

Molecular Phenotypes of Null Alleles in Cells (MorPhiC) Pre-Application Webinar

Ajay Pillai, Adam Felsenfeld, Colin Fletcher, & Riley Wilson
National Human Genome Research Institute, Division of Genome Sciences
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Relevant Links

Current FOA: RFA-HG-22-019: Molecular Phenotypes of Null Alleles in Cells (MorPhiC) Phase I: Data Analysis and Validation Centers grants.nih.gov/grants/guide/rfa-files/RFA-HG-22-019.html

Email Contact: morphicprogram@nih.gov

Webinar Link: genome.gov/event-calendar/MorPHiC-data-analysis-validation-centers-pre-application-webinar

Program Webpage: genome.gov/research-funding/Funded-Programs-Projects/Molecular-Phenotypes-of-Null-Alleles-in-Cells

FAQ: genome.gov/event-calendar/MorPhiC-pre-application-webinar/FAQ

2020 NHGRI Strategic Vision: genome.gov/2020SV

Past FOAs:

- RFA-HG-21-029: Data Production Research and Development Centers (UM1 Clinical trials not allowed)
- RFA-HG-21-031: Data Resource and Administrative Coordinating Center (U24 Clinical trials not allowed)

Format

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Overall Introduction (Adam Felsenfeld) (10 min)

RFA-HG-22-019: Data Analysis and Validation Centers (Ajay Pillai) (15 min)

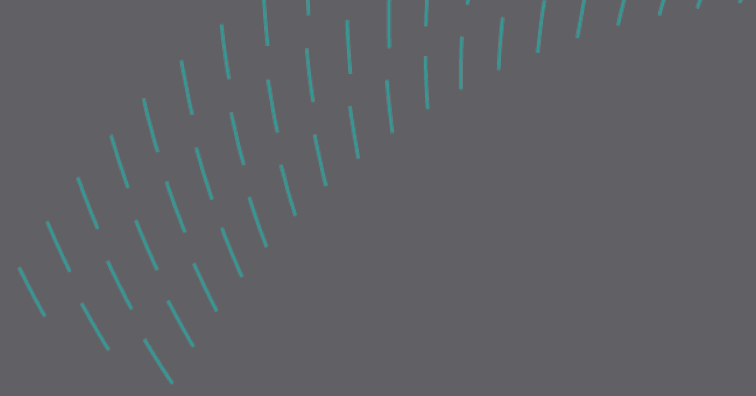
Questions: 30 mins

To Note

- **This call will be recorded and posted to the NHGRI MorPhiC Web pages.**
- **Your questions may be rendered into general FAQ's, with our answers, that will be linked to the MorPhiC Web pages.**
- **You do not have to identify yourself to ask a question.**
- **Please ask questions in the Q&A.**



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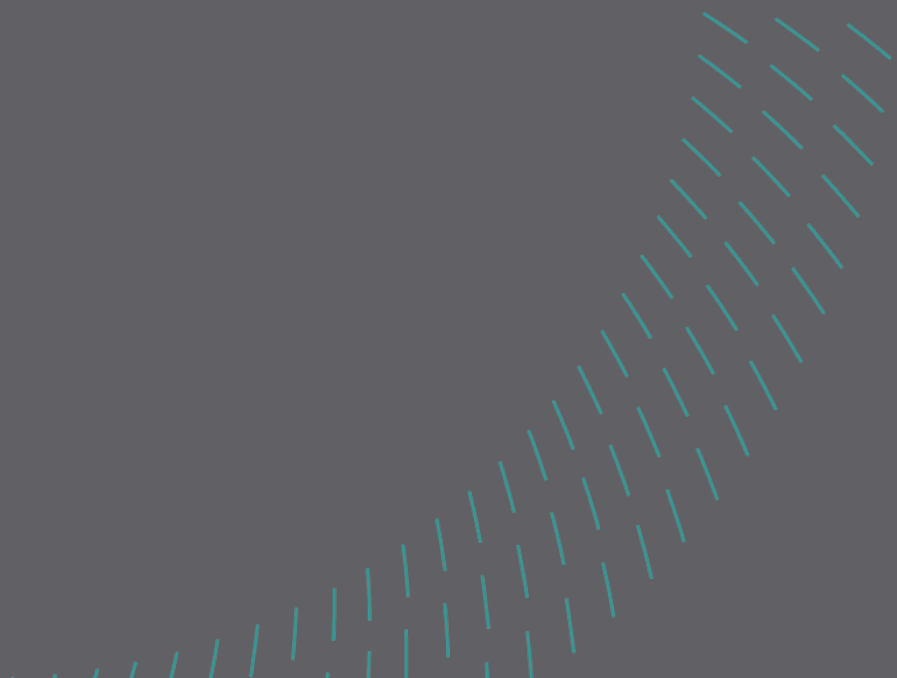
Part 1: Overview

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MorPhiC

Long-term (more than 5 years): Develop a catalog of molecular and cellular phenotypes for null alleles in human, across the genome.

