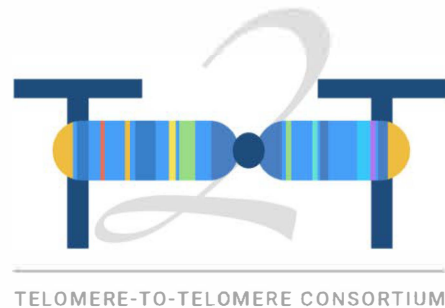


The Human Pangenome Project: Creating a Reference that Better Represents Human Global Genetic Diversity



TOWARDS A
COMPLETE
REFERENCE OF
HUMAN GENOME
DIVERSITY

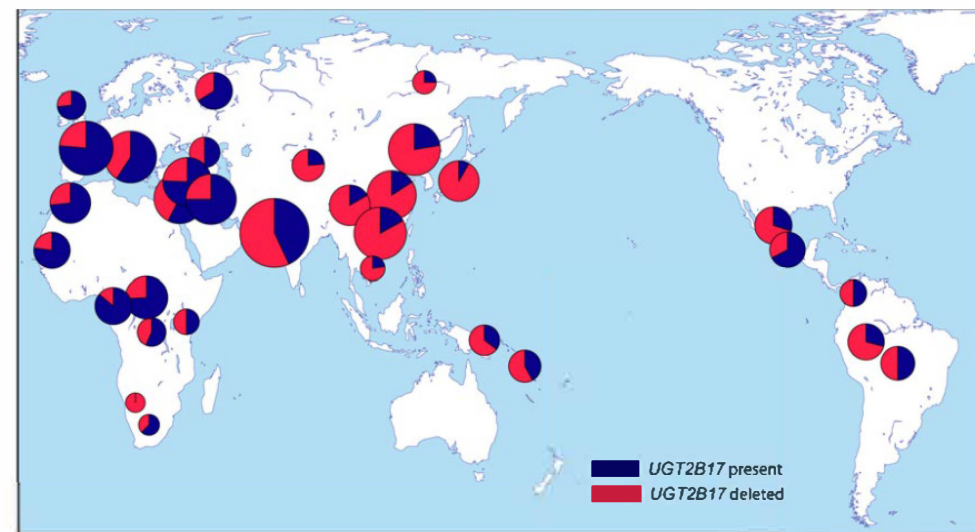


Deanna Church
Presenting

The current human reference fails to faithfully represent a single individual genome much less the genomes of a global population.

RP11 hap1
UGT2B17

RP11 hap2



- **The human reference genome** is a foundational resource in human genetics and like most technology-driven resources, is overdue for an upgrade.
- The current structure is a **linear monoploid representation containing mixed haplotypes with too many gaps and errors. Additionally, the underlying sequence is predominately from a single individual.**
- **Mapping limitations of short reads and inherent reference biases** means we have missed more than 70% of structural variants in traditional whole-genome sequencing studies

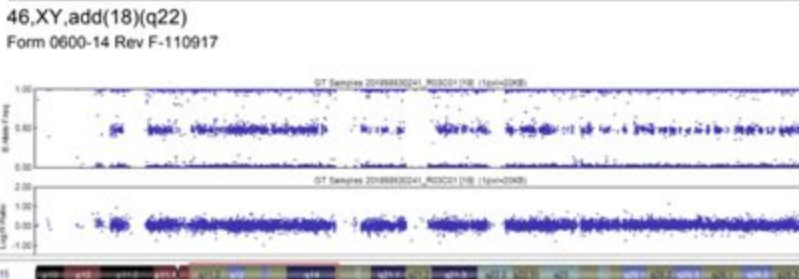
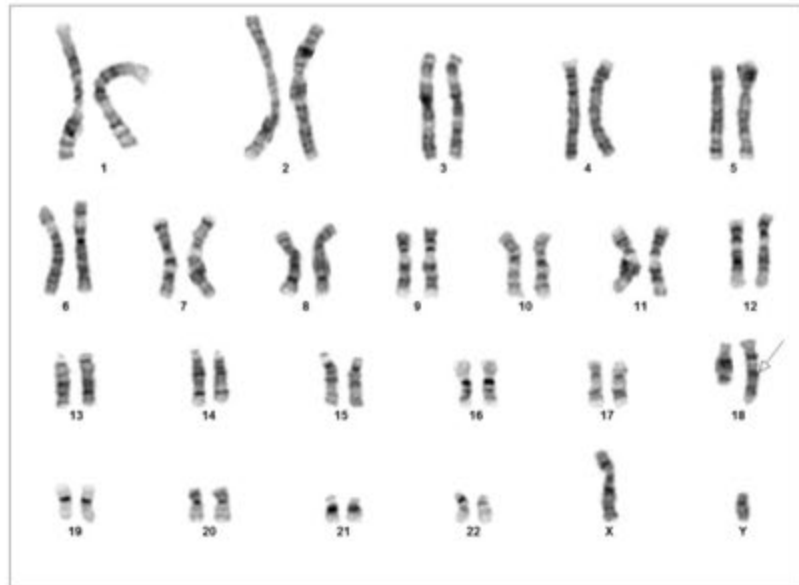


Human Pangenome Reference Consortium

- Improve representation of **global genomic diversity** (>350 diverse diploid references)
- **Prioritizing quality**: we aim to release a complete (T2T) and comprehensive map of genome variation
- **Develop a new, non-linear reference data structure** and foster an innovative ecosystem of pangenomic tools
- Outreach, Education and Implementation

Multi-Center Sequencing Technology and Production: Optimized for Efficiency and Quality

