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Capturing RNA Sequence and Transcript Diversity, From Technology Innovation to Clinical Application

Jennifer Strasburger

Program Director,
Division of Genome Sciences

Brenton Graveley

UConn Health Department of Genetics & Genome
Sciences, Institute for Systems Genomics
Executive Committee Member

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National Human Genome
Research Institute

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NHGRI 2020 Strategic Vision

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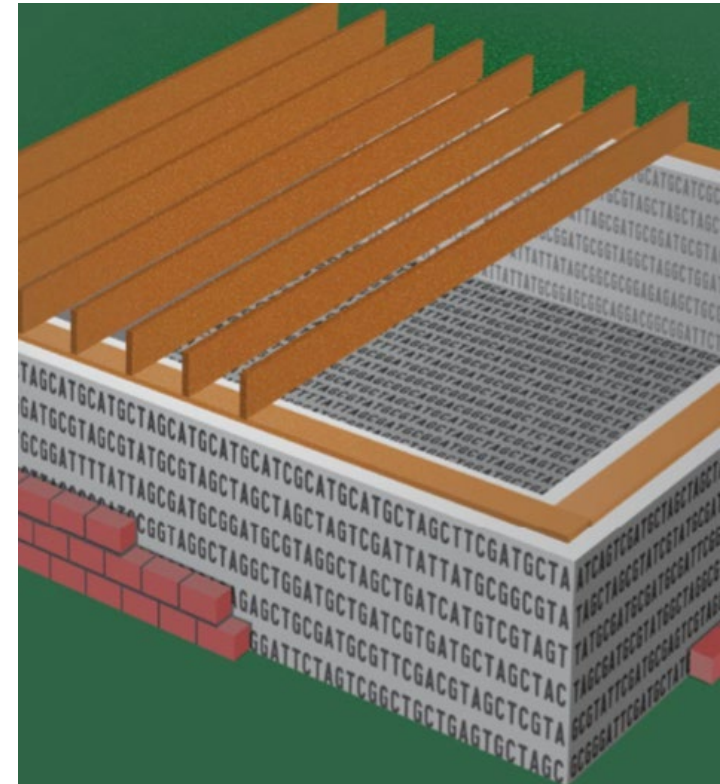
Green, E.D., Gunter, C., Biesecker, L.G. *et al.* Strategic vision for improving human health at The Forefront of Genomics. *Nature* 586, 683–692 (2020).

Building a Robust Foundation

Genome Structure & Function:

*“Technologies for generating DNA sequence and other datatypes (for example, transcriptomic data, epigenetic data, and functional readouts of DNA sequences) need to be enabled at orders-of-magnitude **lower costs**, at **single-cell resolution**, at **distinct spatial locations** within tissues, and **longitudinally over time**.”*

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NHGRI's RNA Goals

- Direct RNA sequencing
- Modified RNA bases
- Transcript diversity
- Synthetic RNA with modifications
- Multi-omic approaches to disease and risk
- Functional genomics

A call from the RNA community

- Technologies needed to sequence full-length RNA directly
- Address 140+ chemical RNA modifications
- Defects related to modifications account for 100+ human diseases
- Informatics to detect and identify all modifications & splice variants

Nat Genet 53, 1113–1116 (2021)

 Check for updates

[comment](#)

A call for direct sequencing of full-length RNAs to identify all modifications

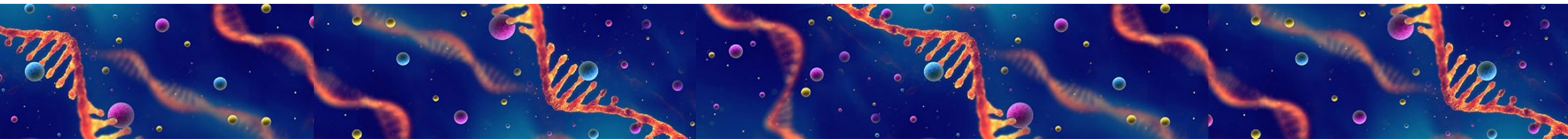
For most organisms, DNA sequences are available, but the complete RNA sequences are not. Here, we call for technologies to sequence full-length RNAs with all their modifications.

Juan D. Alfonzo, Jessica A. Brown, Peter H. Byers, Vivian G. Cheung, Richard J. Maraia and Robert L. Ross

Plus 50 signatories

Capturing RNA Sequence and Transcript Diversity, From Technology Innovation to Clinical Application

- Virtual workshop hosted by NHGRI & NIEHS
- May 24-26, 2022



www.niehs.nih.gov/news/events/pastmtg/2022/rnaworkshop2022/index.cfm

The Planning Team

Thanks to the many others who participated and helped with planning efforts!

Executive Committee:

- Vivian Cheung
- Pete Dedon
- Brenton Graveley
- Blanton Tolbert

Avanti Corporation:

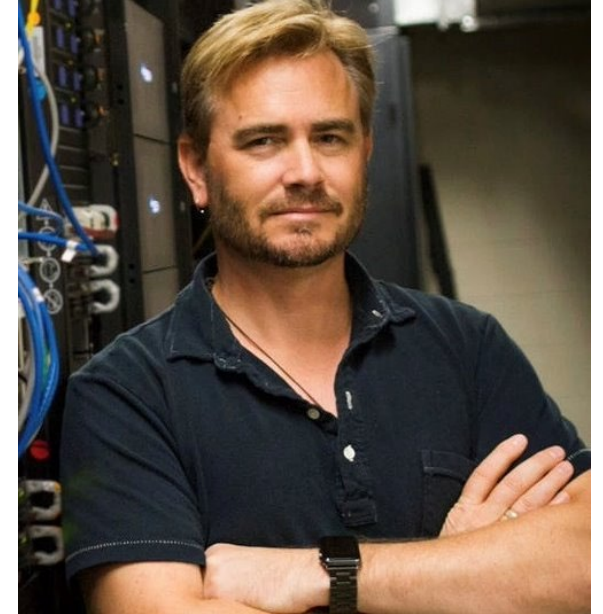
- Justin Crane
- Manashree Malpe
- Devan Smith-Brown