IDEAL:

- Consistent (i.e., standardized, well characterized assays)
- Null (or strong l.o.f.) alleles/KOs
- Informative M&C phenotypes; in multicellular systems
- “All” genes



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Why?

- **Lack of human KOs; there are mouse and other KOs, but not molecular/cellular phenotypes.**
- **Strong alleles are useful for interpreting other alleles (incl. noncoding).**
- **Resource for insight into pathways. Complement to cis-reg initiatives (e.g., IGVF)**
- **Collection of disease models**
- **Others...**



Purpose of Phase 1

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Understand main barriers to the “long term goal”, by getting started at scale of ~1000 loci/5 years

- Criteria for selecting genes to learn lessons about doing this genome-wide
- Optimize making alleles
- Selecting cellular systems and assays for informativeness, generalizability, and scalability (tradeoffs!)
- Understand scale (costs, throughput)
- Raise and address challenges (pleiotropy, cell non-autonomy, compensation, variability)
- Understand/improve value of data (management, dissemination, quality, reproducibility, validation, variability, “use cases”, interoperability, etc.) and feed back into data production/design

Phase 1: To inform feasibility, value, and design of Phase 2

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MorPhiC Structure

Three components:

- Data production: 4 awards by Oct 2022
- Data resource and admin coordination: 1 award by Oct 2022
- **Data analysis and validation—this FOA**



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MorPhiC Overall: General advice (1)

- **Cooperative Agreements**
 - **Substantial NHGRI program management**
 - **Collaborative tasks (e.g., sample prioritization, QC and data format discussions, data flow between the three component, etc.). Read the FOA *Terms & Conditions* for how this will be managed**
 - **Flexibility to set, and adjust, milestones (needed in a complex program)**
 - **There will be a “kick-off” meeting after grants are funded to establish consortium**
- **Letters of Intent– not required, but encouraged (Oct 1).**



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MorPhiC Overall: General advice (2)

- Always read the Review Criteria section of any FOA. This is what the reviewers will use to evaluate.
- Please read the instructions to applicants for the “Research Plan” sections.
- FOA’s have a separate Resource Sharing section. *Will be considered in score.*
- Please read the section on “Review and Selection”. It lists criteria that NHGRI may apply in selecting among well-scored applications.
- Read the Budget section (minimum time commitments; consortium meetings).
- Choose letters of support judiciously.

MorPhiC Overall: Diversity & Funding

NHGRI especially encourages applications from

- **investigators from demographic groups or institutions that are generally underrepresented in genomic science**
- **new investigators**
- **experienced investigators who are new to genomic science**
- **investigators that have not previously participated in a NHGRI consortium or program**



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MorPhiC Overall: FAQ's

Please look out for these in the next month on the website genome.gov/event-calendar/MorPhiC-pre-application-webinar/FAQ

It will be updated.



Part 2: RFA HG-22-019

Data Analysis & Validation Centers



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RFA HG-22-019: Outline

Primary goals

- data variability is controllable,
- data is useful to understand basic biological processes,
- data is interpretable for undertaking future hypothesis-driven science by the community.

Projects with high potential to illuminate strengths and weaknesses

Community utilization & feedback



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RFA HG-22-019: Responsiveness

The FOA has a list of non-responsive criteria:

- Wet-lab data generation
- Do not propose to use MorPhiC data
- Do not address collaborations within Consortia
- Do not have a data sharing plan.



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RFA HG-22-019: Challenges

Integrating data: between MorPhiC labs and 'related datasets'

- Identify & Correct technical bias
- Exptl design
- Effect of KO vs background changes
- Constructing representative GRNs
- Data integration: Multiome measurements

Sufficient metadata reflecting biology & data generation.

How can small labs use the data and models for downstream experiments?



RFA HG-22-019: Consortium responsibilities

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How good/useful is the data?

Metadata and APIs and data access

Merits of common pipelines (leadership role)



RFA HG-22-019: Application & Review

Please follow instructions in the FOA about the Research Plan/Research Strategy sections.

Budget: 350K direct cost/yr (max); 5 years

Separate Data Sharing section & it is reviewed.

Specific review criteria in the FOA.