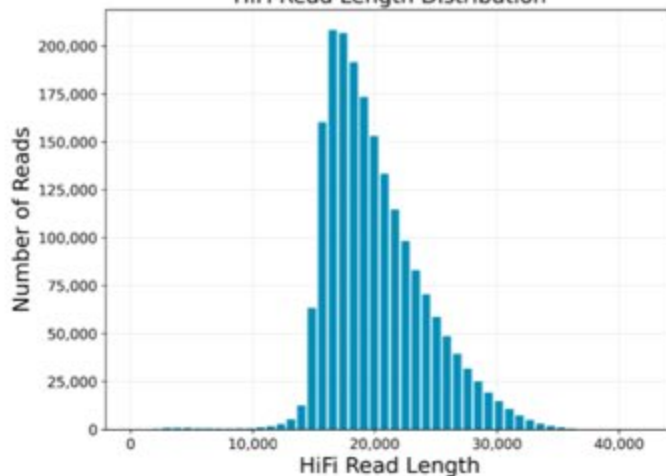
Cell line stability/QC



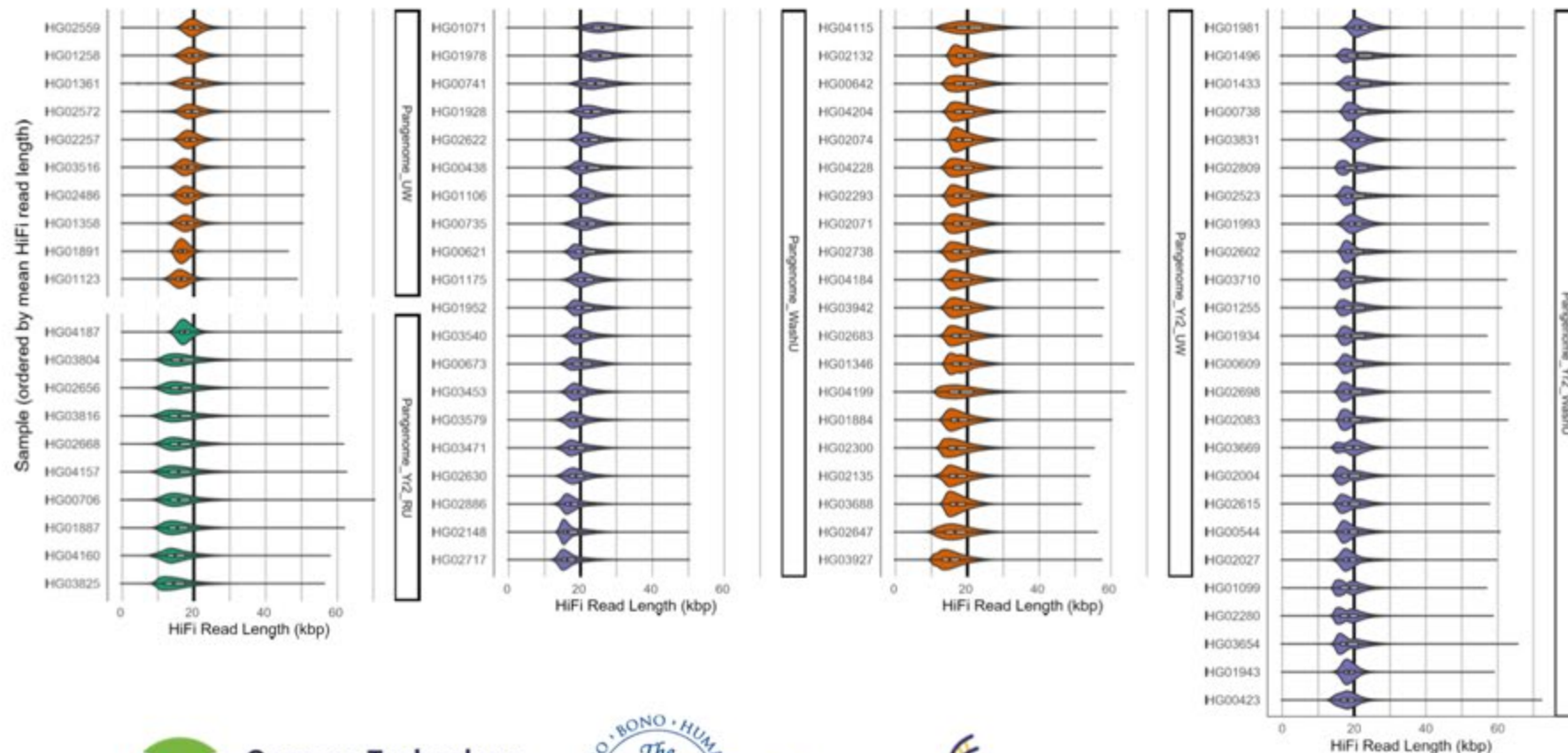
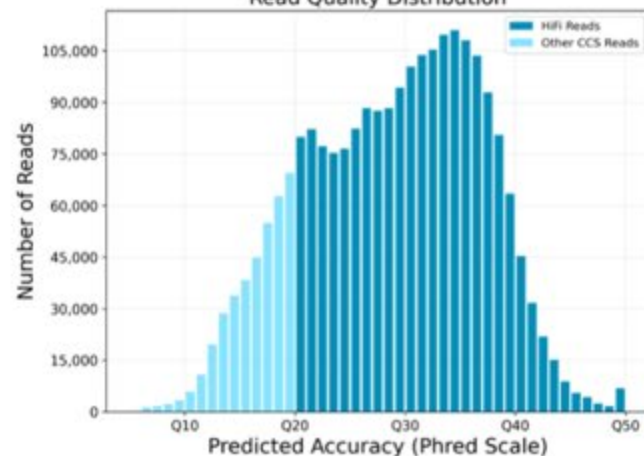
Optimized and Consistent Long-Read HiFi Production



