analysis
- Generate position-specific scoring matrices for subsequent searches ("many-against-one" or "one against many")
- Bolster confidence in secondary structure predictions



Considerations

- Absolute sequence similarity

 Create the alignment by lining up as many common characters as possible
- Conservation

 Take into account residues that can substitute for one another and not adversely affect the function of the protein
- Structural similarity

 Knowledge of the secondary or tertiary structure of the proteins being aligned can be used to fine-tune the alignment

General Guidelines

- As with most analyses, concentrate on the protein level rather than on the nucleotide level
 - More informative
 - Less prone to inaccurate alignment ("20 vs. 4")
 - Can "translate back" to nucleotide sequences after doing the alignment

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General Guidelines

- Use a reasonable number of sequences to avoid technical difficulties
 - *Global* alignment method: compute time increases exponentially as sequences are added to the set
 - Most alignment algorithms are ineffective on huge data sets (and may yield inaccurate alignments)
 - Phylogenetic studies resulting from inordinately large data sets are almost impossible
 - Good starting point: 10-15 sequences
 - Ballpark upper limit: 50 sequences

General Guidelines

- Selecting sequences for alignment
 - Sequences should be of about the same length
 - Use closely-related sequences to determine "required" amino acids
 - Use more divergent sequences to study evolutionary relationships
 - Good starting point: use sequences that are 30-70% similar to most of the other sequences in the data set
 - The most informative alignments result when the sequences in the data set are not "too similar", but also not "too different"

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General Guidelines

- Iterative process
 - Perform alignment on small set of sequences
 - Examine the quality of the alignment
 - If alignment good, can add new sequences to data set, then realign
 - If alignment not good, remove any sequences that result in the inclusion of long gaps, then realign

Interpretation

- Absolutely-conserved positions are *required* for proper structure and function
- Relatively well-conserved positions are able to tolerate limited amounts of change and not adversely affect the structure or function of the protein
- Non-conserved positions may "mutate freely," and these mutations can possibly give rise to proteins with new functions

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Interpretation

- Gap-free blocks probably correspond to regions of secondary structure
- Gap-rich blocks probably correspond to unstructured or loop regions

ClustalW2

- Automatic multiple alignment of nucleotide or amino acid sequences
- Implementations
 - Client versions command-line text menu system, all platforms
 - Web-based version http://www.ebi.ac.uk/clustalw2

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Progressive Alignment

- Align two sequences at a time
- Gradually build up the multiple sequence alignment by merging larger and larger sub-alignments, clustering on the basis of similarity
- Uses protein scoring matrices and gap penalties to calculate alignments having the best score
- Major advantages of method
 - Very fast

XXXXXXXXXXXXXXX

• Alignments generally of high quality

Progressive Alignment

>sequence A

VHLTPEEKSAVTALWGKVNVDEVGGEALGRLLVVYPWTQRFFESFGDLST >sequence B

VQLSGEEKAAVLALWDKVNEEEVGGEALGRLLVVYPWTQRFFDSFGDSLN >sequence C

 ${\tt VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSH}$ >sequence D

VLSAADKTNVKAAWSKVGGHAGEYGAEALERMFLGFPTTKTYFPHFDLSH

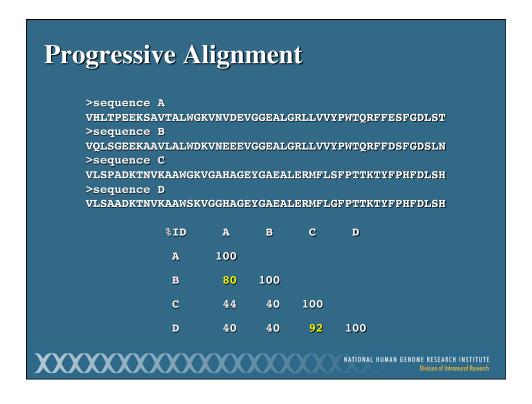
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Progressive Alignment

1. Calculate a similarity score (percent identity) between every pair of sequences to drive the alignment

For N sequences, this requires the calculation of $[N \times (N-1)]/2$ pairwise alignments

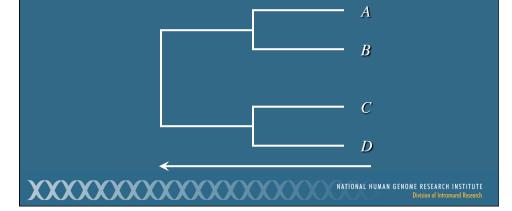
Alignments
6
45
300
1,225
4,950



Progressive Alignment
 Align A with B → alignment AB (fixed) Align C with D → alignment CD (fixed) Represent alignments AB and CD as single sequences
$\begin{array}{cccccccccccccccccccccccccccccccccccc$
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Progressive Alignment

- Align "sequence" AB with "sequence" CD
- Continue following the branching order of the tree, from the tips to the root, merging each new pair of "sequences"



Progressive Alignment: Advantages

- Do "easier" alignments between highly-related sequences first
- Use information regarding conservation at each position to help with more difficult alignments between more distantly-related sequences later on in process

Progressive Alignment: Disadvantages

- If initial alignments are made on distantly related sequences, there may be errors in the initial alignments
- Once an alignment is "fixed", it is not reconsidered, so any errors in the early alignments may propagate through subsequent alignments
- New version of ClustalW2 does provide a "remove first" iteration scheme to attempt to improve alignments