Due Date: Nov 1, 2022

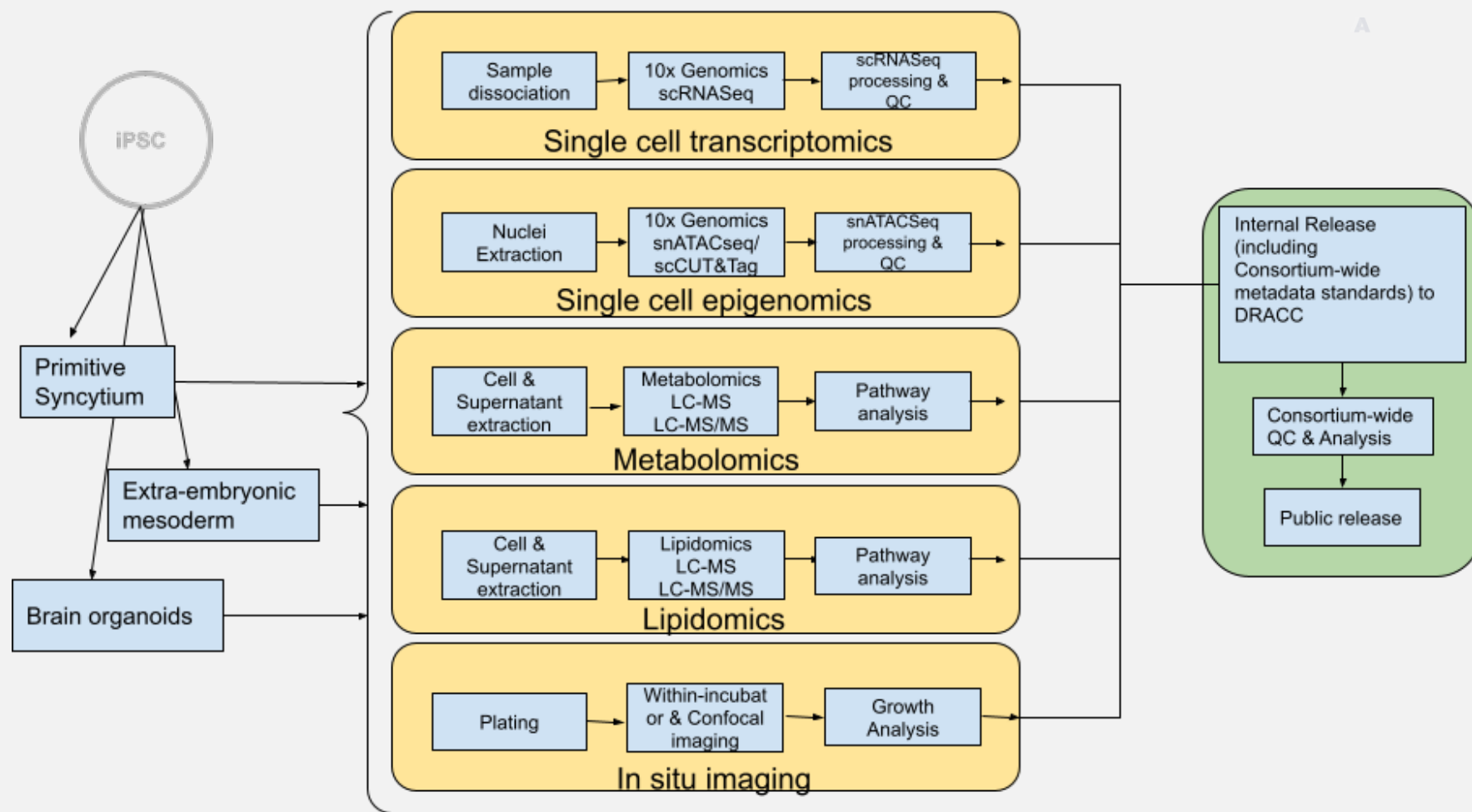


The MorPhiC: Data Production Centers (DPC)

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- Each DPC proposes to analyze
 - their own data (single modality)
 - undertake some limited integrative analysis across their datasets to discover biology