HiFi Read Length Distribution



Read Quality Distribution



Forefront of ultra-long read sequencing innovation

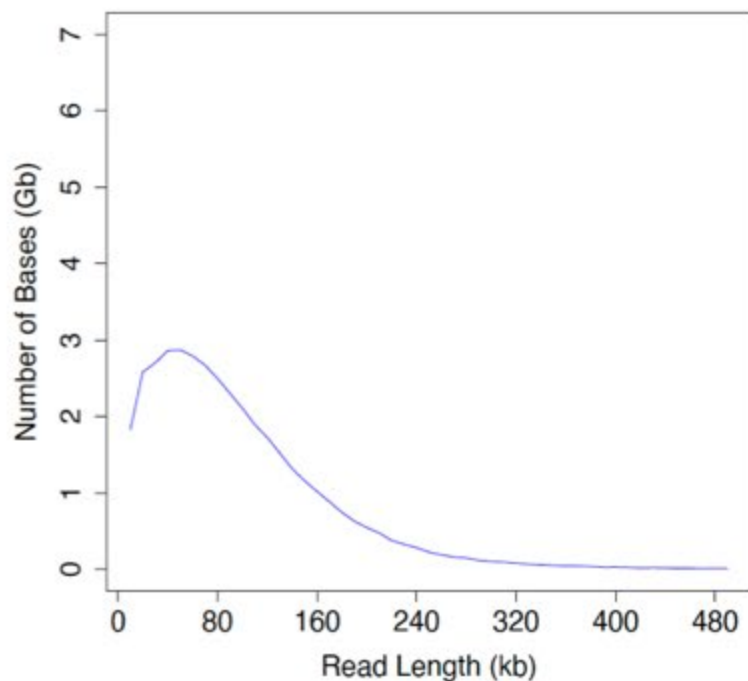
UC SANTA CRUZ Genomics Institute

Oxford NANOPORE Technologies

circulomics

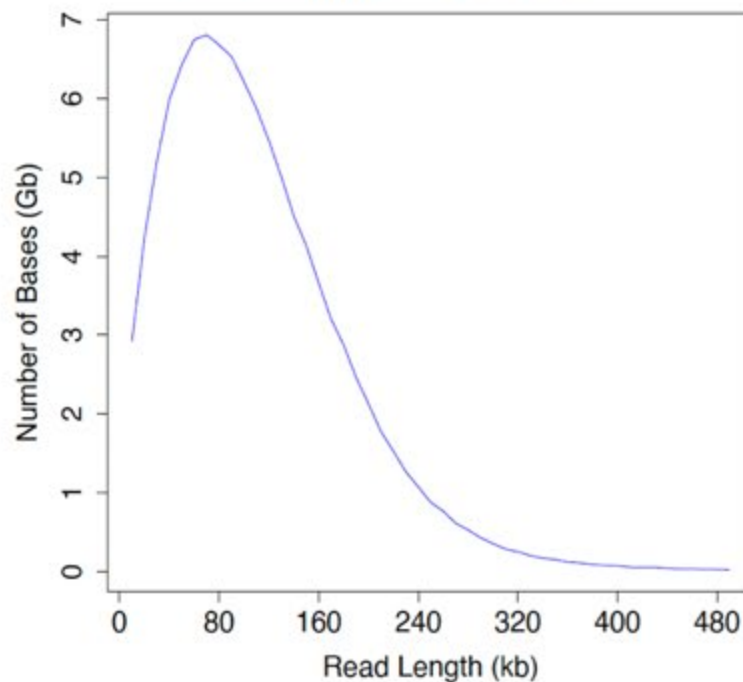
2020

Read N50 ~76 kb
~9X coverage per flow cell
~3.5X coverage in 100 kb+



2021

Read N50 ~73 kb
~30X coverage per flow cell
~9X coverage in 100 kb+



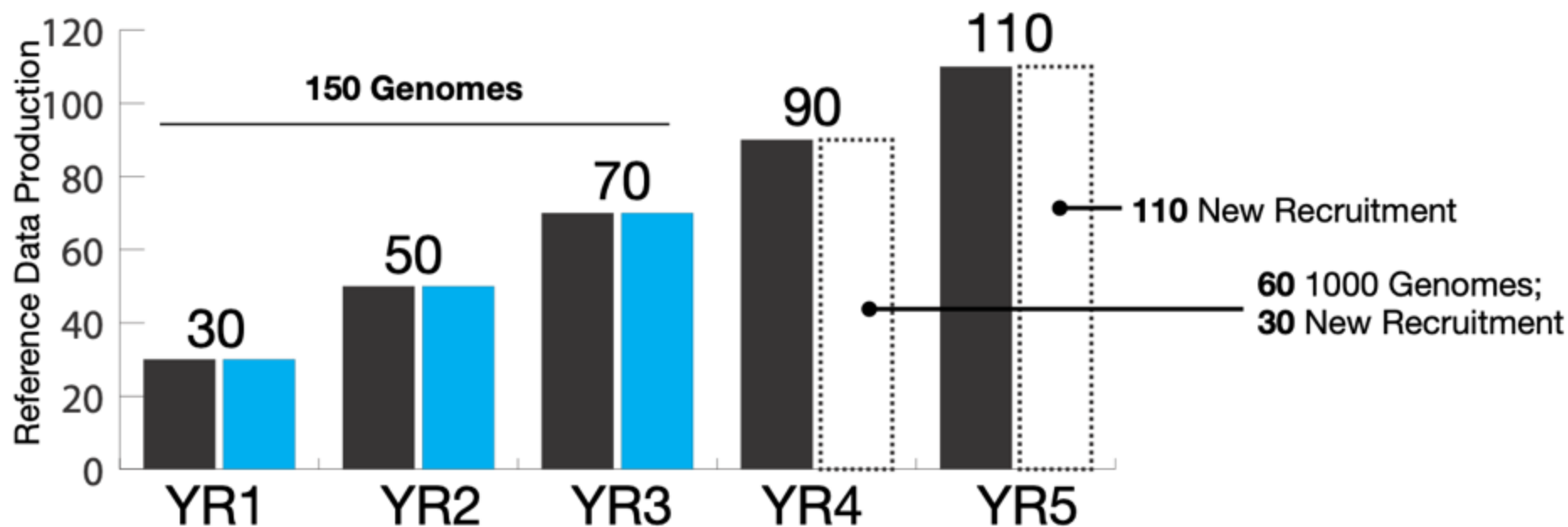
3x increase in throughput



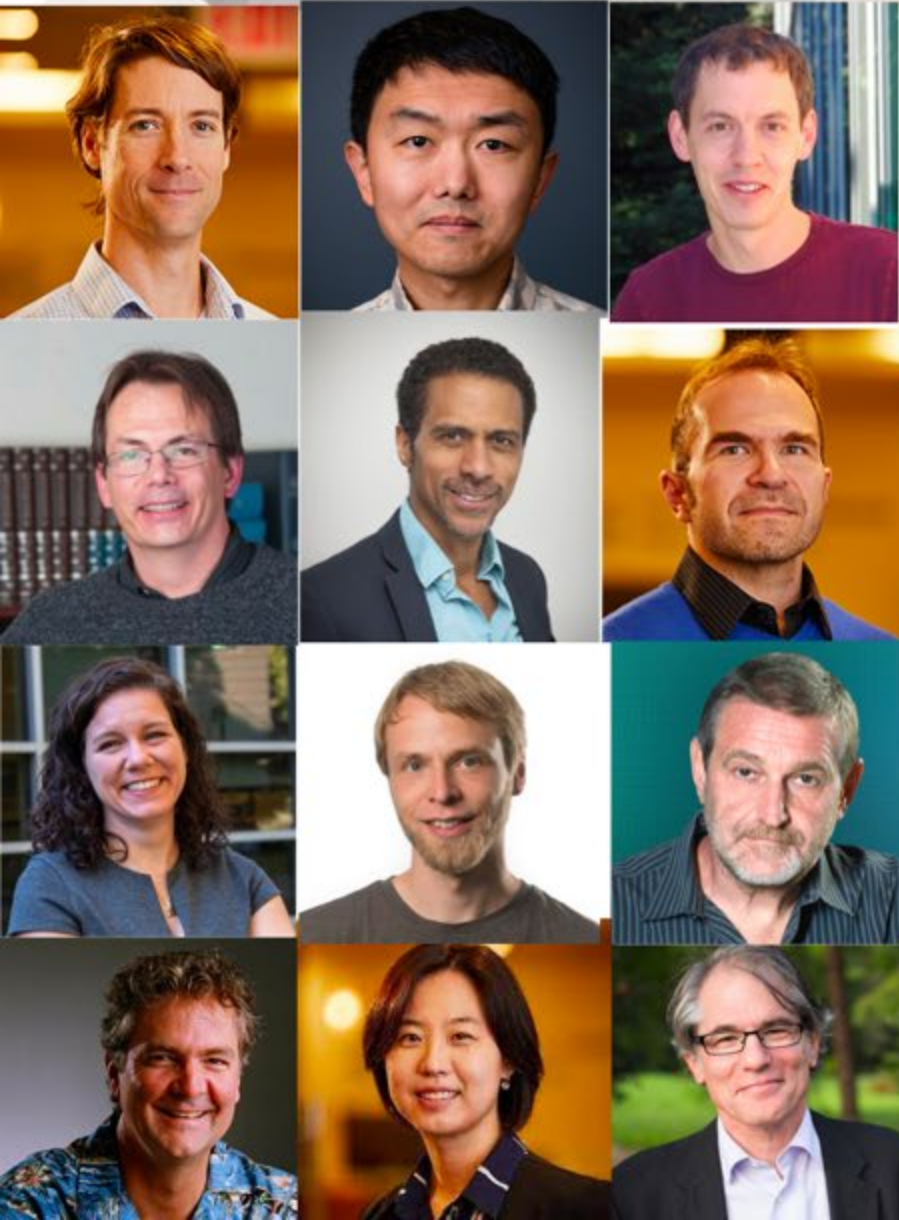
Overview

Population Sampling and Representation (Phase 1): We are representing >99.9% common SNVs (1%) in the 1000 Genomes lines. **(Phase 2)** We are in position for perspective recruitment of remaining 150 individuals (BioMe)

Sequence Technology and Production: Highly efficient multi-center production effort, automated assembly and quality assessment



Innovation in Long Read Assembly Methods



We have assembled the leaders in long-read assembly methods, with an emphasis in researchers involved in finishing and repeat assemblies.