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ClustalW2 Output

- Pairwise scores
- Multiple sequence alignment (.aln)
 - Alternative formats available:

GCG

Phylip

PIR

GDE

ClustalW2 Output

- Cladogram
 - Tree assumed to be an estimate of a phylogeny
 - Branches are of equal length

- Cladograms show common ancestry, but do not provide an indication of the amount of "evolutionary time" separating taxa
- Phylogram
 - Tree that is assumed to be an estimate of phylogeny
 - Branch lengths proportional to the amount of inferred evolutionary change

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ClustalW2 Conservation Patterns

• Conservation patterns in multiple sequence alignments usually follow the following rules:

[WYF] Aromatics
[KRH] Basic side chains (+)

[DE] Acidic side chains (–)

[GP] Ends of helices
[HS] Catalytic sites

[C] Cysteine cross-bridges

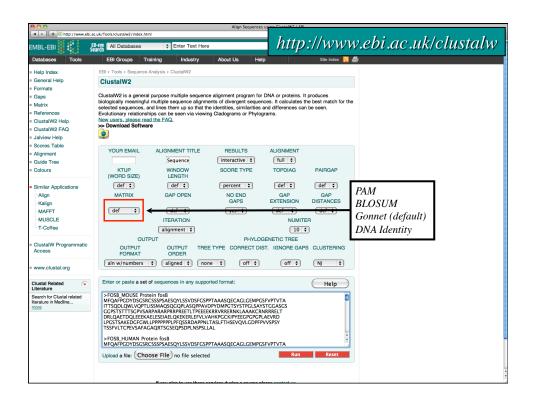
ClustalW2 Conservation Patterns

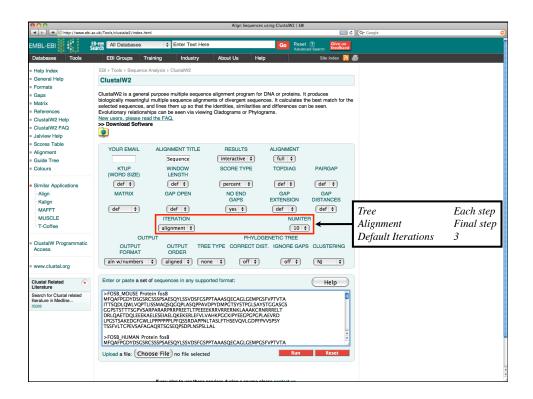
- Interpretation is *empirical* there is no parallel to the *E*-values seen in BLAST searches to assess "significance"
 - * entirely conserved column (want in at least 10% of positions)
 - conserved"

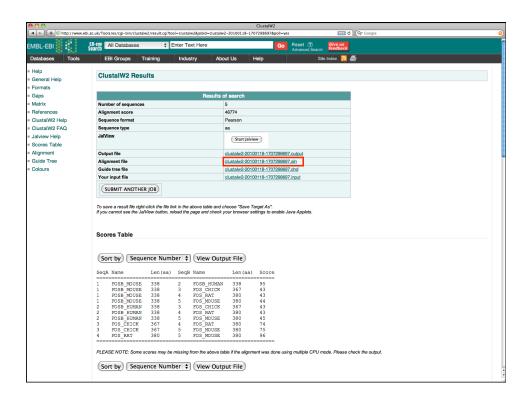
 (according to color table)
 - "semi-conserved"

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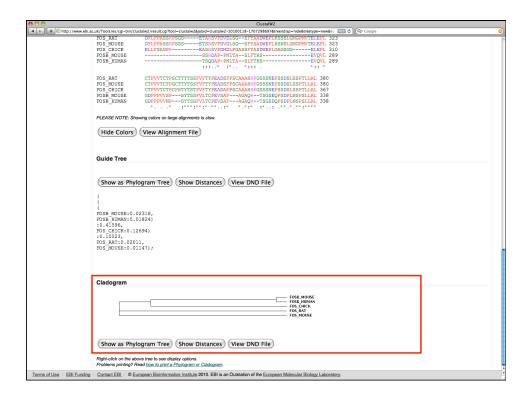
ClustalW Colors AVFPMILW RED Small (small+ hydrophobic (incl.aromatic -Y)) DE BLUE Acidic RK MAGENTA Basic - H STYHCNGQ GREEN Hydroxyl + sulfhydryl + amine + G Others Grey Unusual amino/imino acids etc

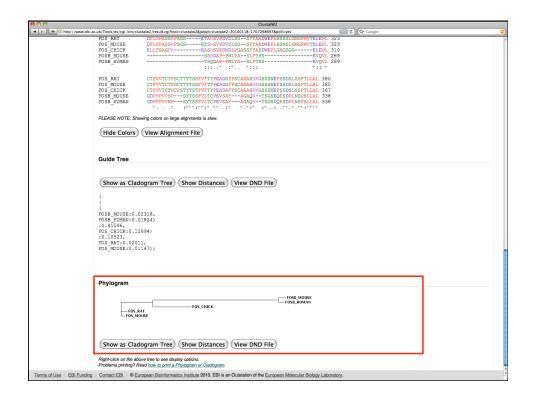












Jalview

- Java applet available within ClustalW2 results
- Used to manually edit ClustalW2 alignments
- Color residues based on various properties
- Pairwise alignment of selected sequences
- Consensus sequence calculations
- Removal of redundant sequences
- Calculation of phylogenetic trees

• Color PostScript output