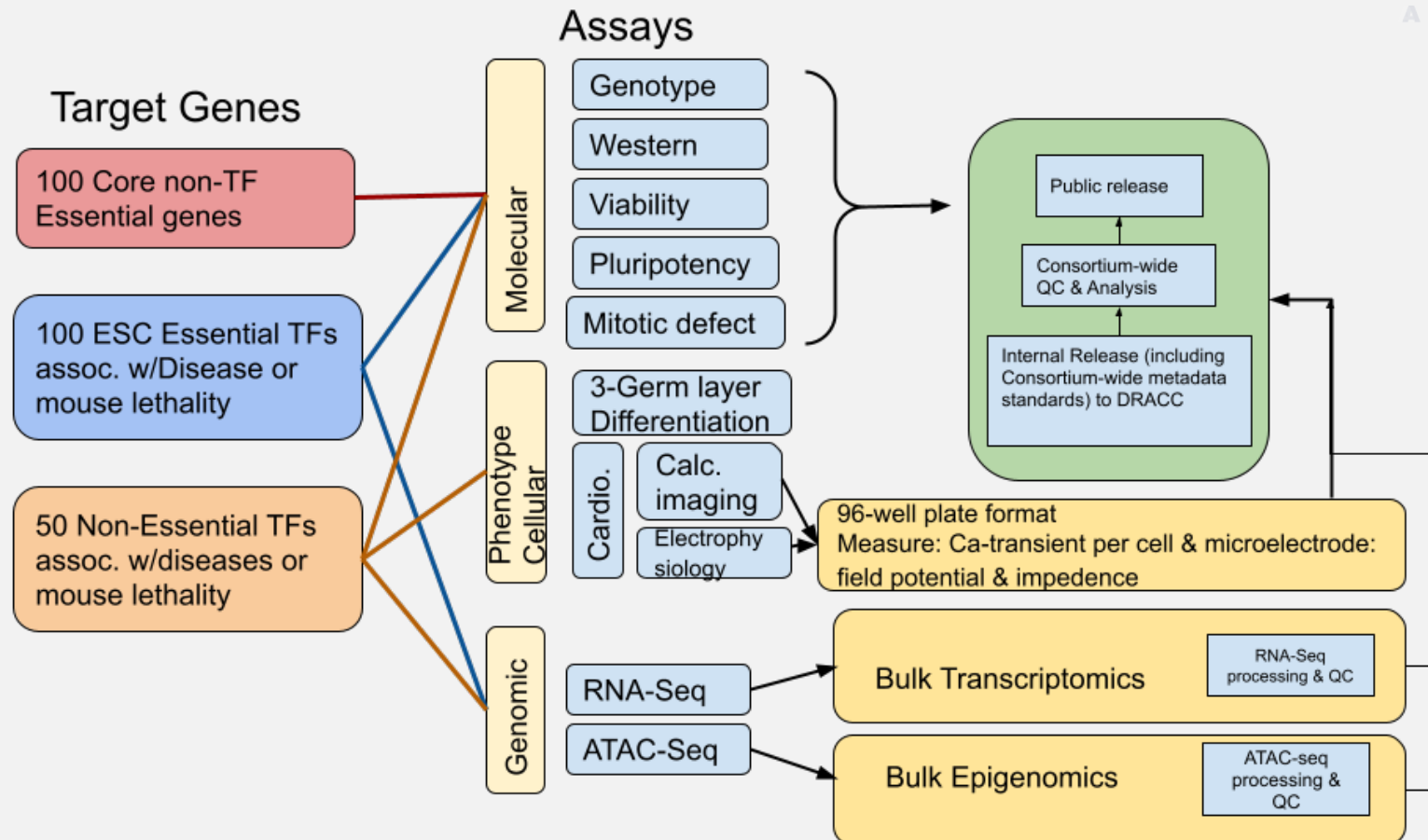
Project 1



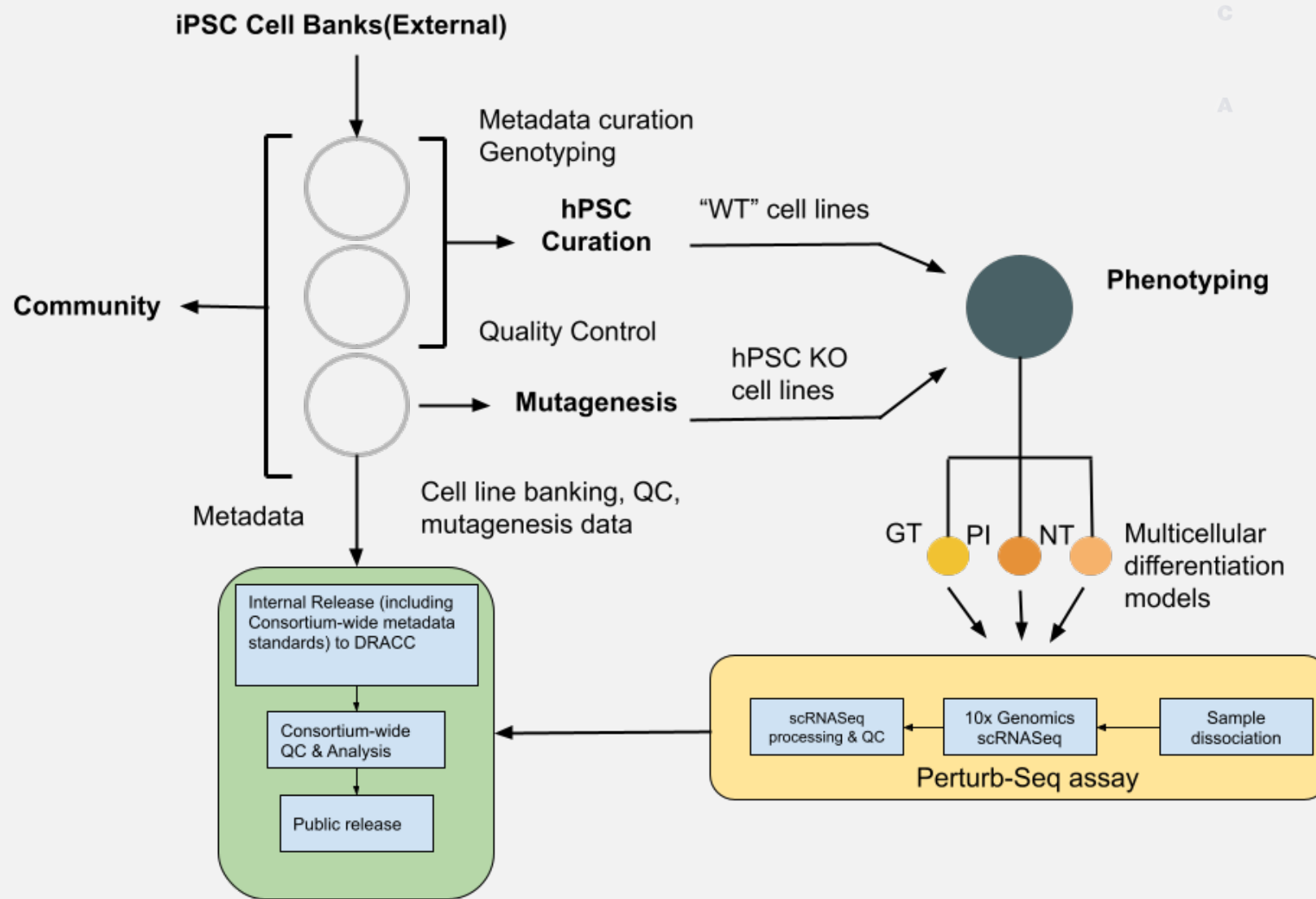
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