Earth's heart of iron begins
to yield its secrets p. 18

Microglia in chronic pain recovery
and relapse pp. 33 & 86

Particle acceleration
in a nova explosion p. 77

Science

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1 APRIL 2022
SPECIAL ISSUE
science.org

AAAS

FILLING THE GAPS

Closing in on a complete
human genome p. 42

SPECIAL ISSUE

Completing the human genome

The complete sequence of a human genome



BY SERGEY NURK, SERGEY KOREN, ARANG RHIE, MIKKO RAUTIAINEN, ANDREY V. BZIKADZE, ALLA MIKHEENKO, MITCHELL R. VOLLGER, NICOLAS ALTEMOSE, LEV URALSKY, ARIEL GERSHMAN, [...] ADAM M. PHILLIPPY **+89 authors** • 31 MAR 2022 : 44-53

Epigenetic patterns in a complete human genome



BY ARIEL GERSHMAN, MICHAEL E. G. SAURIA, XAVI GUITART, MITCHELL R. VOLLGER, PAUL W. HOOK, SAVANNAH J. HOYT, MITEN JAIN, ALAINA SHUMATE, ROHAM RAZAGHI, SERGEY KOREN, [...] WINSTON TIMP **+9 authors** • 01 APR 2022

Segmental duplications and their variation in a complete human genome



BY MITCHELL R. VOLLGER, XAVI GUITART, PHILIP C. DISHUCK, LUDOVICA MERCURI, WILLIAM T. HARVEY, ARIEL GERSHMAN, MARK DIEKHANS, ARVIS SULOVARI, KATHERINE M. MUNSON, ALEXANDRA P. LEWIS, [...] EVAN E. EICHLER **+9 authors** • 01 APR 2022

From telomere to telomere: The transcriptional and epigenetic state of human repeat elements



BY SAVANNAH J. HOYT, JESSICA M. STORER, GABRIELLE A. HARTLEY, PATRICK G. S. GRADY, ARIEL GERSHMAN, LEONARDO G. DE LIMA, CHARLES LIMOUSE, REZA HALABIAN, LUKE WOJENSKI, MATIAS RODRIGUEZ, [...] RACHEL J. O'NEILL **+16 authors** • 01 APR 2022

A complete reference genome improves analysis of human genetic variation



BY SERGEY AGANEZOV, STEPHANIE M. YAN, DANIELA C. SOTO, MELANIE KIRSCH, SAMANTHA ZARATE, PAVEL AVDEYEV, DYLAN J. TAYLOR, KISHWAR SHAFIN, ALAINA SHUMATE, CHUNLIN XIAO, [...] MICHAEL C. SCHATZ **+22 authors** • 01 APR 2022

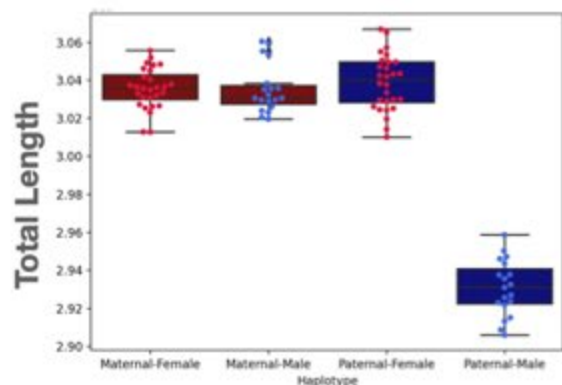
Complete genomic and epigenetic maps of human centromeres



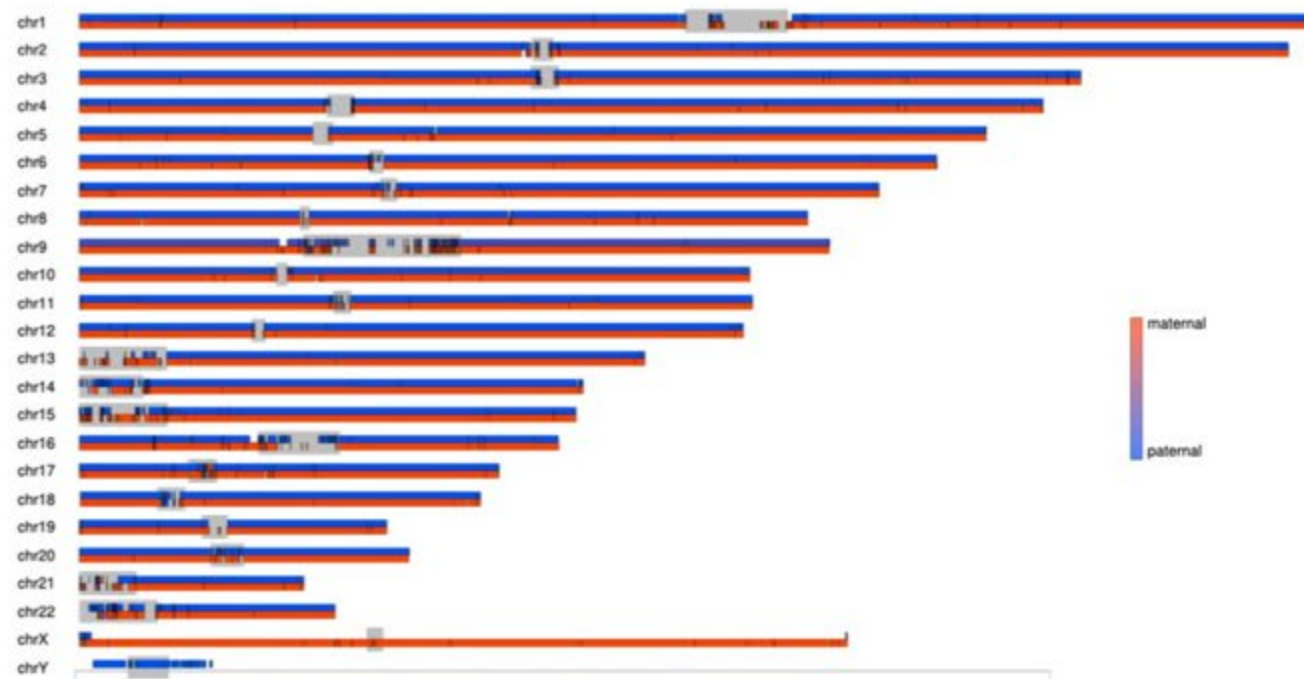
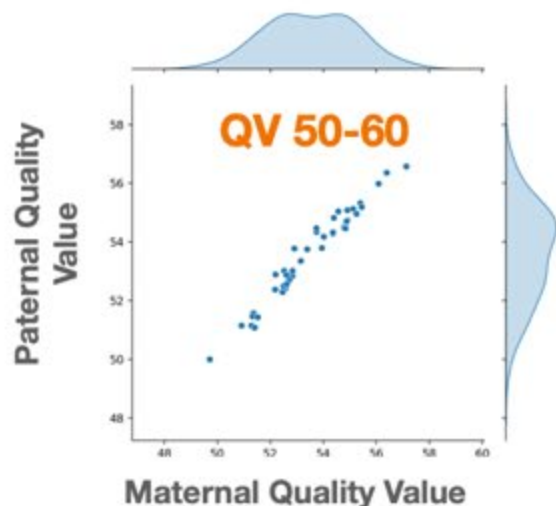
BY NICOLAS ALTEMOSE, GLENNIS A. LOGSDON, ANDREY V. BZIKADZE, PRAGYA SIDHWANI, SASHA A. LANGLEY, GINA V. CALDAS, SAVANNAH J. HOYT, LEV URALSKY, FEDOR D. RYABOV, COLIN J. SHEW, [...] KAREN H. MIGA **+48 authors** • 01 APR 2022