NHGRI & NIEHS Staff:

- Katie Bardsley
- Michelle Heacock
- Kim McAllister
- Alicia Ramsaran
- Dan Shaughnessy
- Mike Smith
- Jennifer Strasburger
- Fred Tyson
- Leroy Worth

Brenton Graveley

- UConn Health
- Chair of the Department of Genetics and Genome Sciences
- Associate Director of the Institute for Systems Genomics
- Workshop Executive Committee Member
- Former NACHGR Member



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State of the Field

- Most RNAs involved in active gene expression are **very long** (>1kb), **fragile**, and **low abundance**
- Functionally **relevant information** within RNA is **not captured** by its encoded sequence
- Most RNA sequencing is done with **short reads, bulk samples, and inefficient enzymes**
- Impossible to **detect RNA modifications transcriptome-wide**, with known stoichiometry
- **Machine learning approaches and methods to interpret and manage the data** are lacking

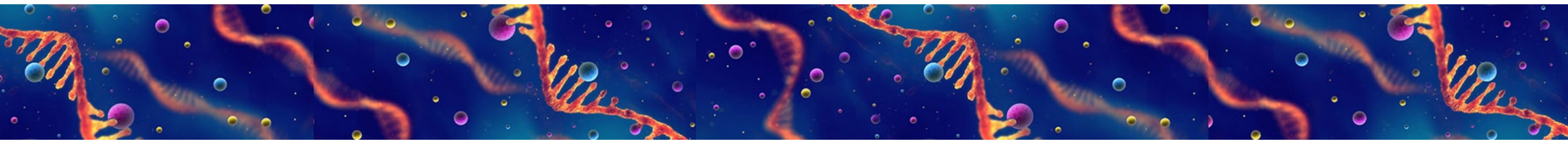
Workshop Objectives

- Identify **technologies** needed to **comprehensively characterize RNA**
- Determine **infrastructure, bioinformatics, and other resource needs**
- Consider the steps needed to **facilitate rapid adoption** of these technologies
- Identify how to best incorporate **public outreach and workforce development**

Process

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- Participants discussed challenges and opportunities throughout the workshop.
- Key take-aways were synthesized by the planning team, reviewed by the Executive Committee, and appear in the Executive Summary on genome.gov.



www.genome.gov/event-calendar/97th-Meeting-of-National-Advisory-Council-for-Human-Genome-Research

Priority Areas

- Diversity, Inclusion, & Training
- Technology Development
- Biological Standards & Centralized Resources
- Bioinformatics, Computation, & Data Science
- A Path to Clinical Implementation
- Environmental Exposure and Stress

Diversity, Inclusion, & Interdisciplinary Training

- **Additional funding** across career stages to **promote diversity**.
- **Outreach components** in large NIH-funded consortium and center projects.
- Support of **undergraduate and graduate-level training**.
- **Public Outreach**.
- **Interdisciplinary virtual and in-person training** in both wet and dry lab methods available and accessible to all professional levels.

Technology Development

- **New technologies** for full-length, direct RNA analysis.
- **New MS capabilities** as well as improvements of existing instrumentation.
- Focused efforts to harness the **full capability of enzymes** for RNA analysis.
- Creating a **library of long, synthetic RNAs** with specific modifications.
- Detecting, locating, and analyzing **interactions between DNA, RNA, proteins, and small molecules.**

Biological Standards & Centralized Resources



- “**Gold standard**” reference set of synthetic and cellular RNA splice isoforms with known chemical modifications.
- **Centralized resource** with banked tissues, cell lines, and exemplar viral strains.
- Consortium or coordination center-directed efforts to **stimulate collaborations** for both wet and dry lab advances in direct RNA analysis
- Centralized resource for **direct RNA sequencing**.

Bioinformatics Tools, Computational Resources, & Data Standards

- **RNA database** of different RNA types, includes normal and diseased tissue.
- **Standardized** nomenclature, file formats, pipelines, and software.
- **Machine learning/AI-based tools.**
- **RNA training sets.**
- **RNA secondary structure prediction** that considers chemical modifications.

Environmental Exposure and Stress

- Tools/reagents to study stress-, **environmental exposure-induced modification** in normal and disease states.
- Tools/reagents to map **temporal and spatial dynamics** of RNA modifications in different cellular and tissue contexts.
- Methods to interrogate impact of stress on RNA relative to **signaling, RNA-protein interactions, and trafficking.**

A Path to Clinical Implementation

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- Small molecule libraries **targeting RNA structures**.
- RNA-modifying **enzymes** as potential **drug targets**.
- Improved **delivery technologies** to target specific tissues and cell types.
- New **therapeutic** development pipelines.
- High-grade **oligonucleotide** discovery and synthesis;
GLP manufacturing for pre-clinical and clinical studies.

Conclusions

Identified opportunities in:

- Direct RNA sequencing with modification identification.
- The broader arena of RNA structure, RBPs, and biology.
- RNA as therapeutic and drug target.
- Developing the capabilities to obtain a truly comprehensive view of the transcriptome.



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Discussion



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