Project 2

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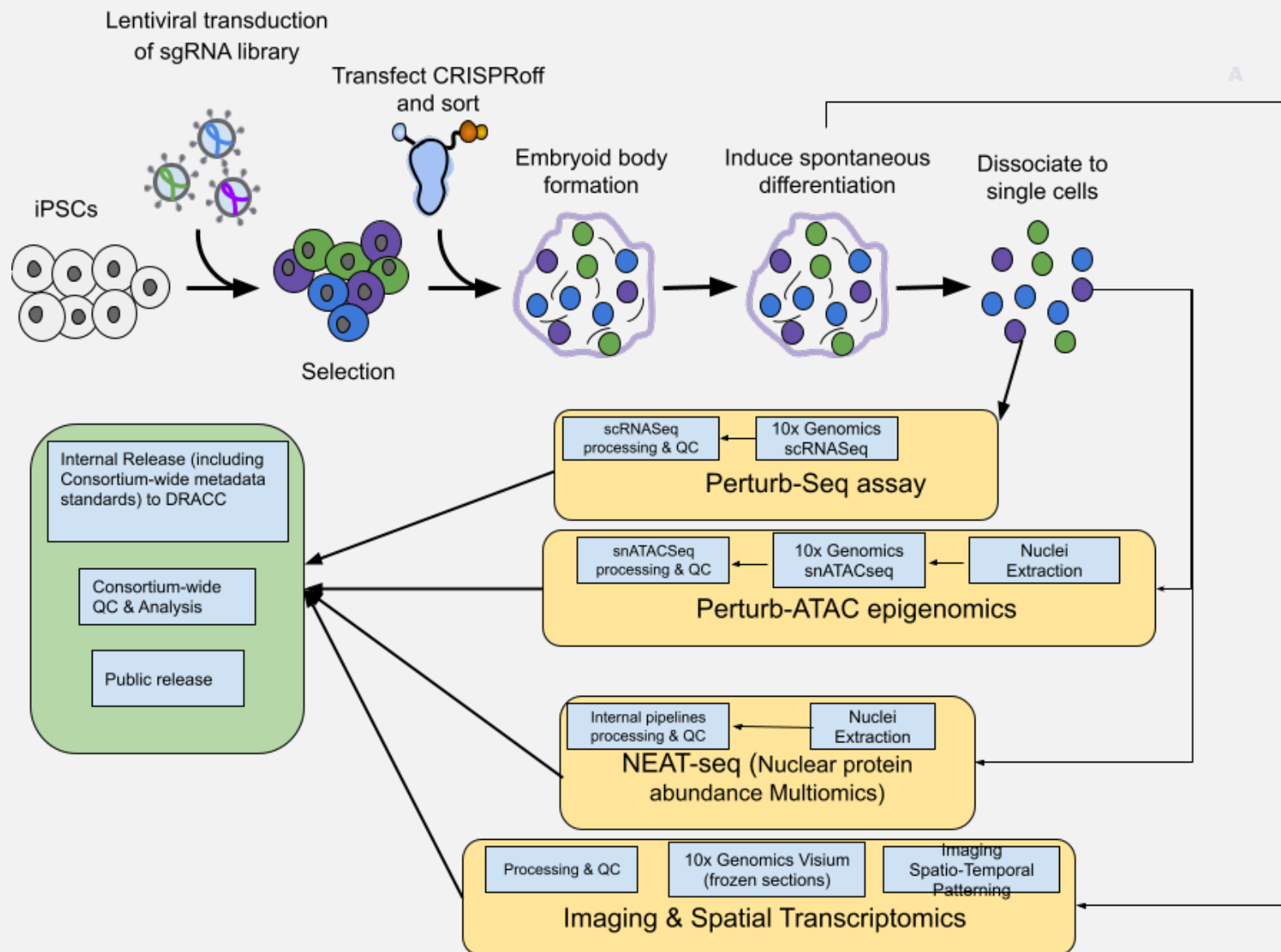