Automated assembly standards: High-quality References

We tested the current best practices in sequencing technologies and automated assembly algorithms on one human sample, HG002, an openly- consented Ashkenazi individual from the Personal Genome Project

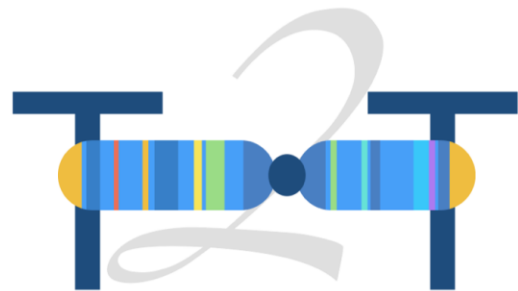
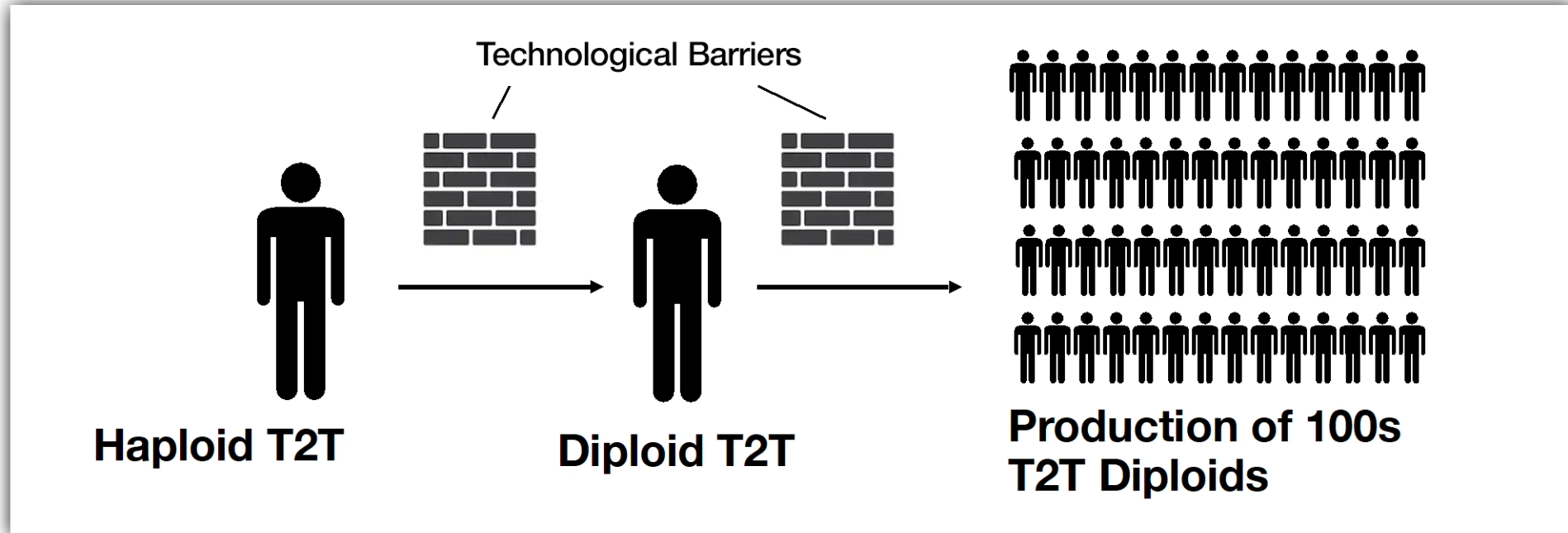


T2T-CHM13
3.055 Gb



Near complete chromosome scaffolds between HPRC-HG002 **maternal** and **paternal** assemblies.

One genome is not enough....



TELOMERE-TO-TELOMERE CONSORTIUM

TOWARDS A
COMPLETE
REFERENCE OF
HUMAN GENOME
DIVERSITY



Population Sampling and Representation

Goals:

- Establish a framework to define diversity and prioritize cell lines from 350 globally diverse participants to build a new pangenome reference
- Aim to capture most common variants, defined as variants at $>1\%$ frequency, in human populations globally

Challenges:

- Incomplete understanding of the full spectrum of human genetic diversity
- Not one way to define diversity in this study: multiple well motivated approaches proposed for sample selection

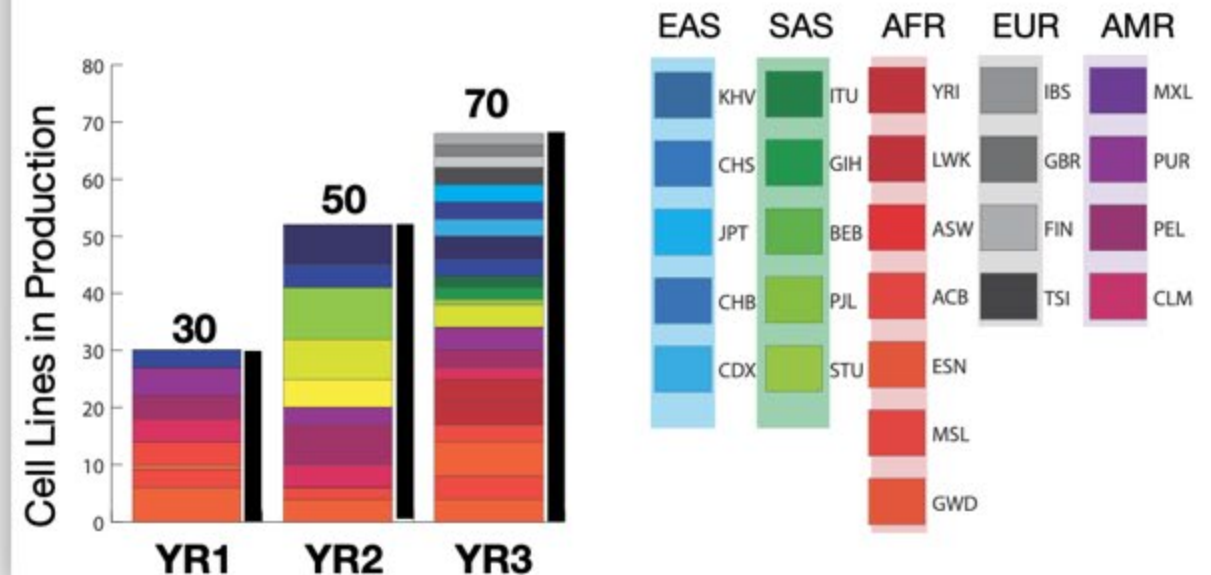
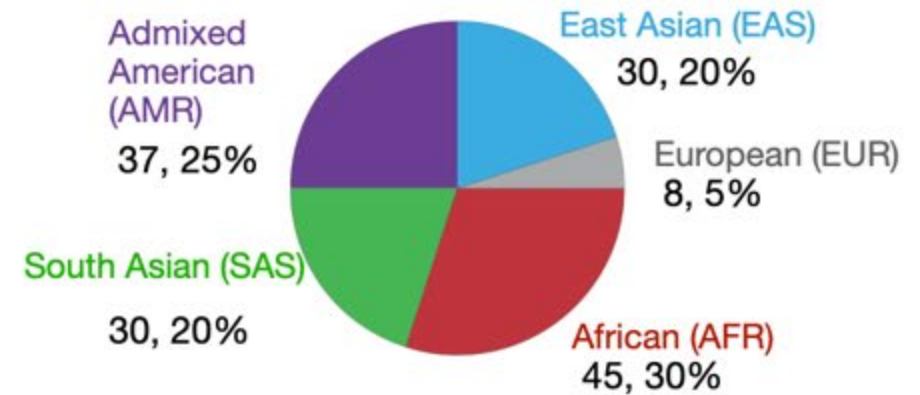
Phase 1 HPRC Sample Selection



- ✓ Cover genetic and geographic diversity
- ✓ Availability of low passage cell lines
- ✓ Availability of trios/parental data (YR1-2)



Phase 1 Production: 150 total lines



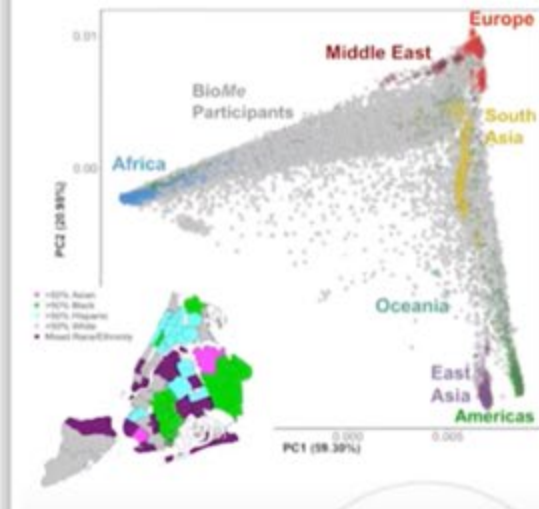


Mount Sinai
The Charles Bronfman
Institute for Personalized Medicine

**BioMe
Biobank**
Your history is our future



Eimear Kenny, PI



Mount Sinai's BioMe BioBank:

>70,000 participants from 160 countries, with sequencing data for more than 40,000 participants.



Washington University, St. Louis Recruitment Center:

New recruitment from African American communities

Phase 2: Prioritization of New Participants

- **Model 1:** Maximizing common variant diversity

Leveraging sequence data, iteratively select samples that maximize common variant coverage in an out-of-sample dataset

- **Model 2:** Maximizing genetic divergence

Using sequence/array data, plot PCA and select participants to represent the continuum of genetic dissimilarity in principle component (PC) space

- **Model 3:** Targeting underrepresented populations based on self-reporting/geographical data

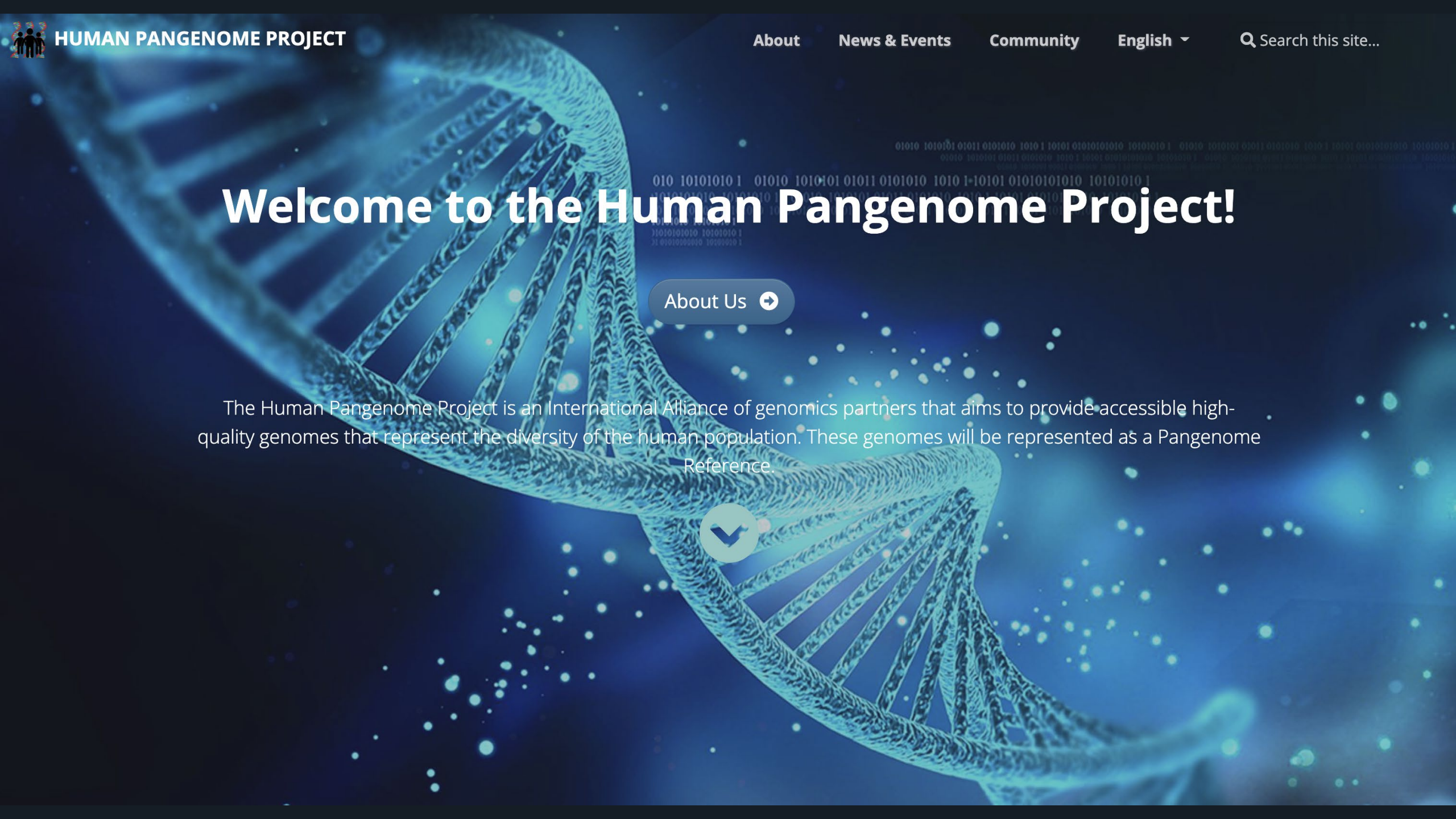
Using country-of-origin data, designate countries/subcontinental regions of interest to the project, and select participants from those regions