Project 3



GT: 2D gastruloid; PI: Pancreatic Islet Organoids; NT: neuron/astrocyte/microglia tri-culture

Project 4

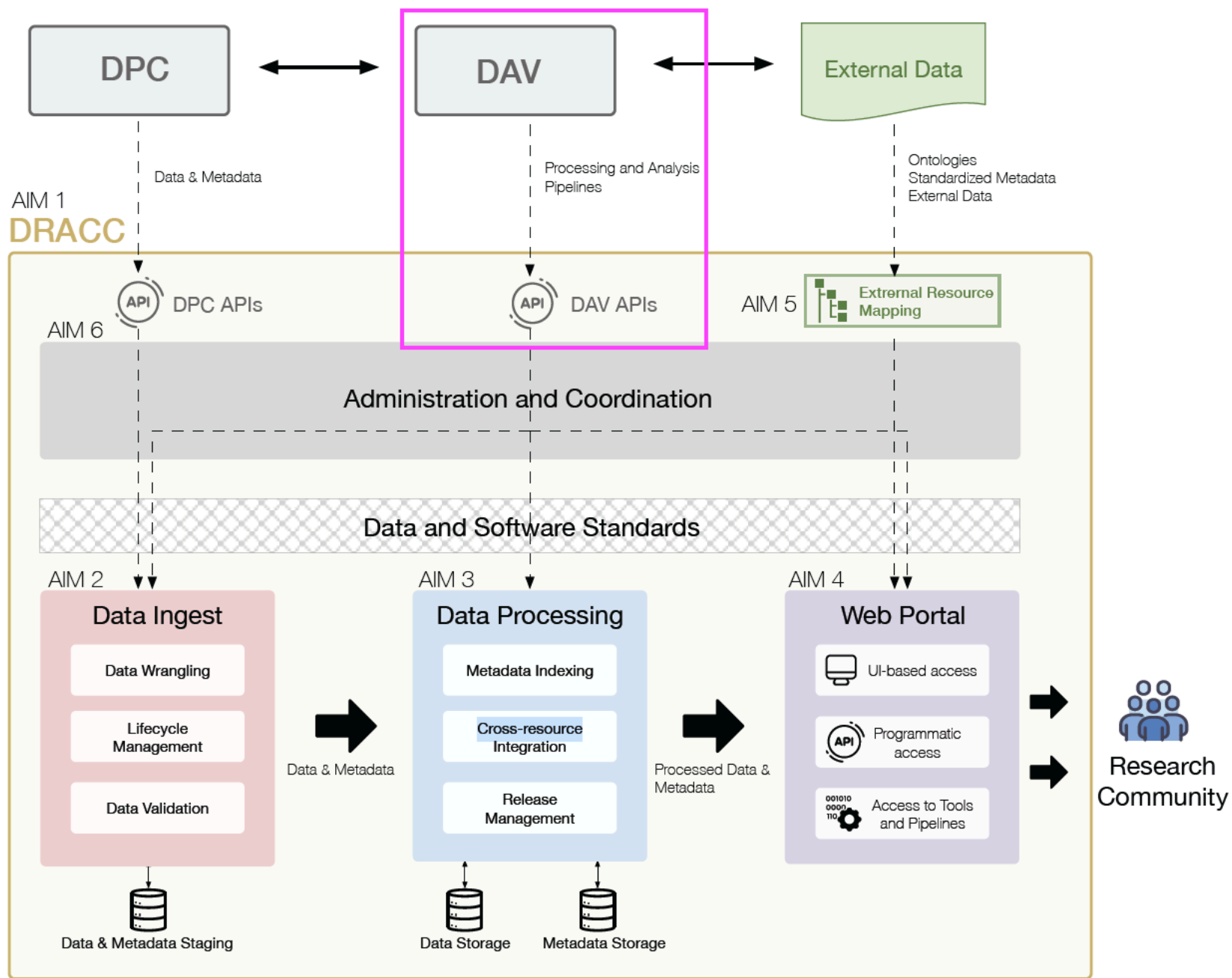


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The MorPhiC: Data Coordination Center

- The DRACC will be doing many things
 - A generic summary follows

Data Coordination





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RFA HG-22-019

Questions?



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Part 4: Tabular formatted Morphic Experimental Data

Project 1

Classes of Targeted Genes

Number of Knockouts

- ✓ 250 genes over 5 years
- ✓ 391 alleles over 5 years
- ✓ Main method: CRISPR-Cas9 KO
- ✓ gene selection criteria will focus on genes expressed at these early time points.
- ✓ Following classes will be included (and others based on Consortium discussion): developmental regulators; genes with overt transcriptional effects; others with no known transcriptional/chromatin regulation
- ✓ separate gene selection criteria for neuroectoderm lineage and extra-embryonic lineage

Classes of Targeted Genes

Cellular Models

Main workhorse cellular system for large scale data generation

- ✓ differentiated cells: KOLF2.1 (46;XY)

Other cellular systems that will be tested

- ✓ iPSC's will subsequently be differentiated into two cell lineages: (1) the extra-embryonic, trophoblast and extra-embryonic mesoderm in monolayer cultures and (2) the neuroectodermal (early embryonic lineages in organoid cultures along the neuroectoderm lineage)

Types of Assays

Main workhorse assays

- ✓ sc-transcriptomics at 4 different time points
- ✓ sn-epigenetics
- ✓ detailed high-resolution bulk proteomics & lipidomics (75 genes over 5 years): LC-MS and LC-MS/MS
- ✓ In biological triplicate (replication)

Other assays that will be used in a targeted manner

- ✓ differentiation schemes will be temporally imaged to capture phenotypes & morphology
- ✓ Stable isotope labeled spike-in standards will be used NIST (SRM 1950) & QStd in all metabolomics and lipid assays

Other General Scientific Questions to be Explored

- ✓ Replicate a number of KOs (18 genes with robust cellular phenotypes) in three additional iPSC lines to address impacted by sex and/or genetic background (1 European female, 2 lines of non-European ancestry male & female)
- ✓ Knockout clones will also be reverted to wild-type by a second round of CRISPR-based editing to investigate: reversion of phenotype; evidence for cellular adaptation responses; contribution of genetic drift?
- ✓ generate protein-tagged 'degron' alleles for the same set of five genes to test an orthogonal method for conditional, reversible gene knockouts

- ✓ phenotyping efforts, in Phase I of MorPhiC, will focus on relatively early events in development
- ✓ compare KO strategies—editing biallelic premature termination codon, exon and gene deletion alleles for 24 genes assay based on transcription changes to identify reliable strategy for null allele generation

Project 2

Classes of Targeted Genes

Cellular Models

Types of Assays

Other General Scientific Questions to be Explored

Number of Knockouts

✓ 250 genes in iPSC
 ✓ Main method: conditional null phenotype by using the Auxin inducible degron technology (AID)

Main **workhorse cellular system for large scale data generation**

✓ Female hiPSC line
 ✓ After selection of single cell clones (after transfection) a number of QC steps will be undertaken

Main **workhorse assays**

✓ bulk RNA-Seq at 0, 6 & 48 hrs after null allele induction for 100 TFs
 ✓ bulk ATAC-Seq at 0, 6, & 48 hrs after null allele induction for 100 TFs
 ✓ IncuCyte live cell imaging platform to study viability, proliferation & mitotic defects after induction of null phenotype with auxin treatment

✓ Fitness & Stemness phenotype after null allele using FACS
 ✓ Differentiation propensity of 250 alleles into 3 germ layers. Mesoderm—cardiomyocytes, endoderm—hepatocyte-like cells, ectoderm—sensory neurons in a 10-day timeline.
 ✓ Cardiomyocytes: mature d30 cardiomyocytes will be studied for a limited number of KO alleles using (1) high-throughput Ca-transient measurement, and (2) microelectrode array measurement of field potential duration, beat rate etc.