Partnership with ELSI Team for Consent and Outreach



Embedded
Ethics Team

- **Formal Review of Consent Language (compatibility with 1000 genomes consent)**
- **Design and review of outreach materials for prospective recruitment**
- **Alignment with internal ethics/genomic review board at BioMe**
- **Review of the use of external IRB ( BRANY)**



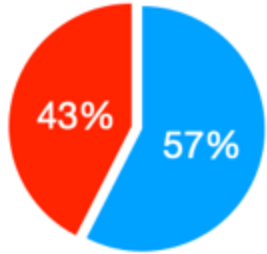
Welcome to the Human Pangenome Project!

[About Us](#) ➡

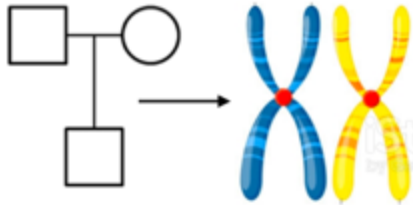
The Human Pangenome Project is an International Alliance of genomics partners that aims to provide accessible high-quality genomes that represent the diversity of the human population. These genomes will be represented as a Pangenome Reference.



Improving Population Sampling and Representation



Continue reference production of 1000 genomes (200/350)



Move away from dependency on trios (priority based on genomic diversity) Establish new assembly methods that do not rely on parental data

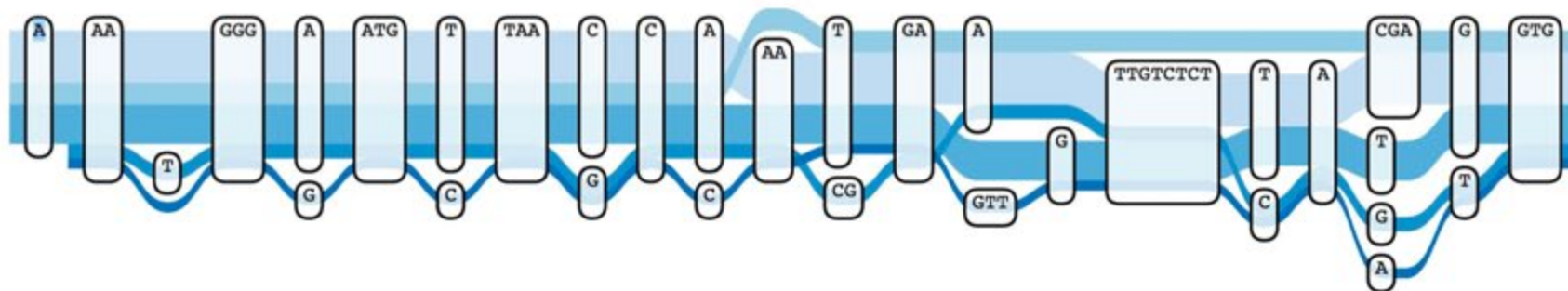
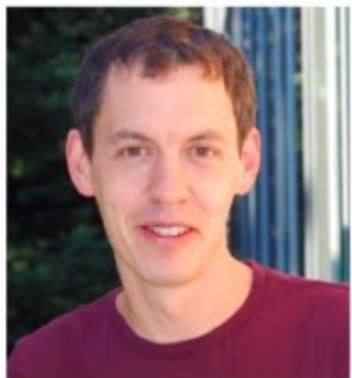


Launch new recruitment efforts and establish 100 new LCLs (NHGRI Collection: Human Pangenome) outside of 1000 Genome Cell lines