Classes of Targeted Genes

✓ 200 essential genes
 ✓ 50 non-essential TFs

Other cellular systems that will be tested

Other assays that will be used in a targeted manner

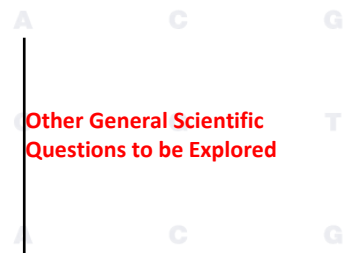
✓ Genotype
 ✓ Western

✓ Compare ~50 clonal CRISPR KO with AID-mediated protein depletion (RNA and ATAC)
 ✓ AID-knock-in strategy allows controlling the timing and the level of the target protein degradation and it is reversible (multiple downstream questions can be addressed).

Classes of Targeted Genes

Cellular Models

Types of Assays



Other General Scientific Questions to be Explored

Number of Knockouts

- ✓ Generate clonal KO lines for ~250 genes
- ✓ generate "population KO" for 3-5 selected genes using all 96 curated hPSC lines
- ✓ Method: CRISPR-Cas9 based HDR

Main workhorse cellular system for large scale data generation

- ✓ 96 curated (existing) hPSC lines; equal male, female; emphasize diverse ancestry; healthy
- ✓ array individual lines in 96-well format, and then pool the lines
- ✓ clonal KO lines for ~250 genes, select 3-5 diverse and representative hPSC lines, including RUES2
- ✓ Differentiation (2D gastruloid, neuron-glia tri-culture, and islet-like organoids) starting from 96 curated hPSC lines in a pooled format

Main workhorse assays

- ✓ scRNA-seq (Perturb-Seq; 10x)
- ✓ Perturb-Seq on pooled clonal KO 5-10 diverse hPSC lines
- ✓ (Custom scripts) for Perturb-Seq demultiplexing and assignment of reads and sgRNA.

- ✓ Phenotyping in diverse genetic backgrounds
- ✓ Variability of differentiation behavior (including proliferation related issues in complex cell systems)
- ✓ Some genes (due to pleiotropy) may need KO after differentiation

Classes of Targeted Genes

- ✓ 161 genes required for normal embryonic development in mice (distribution of genes with various mouse phenotypes)
- ✓ 50 genes linked to T2D and Alzheimer's disease based on human genetic studies
- ✓ Set of +ve & -ve control genes to QC phenotype assays

Other cellular systems that will be tested

- ✓ Primary human islets

Other assays that will be used in a targeted manner

- ✓ Improvements in power calculations to determine # of cells screened/sample (extensions into pooled screens).

	Classes of Targeted Genes		Cellular Models		Types of Assays		Other General Scientific Questions to be Explored
Project 4	<p>Number of Knockouts</p> <ul style="list-style-type: none"> ✓ Method: Use CRISPRoff (directly target mRNA and protein levels) ✓ 250 genes 	<p>Main workhorse for large scale data generation</p>	<ul style="list-style-type: none"> ✓ iPSC cells ✓ Embryoid body (all 3 germ lineages at equal representation) ✓ iPSC from 2 Asian, 2 Hispanic, 2 African American, 4 European (1 male and 1 female) ages: 24-72. 	<p>Main workhorse assays</p>	<ul style="list-style-type: none"> ✓ Perturb-Seq (pooled screening) ✓ Perturb-ATAC (pooled screening) ✓ (Custom scripts) for Perturb-Seq demultiplexing and assignment of reads and sgRNA. 	<ul style="list-style-type: none"> ✓ Demonstrate that CRISPRon can functionally reverse CRISPRoff null allele phenotypes in a polyclonal population of iPSCs and in embryoid bodies. 	
	<p>Classes of Targeted Genes</p> <ul style="list-style-type: none"> ✓ Priority: transcription factors and epigenetic regulators 	<p>Other cellular systems that will be tested</p>	<ul style="list-style-type: none"> ✓ Each CRISPRoff will be followed by CRISPRon for validation studies. 	<p>Other assays that will be used in a targeted manner</p>	<ul style="list-style-type: none"> ✓ NEAT-seq (simultaneously measure RNA, Chromatin, intracellular proteins—pooled screening) ✓ Non-pooled EB generation with null alleles in each cell (20-30 prioritized genes) followed by serial confocal imaging every 2hrs from 24-72hrs. ✓ Followed by freezing/sectioning for 10x Visium (spatial profiling). 	<ul style="list-style-type: none"> ✓ Explore genetic compensation ✓ Explore the influence of paralogs on 'null phenotypes after KO' 	



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