Establish new collaborations and international partnerships

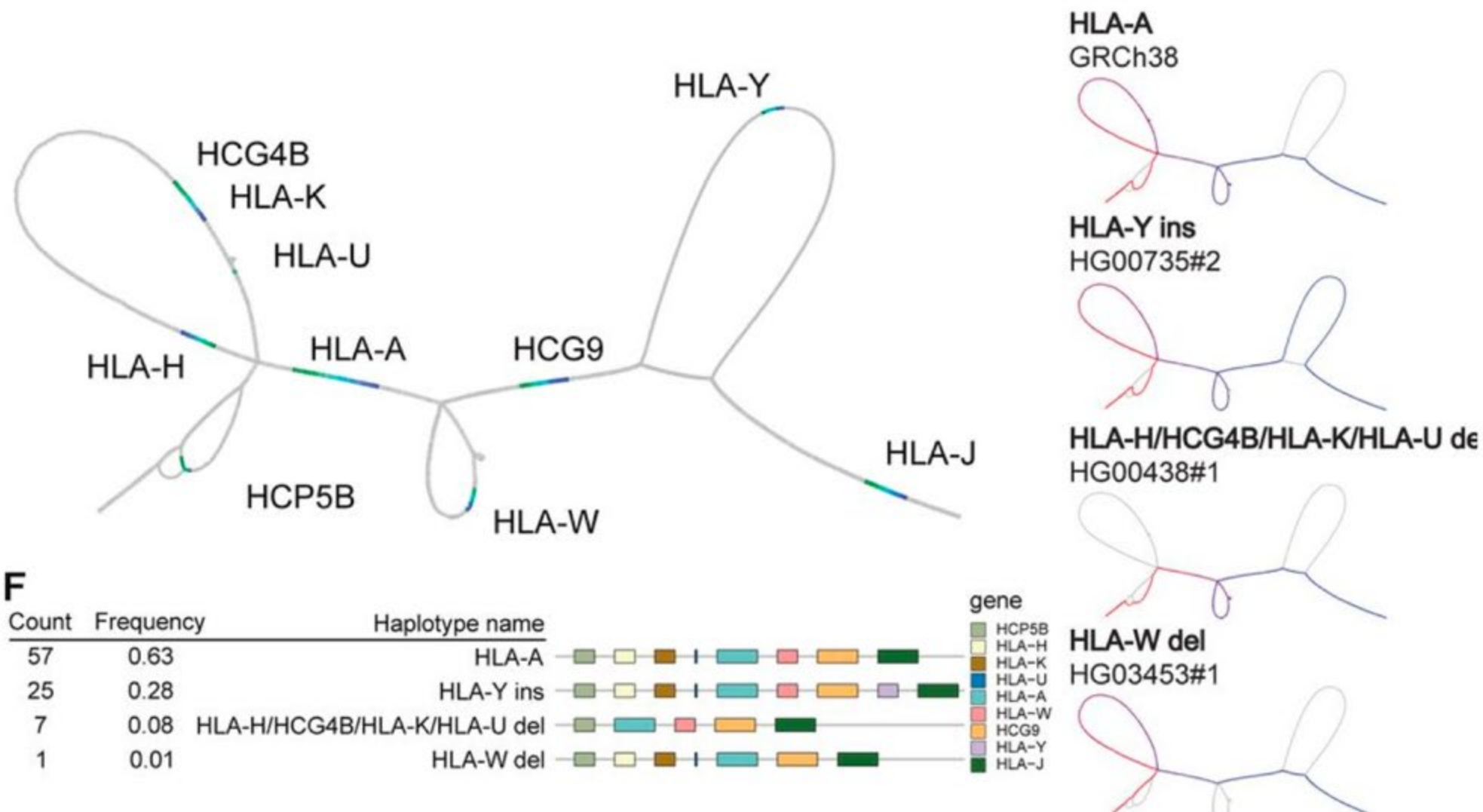
The Human Pangenome



Alignments of high-quality assemblies were performed using three different methods, pioneered by our team:

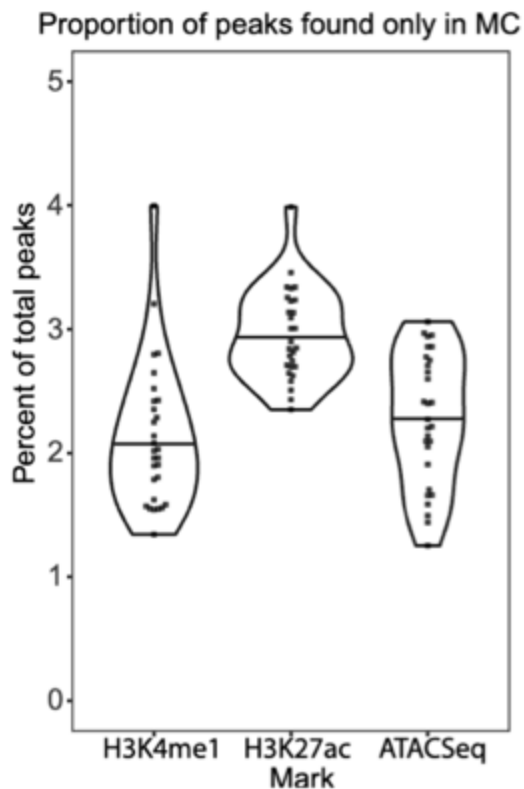
Minigraph (Li et al., 2020),
Minigraph-Cactus (MC)
PanGenome Graph Builder (PGGB).

HLA-A: Aligning Complex Pangenome Loci



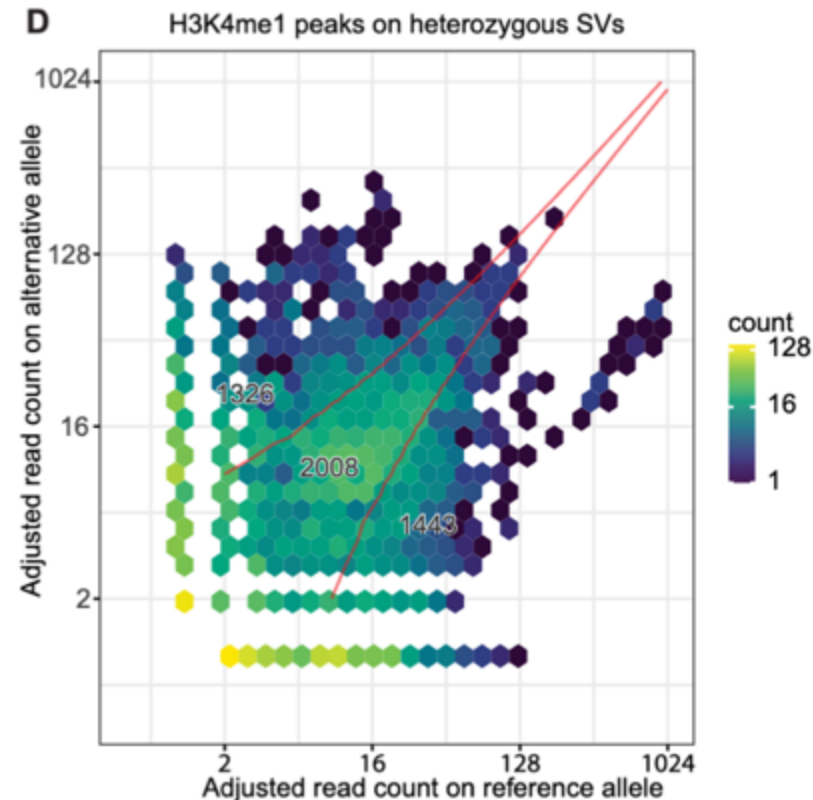
Applications

C



A pangenome approach captures annotation missing in a linear reference

D



A pangenome approach allows annotation at heterozygous structural variant sites

Remaining challenges

- Sample selection and resource development
 - Ensuring this project is truly international in scope
 - Ensuring that we common ($> 1\%$ MAF) variation is included
 - Development of usable cell lines (iPSCs, LCLs, etc) for experimental work
- Pangenome implementation
 - Will one graph representation rule them all, or do we need different graphs for different applications?
- Pangenome adoption
 - In a world where many people still use GRCh37, how do we encourage adoption of this resource?

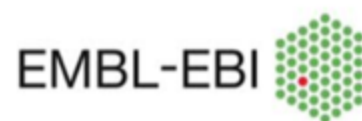
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Ryan Lorig-Roach
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Glenn Hickey Jonas Sibbesen



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Milinn Kremitzki
Haley Abel
Eddie Belter
Derek Albracht
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Chris Markovic
Tina Lindsay



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at Mount Sinai



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Nanibaa' Garrison (UCLA)
Bob Cook-Deegan (ASU)
Alice Popejoy (UC Davis)



Heng Li Haoyu Cheng
Shilpa Garg Xiaowen Feng

Valerie Schneider Chunlin Xiao
Terence Murphy Françoise Thibaud-Nissent
Paul Kitts



Company Partnerships

Alissa Resch Brittany Kerr Brittney Martinez Ellen Kelly



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HUMAN GENOME
DIVERSITY



NIH/NHGRI Funding:
U01HG010971, U41HG010972

Justin Zook



Google
DeepVariant



Dovetail
GENOMICS



PACBIO



Erich Jarvis Olivier Fedrigo Giulio Formenti Sadye Paez
Lauren Shalmiye